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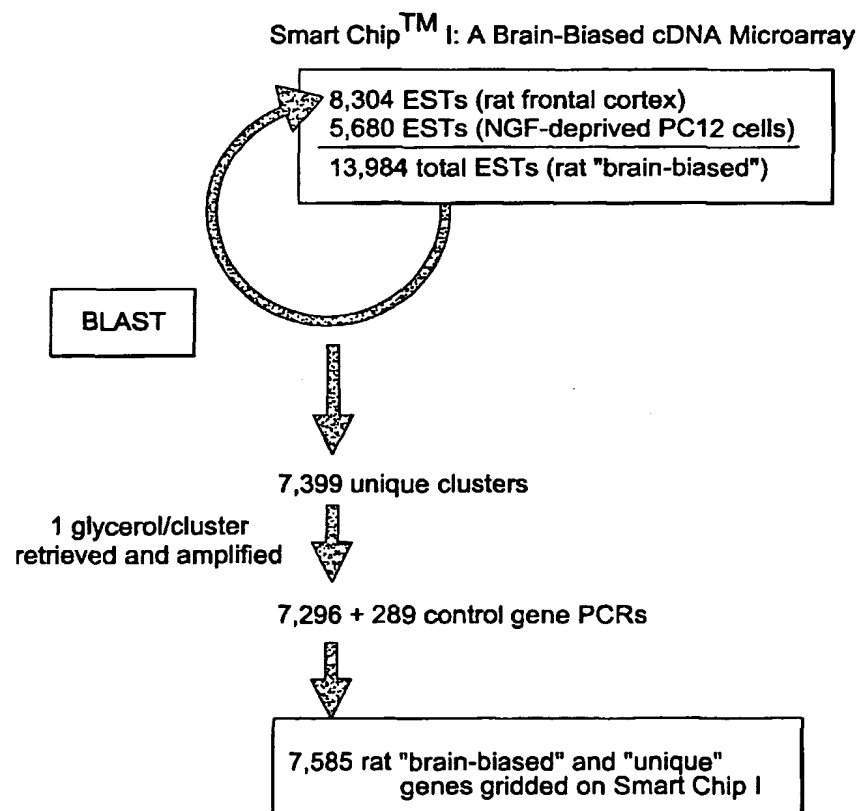
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(54) Title: NUCLEIC ACID MOLECULES DERIVED FROM RAT BRAIN AND PROGRAMMED CELL DEATH MODELS



(57) Abstract: The invention is directed to human homologs of nucleic acid molecules derived from rat brain and programmed cell death expression libraries. These molecules can constitute microarrays of expressed sequences useful for analyzing gene expression in various biological contexts, including development, differentiation, and disease, both *in vivo* and *in vitro*. The nucleic acid molecules are useful for diagnosis, treatment, and drug discovery. The nucleic acid molecules are useful for creating microarrays for transcriptional profiling. The invention further provides peptides encoded by the nucleic acid molecules, useful for methods of diagnosis, treatment, and drug discovery. The invention specifically relates to nucleic acid molecules involved in programmed cell death.

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NUCLEIC ACID MOLECULES DERIVED FROM RAT BRAIN AND PROGRAMMED CELL DEATH MODELS

FIELD OF THE INVENTION

The invention relates to nucleic acid molecules derived from rat brain and programmed cell death expression libraries. Also provided are vectors, host cells, and
5 methods for making and using the novel molecules of the invention.

BACKGROUND OF THE INVENTION

A great deal of effort has been expended by the modern scientific research community to identify and sequence genes, particularly human genes. The
10 identification of genes and knowledge of their nucleic acid sequences pave the way for many scientific and commercial advancements, both in research applications and in diagnostic and therapeutic applications. For example, advances in gene identification and sequencing allow the production of the products encoded by these genes, such as by recombinant and synthetic means. Furthermore, identification of
15 genes and the products they encode provide important information about the mechanism of disease and can provide new diagnostic tests and therapeutic treatments for the diagnosis and treatment of disease. Thus, identification and sequencing of genes provide valuable information and compositions for use in the biotechnology and pharmaceutical industries.

20 In multicellular organisms, homeostasis is maintained by balancing the rate of cell proliferation against the rate of cell death. Cell proliferation is influenced by numerous growth factors and the expression of proto-oncogenes, which typically encourage progression through the cell cycle. In contrast, numerous events, including the expression of tumor suppressor genes, can lead to an arrest of cellular
25 proliferation.

In differentiated cells, a particular type of cell death called apoptosis occurs when an internal suicide program is activated. This program can be initiated by a variety of external signals as well as signals that are generated within the cell in response to, for example, genetic damage. Dying cells are eliminated by phagocytes,
30 without an inflammatory response.

Programmed cell death (PCD) is a highly regulated process (Wilson (1998) *Biochem. Cell. Biol.* 76:573-582). The death signal is then transduced through various signaling pathways that converge on caspase-mediated degradative cascades resulting in the activation of late effectors of morphological and physiological aspects of apoptosis, including DNA fragmentation and cytoplasmic condensation. In addition, regulation of programmed cell death may be integrated with regulation of energy, redox- and ion homeostasis in the mitochondria (reviewed by Kroemer (1998) *Cell Death and Differentiation* 5:547), and/or cell-cycle control in the nucleus and cytoplasm (reviewed by Choisy-Rossi and Yonish-Rouach (1998) *Cell Death and Differentiation* 5:129-131; Dang (1999) *Molecular and Cellular Biology* 19:1-11; and Kasten and Giordano (1998) *Cell Death and Differentiation* 5:132-140). Many mammalian genes regulating apoptosis have been identified as homologs of genes originally identified genetically in *Caenorhabditis elegans* or *Drosophila melanogaster*, or as human oncogenes. Other programmed cell death genes have been found by domain homology to known motifs, such as death domains, that mediate protein-protein interactions within the programmed cell death pathway.

The mechanisms that mediate apoptosis include, but are not limited to, the activation of endogenous proteases, loss of mitochondrial function, and structural changes such as disruption of the cytoskeleton, cell shrinkage, membrane blebbing, and nuclear condensation due to degradation of DNA. The various signals that trigger apoptosis may bring about these events by converging on a common cell death pathway that is regulated by the expression of genes that are highly conserved.

Caspases (cysteine proteases having specificity for aspartate at the substrate cleavage site) are central to the apoptotic program, are. These proteases are responsible for degradation of cellular proteins that lead to the morphological changes seen in cells undergoing apoptosis. One of the human caspases was previously known as the interleukin-1 β (IL-1 β) converting enzyme (ICE), a cysteine protease responsible for the processing of pro-IL-1 β to the active cytokine. Overexpression of ICE in Rat-1 fibroblasts induces apoptosis (Miura *et al.* (1993) *Cell* 75:653).

Many caspases and proteins that interact with caspases possess domains of about 60 amino acids called a caspase recruitment domain (CARD). Apoptotic proteins may bind to each other via their CARDS. Different subtypes of CARDS may

confer binding specificity, regulating the activity of various caspases. (Hofmann *et al.* (1997) *TIBS* 22:155).

The functional significance of CARDs have been demonstrated in two recent publications. Duan *et al.* (1997) *Nature* 385:86 showed that deleting the CARD at the N-terminus of RAIDD, a newly identified protein involved in apoptosis, abolished the ability of RAIDD to bind to caspases. In addition, Li *et al.* (1997) *Cell* 91:479 showed that the N-terminal 97 amino acids of apoptotic protease activating factor-1 (Apaf-1) was sufficient to confer caspase-9-binding ability.

Thus, programmed cell death (apoptosis) is a normal physiological activity necessary to proper and differentiation in all vertebrates. Defects in apoptosis programs result in disorders including, but not limited to, neurodegenerative disorders, cancer, immunodeficiency, heart disease and autoimmune diseases (Thompson *et al.* (1995) *Science* 267:1456).

In vertebrate species, neuronal programmed cell death mechanisms have been associated with a variety of developmental roles, including the removal of neuronal precursors which fail to establish appropriate synaptic connections (Oppenheim *et al.* (1991) *Annual Rev. Neuroscience* 14:453-501), the quantitative matching of pre- and post-synaptic population sizes (Herrup *et al.* (1987) *J. Neurosci.* 7:829-836), and sculpting of neuronal circuits, both during development and in the adult (Bottjer *et al.* (1992) *J. Neurobiol.* 23:1172-1191).

Inappropriate apoptosis has been suggested to be involved in neuronal loss in various neurodegenerative diseases such as Alzheimer's disease (Loo *et al.* (1993) *Proc. Natl. Acad. Sci.* 90:7951-7955), Huntington's disease (Portera-Cailliau *et al.* (1995) *J. Neurosci.* 15:3775-3787), amyotrophic lateral sclerosis (Rabizadeh *et al.* (1995) *Proc. Natl. Acad. Sci.* 92:3024-3028), and spinal muscular atrophy (Roy *et al.* (1995) *Cell* 80:167-178).

In addition, improper expression of genes involved in apoptosis has been implicated in carcinogenesis. Thus, it has been shown that several "oncogenes" are in fact involved in apoptosis, such as in the Bcl family.

Accordingly, genes involved in apoptosis are important targets for therapeutic intervention. It is important, therefore, to identify novel genes involved in apoptosis or to discover whether known genes function in this process.

Nucleic acid probes have long been used to detect complementary nucleic acid sequences in a nucleic acid of interest (the "target" nucleic acid). In some assay formats, the nucleic acid is tethered, i.e., by covalent attachment, to a solid support. Arrays of nucleic acid sequences immobilized on solid supports have been used to
5 detect specific nucleic acid sequences in a target nucleic acid. See, e.g., PCT patent publication Nos. WO 89/10977 and 89/11548. Others have proposed the use of large numbers of nucleic acid sequences to provide the complete nucleic acid sequence of a target nucleic with methods for using arrays of immobilized nucleic acid sequences for this purpose. See U.S. Pat. Nos. 5,202,231 and 5,002,867 and PCT patent
10 publication No. WO 93/17126.

The development of specific microarray technology has provided methods for making very large arrays of nucleic acid sequences in very small physical arrays. See U.S. Pat. No. 5,143,854 and PCT patent publication Nos. WO 90/15070 and
92/10092, each of which is incorporated herein by reference. U.S. patent application
15 No. 082,937, filed Jun. 25, 1993, describes methods for making arrays of sequences that can be used to provide the complete sequence of a target nucleic acid and to detect the presence of a nucleic acid containing a specific nucleotide sequence. Thus, microfabricated arrays of large numbers of nucleic acid sequences, called "DNA
chips" offer great promise for a wide variety of applications.

20

SUMMARY OF THE INVENTION

The present invention is based on the identification of novel nucleic acid molecules derived from rat brain and programmed cell death cDNA libraries.

Thus, in one aspect, the invention provides an isolated nucleic acid molecule
25 that comprises a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

The invention also provides an isolated fragment or portion of any of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complement of the
30 sequences shown in SEQ ID NOS: 1-6, 8, and 10. In some embodiments, the fragment is useful as a probe or primer, and/or is at least 15, at least 18, or at least 20, 22, 25, 30, 35, 50, 100, 200 or more nucleotides in length.

In another embodiment, the invention provides an isolated nucleic acid molecule that comprises a nucleotide sequence that is at least about 60% identical, about 65% identical, about 70% identical, about 80% identical, about 90% identical, about 95% identical, about 96% identical, about 97% identical, about 98% identical, or about 99% or more identical to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

In another embodiment, the invention provides an isolated nucleic acid molecule that hybridizes under highly stringent conditions to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

The invention further provides nucleic acid vectors comprising the nucleic acid molecules described above. In one embodiment, the nucleic acid molecules of the invention are operatively linked to at least one expression control element.

The invention further includes host cells, such as bacterial cells, fungal cells, plant cells, insect cells and mammalian cells, comprising the nucleic acid vectors described above.

In another aspect, the invention provides isolated gene products, proteins and polypeptides encoded by nucleic acid molecules of the invention.

The invention further provides antibodies, including monoclonal antibodies, or antigen-binding fragments thereof, which selectively bind to the isolated proteins and polypeptides of the invention.

The invention also provides methods for preparing proteins and polypeptides encoded by isolated nucleic acid molecules described herein by culturing a host cell containing a vector molecule of the invention.

Additionally, the invention provides a method for assaying for the presence of a nucleic acid sequence, protein or polypeptide of the present invention, in a biological sample, e.g., in a tissue sample, by contacting said sample with an agent (e.g., an antibody or a nucleic acid molecule) suitable for specific detection of the nucleic acid sequence, protein or polypeptide.

The invention also provides a kit comprising a nucleic acid probe which hybridizes to a nucleotide sequence of claim 1 and instructions for use, and a kit comprising an agent which binds to a polypeptide of claim 10 and instructions for use.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 illustrates the construction of the "Smart Chip™ I". cDNAs were cloned from rat frontal cortex and from differentiated PC12 cells deprived of nerve growth factor, a model of programmed cell death as described in detail in the experimental section. PC12 cells are an adrenal gland cell line from rat that provides a pre-neuron set that can be differentiated *in vitro*. The application of nerve growth factor induces the formation of axons and dendritic structures. This serves as a model for neuronal differentiation. When the nerve growth factor is withdrawn, the cells undergo programmed cell death (apoptosis). Approximately 300 control nucleic acid sequences (of known function) were added as an internal control and for transcriptional profiling of the cloned cDNA sequences. These sequences were then subjected to BLASTX analysis to determine the correspondence between the cDNA and a known cDNA and to determine to which protein family, if any, the proteins encoded by each cDNA belong. Computer analysis was used to assemble the cDNA sequences into unique clusters. The majority of the clusters as well as control genes were gridded on Smart Chip™ I.

Figure 2 shows the coefficient of variation (standard deviation/mean for triplicate hybridizations) after normalization for each array element plotted against the mean intensity for the gene (gene expression intensity). The figure shows the moving average (with a window of 200) for three different mRNA probes, 3 hour KCl-withdrawn, 3 hour control, and 6 hour control (See the examples and figures 3 and 4). As is typical for all probes, past a threshold of 30 to 40, the coefficient of variation averages below 0.2. The inset compares one triplicate hybridization (Filter Y) to another (Filter Z). Each point represents a different gene graphed on log-log axis comparing the intensity measured on one filter versus the other.

Figure 3 shows temporal expression clusters observed following KCl and serum withdrawal. A hierarchical clustering algorithm was used to cluster genes based on expression patterns across 10 time points (from left to right), 1, 3, 6, 12, and 24 hours post-KCl/serum-replacement (sham), and 1, 3, 6, 12, and 24 hours post-KCl/serum-withdrawal (treatment) (See Examples). Expression values for each gene were scaled based on the number of standard deviations from the mean intensity of each gene across all 10 time points. Scaled expression values are color-coded such

that red, yellow, and blue indicated above, at, and below mean intensity, respectively. The correlation between expression patterns of neighboring genes is depicted by the dendrogram on the right. Genes regulated by programmed cell death (KCl/serum-withdrawal alone) are enlarged in B. Representative non-scaled gene expression bar graphs with standard deviation error bars are aligned next to the four major clusters for Late Effector, Middle, Early, and Immediate Early gene expression classes. Regulated genes within each temporal expression class are listed in order of hierarchical clustering in SEQ ID NOS: 1-6, 8, and 106.

Figure 4 shows expression clusters for all of the CGN programmed cell death models (KCl and serum withdrawal, KCl withdrawal alone, and kainate treatment). Figure 4A shows a self-organizing map (SOM) algorithm (See, e.g. Kohonen, Self Organizing Maps: Springer, Berlin (1997)) that was used to cluster genes based on expression in 26 experiments (in order: serum added back, 1, 3, 6, 12, 24 hours; KCl/serum withdrawal, 1, 3, 6, 12, 24 hours; controls for KCl withdrawal, 1, 3, 6, 12 hours; KCl withdrawal alone, 1, 3, 6, 12 hours; controls for kainate treatment, 2, 4, 8, 12 hours; kainate treatment, 2, 4, 8, 12 hours; see examples for experimental details). As shown, a 5 x 4 geometry was used to organize the genes into 20 groups. A cluster (3,3) of 17 programmed cell death-induced genes is highlighted. The inset shows a tiled depiction of all the genes in the (3,3) cluster; red = above mean expression, white = mean expression, blue = below mean expression; the tiles are ordered in columns as indicated above for experimental order; each row represents a different array element gene in the order indicated by distance from the cluster centroid. Caspase 3, a gene involved in apoptosis, is part of the array and depicted in the raw values graph (i.e. relative expression in the 26 experiments); each experiment is represented in order on the x-axis; the y-axis indicates gene expression intensity.

Figure 4 B, C, D, and E show the raw gene expression intensity plotted for a representative gene from programmed cell death-regulated, regulated by KCl withdrawal only, immediate early genes, and serum-repressed constitutive expression classes, respectively. Each panel shows the data for a representative member of the cluster (indicated in the gene list by *), along with a list of genes included in the expression cluster.

Figure 4B shows the raw gene expression intensity for a gene representative from the list on the right. The graph shows increased expression with KCl and serum withdrawal, and kainate treatment. Accordingly, genes with these characteristics are designated "programmed cell death regulated." The list of genes with this pattern (on the chip) is shown on the right. Known genes include genes regulated in apoptosis.

Figure 4C lists genes which show increased expression after withdrawal of KCL or KCL and serum, but following kainate treatment. The list includes genes known to be involved in apoptosis.

Figure 4D shows genes that demonstrate constitutive immediate early expression.

Figure 4E shows genes that demonstrate constitutive expression in the absence of serum. The list on the right shows that this class contains mediators of programmed cell death.

Figure 5 shows information relating to various NARC genes. Accordingly the first column gives the NARC (neuronal apoptosis regulated candidate) designation. The second column provides specific information, such as the number of nucleotides sequenced, the region sequenced, for example, the 3' untranslated region, information regarding open reading frames, information regarding human orthologs (whose sequences may also be found in SEQ ID NOS: 1-6, 8, and 10), information regarding homology to known amino acid or nucleotide sequences, information regarding function, and other information related to specific physical or functional characteristics. The third column shows the gene expression class as described and designated in Figure 4. The fourth column shows the results of Northern blot hybridization, for example whether expression is restricted to specific organs or ubiquitous, and transcript size.

Figure 6 shows a tabulation of expression data of genes known to be related to programmed cell death, the data being obtained from experiments disclosed herein wherein nucleic acid sequences on the microarray were hybridized to mRNA derived from the two programmed cell death models (see examples). The first column indicates the clone designation. Where the clone is a previously known gene (for example, c-fos and c-jun), the gene name is given rather than the cDNA clone designation. The second column indicates the gene designation for each clone based on a BLASTX search. The third column indicates the expression pattern for each of

the clones. This tabulation can serve as an internal control to assess the fidelity of the experimental conditions and thus can serve as a background to compare the expression pattern of uncharacterized clones in the array. Accordingly, this figure shows a subarray that can serve as an internal control for discovering genes related to apoptosis and cell proliferation.

Figure 7 shows all genes (i.e., that are represented by nucleic acid sequences on the chip) that are regulated in specific experimental conditions described in the examples and shown in Figure 4. Specific genes are clustered (in an underlined category). Each cluster represents clones having a specific expression characteristic.

For example, the first cluster is transiently down-regulated by serum and down-regulated by KCl withdrawal. The second column identifies cDNA clones whose function is previously known. The third column indicates the cluster number. See Figure 4A. In addition, an analysis of the functions of the genes in each cluster showed that within a cluster, certain functional classes of genes may be over-represented. Thus, the material in parentheses indicates the biological functions that are associated with a disproportionate number of genes in the cluster. This includes secretion and synaptic vesicle release (cluster 0,0), cell proliferation (cluster 0,3), secretion/synaptic vesicle release/cytoskeletal reorganization (cluster 1,0), stress response/hormone response (cluster 1,3), stress response/hormone response (cluster 1,4), calcium signal transduction (cluster 2,0), and cytoskeleton/synapse cytoskeleton (cluster 2,4).

Figure 8 summarizes tissue expression data for the Smart Chip I™ microarray elements. The data were obtained by membrane blotting of the microarray against mRNA from testes, brain, heart, smooth muscle, spleen, kidney, skeletal muscle, lung, liver, and pancreatic tissue. Following hybridization with labeled cDNA synthesized from RNA from the indicated tissue type, the signal from each sequence on the array filters was quantitated by phosphorimaging.

Figure 9 provides a list of genes that were shown to be regulated by KCl and serum withdrawal in the microarray experiments described herein.

DETAILED DESCRIPTION OF THE INVENTION

I. Isolated Nucleic Acid Molecules

5 The invention encompasses the discovery and isolation of nucleic acid molecules that are expressed in rat brain and in programmed cell death *in vitro* models (neuronal apoptosis regulated candidates or NARCs) and their human homologs. The sequences of these human homologs are specifically disclosed in SEQ ID NOS:1 (human NARC 9B), 2 (human NARC 8B), 3 (human NARC 2A), 4 (human NARC 16B), 5 (human NARC 10C), 6 (human NARC 1C), 8 (human NARC 1A), and 10 (human NARC 25).

As appropriate, the isolated nucleic acid molecules of the present invention can be RNA, for example, mRNA, or DNA, such as cDNA and genomic DNA. DNA molecules can be double-stranded or single-stranded; single stranded RNA or DNA 15 can be either the coding, or sense, strand or the non-coding, or antisense, strand. The nucleic acid molecule can include all or a portion of the coding sequence of the genes of the invention. Additionally, the nucleic acid molecule can be fused to a marker sequence, for example, a sequence that encodes a polypeptide to assist in isolation or purification of the polypeptide. Such sequences include, but are not limited to, those 20 which encode a glutathione-S-transferase (GST) fusion protein and those which encode a hemagglutinin A (HA) polypeptide marker from influenza.

An "isolated" nucleic acid molecule, as used herein, is one that is separated from nucleic acid which normally flanks the nucleic acid molecule in nature. With regard to genomic DNA, the term "isolated" refers to nucleic acid molecules which 25 are separated from the chromosome with which the genomic DNA is naturally associated. For example, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotides which flank the nucleic acid molecule in the genomic DNA of the cell from which the nucleic acid is derived.

30 Moreover, an isolated nucleic acid of the invention, such as a cDNA or RNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or chemical precursors or other chemicals when chemically synthesized. However, the nucleic acid molecule can be fused to other

coding or regulatory sequences and still be considered isolated. In some instances, the isolated material will form part of a composition (for example, a crude extract containing other substances), buffer system or reagent mix. In other circumstances, the material may be purified to essential homogeneity, for example as determined by
5 PAGE or column chromatography such as HPLC. Preferably, an isolated nucleic acid comprises at least about 50, 80 or 90% (on a molar basis) of all macromolecular species present.

Further, recombinant DNA contained in a vector is included in the definition of "isolated" as used herein. Also, isolated nucleic acid molecules include
10 recombinant DNA molecules in heterologous host cells, as well as partially or substantially purified DNA molecules in solution. "Isolated" nucleic acid molecules also encompass *in vivo* and *in vitro* RNA transcripts of the DNA molecules of the present invention.

The invention further provides variants of the isolated nucleic acid molecules
15 of the invention. Such variants can be naturally occurring, such as allelic variants (same locus), homologs (different locus), and orthologs (different organism), or may be constructed by recombinant DNA methods or by chemical synthesis. Such non-naturally occurring variants can be made using well-known mutagenesis techniques, including those applied to polynucleotides, cells, or organisms.
20 Accordingly, variants can contain nucleotide substitutions, deletions, inversions and/or insertions in either or both the coding and non-coding region of the nucleic acid molecule. Further, the variations can produce both conservative and non-conservative amino acid substitutions.

Typically, variants have a substantial identity with a nucleic acid molecule
25 selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements thereof. Particularly preferred are nucleic acid molecules and fragments which have at least about 60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99% or more identity
30 with nucleic acid molecules described herein.

Such nucleic acid molecules can be readily identified as being able to hybridize under stringent conditions to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the

complements thereof. In one embodiment, the variants hybridize under high stringency hybridization conditions (e.g., for selective hybridization) to a nucleotide sequence selected from the sequences shown in SEQ ID NOS:1-6, 8, and 10.

As used herein, the term "hybridizes under stringent conditions" describes conditions for hybridization and washing. Stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. Aqueous and nonaqueous methods are described in that reference and either can be used. A preferred, example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 50°C. Another example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 55°C. A further example of stringent hybridization conditions is hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 60°C. Preferably, stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 65°C. Particularly preferred stringency conditions (and the conditions that should be used if the practitioner is uncertain about what conditions should be applied to determine if a molecule is within a hybridization limitation of the invention) are 0.5M Sodium Phosphate, 7% SDS at 65°C, followed by one or more washes at 0.2X SSC, 1% SDS at 65°C. The hybridization step may be performed for 4, 8, 12, or 16 hours, and the wash steps are generally 15 minutes or 30 minutes in length.

The percent identity of two nucleotide or amino acid sequences can be determined by aligning the sequences for optimal comparison purposes (e.g., gaps can be introduced in the sequence of a first sequence). The nucleotides or amino acids at corresponding positions are then compared, and the percent identity between the two sequences is a function of the number of identical positions shared by the sequences. In certain embodiments, the length of a sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 60%, and even more preferably at least 70%, 80% or 90% of the length of the reference sequence. The actual comparison of the two sequences can be accomplished by well-known methods, for example, using a mathematical algorithm. A preferred, non-limiting

example of such a mathematical algorithm is described in Karlin *et al.* (1993) *Proc. Natl. Acad. Sci. USA*, 90:5873-5877. Such an algorithm is incorporated into the NBLAST and XBLAST programs (version 2.0) as described in Altschul *et al.* (1997) *Nucleic Acids Res.*, 25:389-3402. When utilizing BLAST and Gapped BLAST
5 programs, the default parameters of the respective programs (e.g., NBLAST) can be used. See <http://www.ncbi.nlm.nih.gov>. In one embodiment, parameters for sequence comparison can be set at score=100, wordlength=12, or can be varied (e.g., W=5 or W=20).

Another preferred, non-limiting example of a mathematical algorithm utilized
10 for the comparison of sequences is the algorithm of Myers and Miller, CABIOS (1989). Such an algorithm is incorporated into the ALIGN program (version 2.0) which is part of the CGC sequence alignment software package. When utilizing the ALIGN program for comparing amino acid sequences, a PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used. Additional
15 algorithms for sequence analysis are known in the art and include ADVANCE and ADAM as described in Torellis and Robotti (1994) *Comput. Appl. Biosci.* 10:3-5; and FASTA described in Pearson and Lipman (1988) *PNAS*, 85:2444-8.

In another embodiment, the percent identity between two amino acid sequences can be accomplished using the GAP program in the CGC software package
20 (available at <http://www.cgc.com>) using either a BLOSUM 63 matrix or a PAM250 matrix, and a gap weight of 12, 10, 8, 6, or 4 and a length weight of 2, 3, or 4. In yet another embodiment, the percent identity between two nucleic acid sequences can be accomplished using the GAP program in the CGC software package (available at <http://www.cgc.com>), using a gap weight of 50 and a length weight of 3.

25 The present invention also provides isolated nucleic acids that contain a fragment or portion that hybridizes under highly stringent conditions to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS:1-6, 8, and 10. In one embodiment, the nucleic acid consists of a fragment of a
30 nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS:1-6, 8, and 10. The nucleic acid fragments of the invention are at least about 15, preferably at least about 18, 20, 23 or 25 nucleotides, and can be 30, 40, 50, 100, 200

or more nucleotides in length. Longer fragments, for example, 30 or more nucleotides in length, which encode antigenic proteins or polypeptides described herein are useful. Additionally, nucleotide sequences described herein can also be contigged (e.g., overlapped or joined) to produce longer sequences (see, for example, 5 <http://bozeman.mbt.washington.edu/phrap.docs/phrap.html>).

In a related aspect, the nucleic acid fragments of the invention are used as probes or primers in assays such as those described herein. "Probes" are oligonucleotides that hybridize in a base-specific manner to a complementary strand of nucleic acid. Such probes include polypeptide nucleic acids, as described in 10 Nielsen *et al.* (1991) *Science*, 254, 1497-1500. Typically, a probe comprises a region of nucleotide sequence that hybridizes under highly stringent conditions to at least about 15, typically about 20-25, and more typically about 40, 50 or 75 consecutive nucleotides of a nucleic acid selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements thereof. More typically, 15 the probe further comprises a label, *e.g.*, radioisotope, fluorescent compound, enzyme, or enzyme co-factor.

As used herein, the term "primer" refers to a single-stranded oligonucleotide which acts as a point of initiation of template-directed DNA synthesis using well-known methods (*e.g.*, PCR, LCR) including, but not limited to those described 20 herein. The appropriate length of the primer depends on the particular use, but typically ranges from about 15 to 30 nucleotides. The term "primer site" refers to the area of the target DNA to which a primer hybridizes. The term "primer pair" refers to a set of primers including a 5' (upstream) primer that hybridizes with the 5' end of the nucleic acid sequence to be amplified and a 3' (downstream) primer that hybridizes 25 with the complement of the sequence to be amplified.

The nucleic acid molecules of the invention such as those described above can be identified and isolated using standard molecular biology techniques and the sequence information provided in the sequences shown in SEQ ID NOS:1-6, 8, and 10. For example, nucleic acid molecules can be amplified and isolated by the 30 polymerase chain reaction using synthetic oligonucleotide primers designed based on one or more of the sequences provided in the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. See generally *PCR Technology: Principles and Applications for DNA Amplification* (ed. H.A. Erlich, Freeman Press, NY, NY,

1992); *PCR Protocols: A Guide to Methods and Applications* (Eds. Innis, *et al.* Academic Press, San Diego, CA, 1990); Mattila *et al.* (1991) *Nucleic Acids Res.* 19:4967; Eckert *et al.* (1991) *PCR Methods and Applications*, 1:17; PCR (eds. McPherson *et al.* IRL Press, Oxford); and U.S. Patent 4,683,202. The nucleic acid
5 molecules can be amplified using cDNA, mRNA or genomic DNA as a template, cloned into an appropriate vector and characterized by DNA sequence analysis.

Other suitable amplification methods include the ligase chain reaction (LCR) (see Wu and Wallace (1989) *Genomics*, 4:560, Landegren *et al.* (1988) *Science*, 241:1077, transcription amplification (Kwoh *et al.* (1989) *Proc. Natl. Acad. Sci. USA*, 86:1173), and self-sustained sequence replication (Guatelli *et al.* (1990) *Proc. Nat. Acad. Sci. USA*, 87:1874) and nucleic acid based sequence amplification (NASBA). The latter two amplification methods involve isothermal reactions based on isothermal transcription, which produce both single stranded RNA (ssRNA) and double stranded DNA (dsDNA) as the amplification products in a ratio of about 30 or
15 100 to 1, respectively.

The amplified DNA can be radiolabelled and used as a probe for screening a cDNA library, mRNA in zap express, ZIPLOX or other suitable vector. Corresponding clones can be isolated, DNA can obtained following *in vivo* excision, and the cloned insert can be sequenced in either or both orientations by art recognized
20 methods to identify the correct reading frame encoding a protein of the appropriate molecular weight. For example, the direct analysis of the nucleotide sequence of nucleic acid molecules of the present invention can be accomplished using well-known methods that are commercially available. See, for example, Sambrook *et al.* *Molecular Cloning, A Laboratory Manual* (2nd Ed., CSHP, New York 1989);
25 Zyskind *et al.* *Recombinant DNA Laboratory Manual*, (Acad. Press, 1988)). Using these or similar methods, the protein(s) and the DNA encoding the protein can be isolated, sequenced and further characterized.

Antisense nucleic acids of the invention can be designed using the nucleotide sequences of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and constructed
30 using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase

the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

Additionally, the nucleic acid molecules of the invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, *e.g.*, the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup *et al.* (1996) *Bioorganic & Medicinal Chemistry*, 4:5). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.*, DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup *et al.* (1996), *supra*; Perry-O'Keefe *et al.* (1996) *Proc. Natl. Acad. Sci. USA*, 93:14670. PNAs can be further modified, *e.g.*, to enhance their stability, specificity or cellular uptake, by attaching lipophilic or other helper groups to PNA, by the formation of

PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996), *supra*, Finn *et al.* (1996) *Nucleic Acids Res.*

24(17):3357-63, Mag *et al.* (1989) *Nucleic Acids Res.* 17:5973, and Peterser *et al.*

5 (1975) *Bioorganic Med. Chem. Lett.* 5:1119.

The nucleic acid molecules and fragments of the invention can also include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, *e.g.*, Letsinger *et al.*

(1989) *Proc. Natl. Acad. Sci. USA*, 86:6553-6556; Lemaitre *et al.* (1987) *Proc. Natl.*

10 *Acad. Sci. USA*, 84:648-652; PCT Publication No. WO88/0918) or the blood brain barrier (see, *e.g.*, PCT Publication No. WO89/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (see, *e.g.*, Krol *et al.* (1988) *Bio-Techniques*, 6:958-976) or intercalating agents (see, *e.g.*, Zon (1988) *Pharm Res.* 5:539-549).

15 Uses of the nucleic acids of the invention are described in detail in below. In general, the isolated nucleic acid sequences can be used as molecular weight markers on Southern gels, and as chromosome markers which are labeled to map related gene positions. The nucleic acid sequences can also be used to compare with endogenous DNA sequences in patients to identify genetic disorders, and as probes, such as to
20 hybridize and discover related DNA sequences or to subtract out known sequences from a sample. The nucleic acid sequences can further be used to derive primers for genetic fingerprinting, to raise anti-protein antibodies using DNA immunization techniques, and as an antigen to raise anti-DNA antibodies or elicit immune responses. Additionally, the nucleotide sequences of the invention can be used
25 identify and express recombinant proteins for analysis, characterization or therapeutic use, or as markers for tissues in which the corresponding protein is expressed, either constitutively, during tissue differentiation, or in disease states.

II. Vectors and Host Cells

30 Another aspect of the invention pertains to nucleic acid vectors containing a nucleic acid selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10. These vectors comprise a sequence of the invention that has been inserted in a sense or antisense orientation. As used herein, the term "vector"

refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors, expression vectors, are capable of directing the expression of genes to which they are operably linked. In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids (vectors). However, the invention is intended to include such other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses) that serve equivalent functions.

Preferred recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell. This means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operably linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (*e.g.*, polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel, *Gene Expression Technology: Methods in Enzymology 185*, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and those which direct expression of the nucleotide sequence only in certain host cells (*e.g.*, tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be

transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein .

5 The recombinant expression vectors of the invention can be designed for expression of a polypeptide of the invention in prokaryotic or eukaryotic cells, *e.g.*, bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors), yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *supra*. Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7
10 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein.
15 Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable
20 separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson (1988) *Gene*, 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding
25 protein, or protein A, respectively, to the target recombinant protein.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann *et al.* (1988) *Gene*, 69:301-315) and pET 11d (Studier *et al.* *Gene Expression Technology: Methods in Enzymology*, 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host
30 RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral

polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 *gn1* gene under the transcriptional control of the *lacUV 5* promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to
5 express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, *Gene Expression Technology: Methods in Enzymology*, 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those
10 preferentially utilized in *E. coli* (Wada *et al.* (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari
15 *et al.* (1987) *EMBO J.* 6:229-234), pMFa (Kurjan and Herskowitz (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.* (1987) *Gene*, 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and pPicZ (InVitrogen Corp, San Diego, CA).

Alternatively, a nucleic acid of the invention can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of
20 proteins in cultured insect cells (*e.g.*, Sf 9 cells) include the pAc series (Smith *et al.* (1983) *Mol. Cell Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology*, 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian
25 expression vectors include pCDM8 (Seed (1987) *Nature*, 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both
30 prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook *et al. supra*.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (*e.g.*, tissue-specific regulatory elements are used to express the nucleic acid).

Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.* (1987) *Genes Dev.* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv. Immunol.* 43:235-275), in particular promoters of T cell
5 receptors (Winoto and Baltimore (1989) *EMBO J.* 8:729-733) and immunoglobulins (Banerji *et al.* (1983) *Cell*, 33:729-740; Queen and Baltimore (1983) *Cell*, 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) *Proc. Natl. Acad. Sci. USA*, 86:5473-5477), pancreas-specific promoters (Edlund *et al.* (1985) *Science*, 230:912-916), and mammary gland-specific
10 promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) *Science*, 249:374-379) and the alpha-fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev.* 3:537-546).

15 The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operably linked to at least one expression control element in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to an mRNA of the invention.
20 Regulatory sequences operably linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression
25 vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub *et al.* (*Reviews - Trends in Genetics*, Vol. 1(1) 1986).

30 Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but also to the progeny or potential

progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

5 A host cell can be any prokaryotic or eukaryotic cell. For example, a nucleic acid of the invention can be expressed in bacterial cells (*e.g.*, *E. coli*), insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

 Vector DNA can be introduced into prokaryotic or eukaryotic cells via
10 conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or
15 transfecting host cells can be found in Sambrook, *et al.* (*supra*), and other laboratory manuals.

 For stable transfection of mammalian cells, it is known that, depending upon
the expression vector and transfection technique used, only a small fraction of cells
may integrate the foreign DNA into their genome. In order to identify and select
20 these integrants, a gene that encodes a selectable marker (*e.g.*, for resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can
be introduced into a host cell on the same vector as that nucleic acid of the invention
25 or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

 A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) a polypeptide of the invention.
30 Accordingly, the invention further provides methods for producing a polypeptide using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding a polypeptide of the invention has been introduced) in a suitable medium

such that the polypeptide is produced. In another embodiment, the method further comprises isolating the polypeptide from the medium or the host cell.

The host cells of the invention can also be used to produce nonhuman transgenic animals. For example, in one embodiment, a host cell of the invention is a
5 fertilized oocyte or an embryonic stem cell into which a nucleic acid of the invention have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous nucleotide sequences have been introduced into their genome or homologous recombinant animals in which endogenous nucleotide sequences have been altered. Such animals are useful for studying the
10 function and/or activity of the nucleotide sequence and polypeptide encoded by the sequence and for identifying and/or evaluating modulators of their activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include
15 non-human primates, sheep, dogs, cows, goats, chickens, amphibians, etc. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, an "homologous recombinant
20 animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, *e.g.*, an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing a nucleic
25 acid of the invention into the male pronuclei of a fertilized oocyte, *e.g.*, by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The sequence can be introduced as a transgene into the genome of a non-human animal. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression
30 of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to the transgene to direct expression of a polypeptide in particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are

described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, U.S. Patent No. 4,873,191 and in Hogan, *Manipulating the Mouse Embryo* (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified
5 based upon the presence of the transgene in its genome and/or expression of mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding the transgene can further be bred to other transgenic animals carrying other transgenes.

10 Homologously recombinant host cells can also be produced that allow the *in situ* alteration of endogenous polynucleotide sequences of the invention in a host cell genome. The host cell includes, but is not limited to, a stable cell line, cell *in vivo*, or cloned microorganism. This technology is more fully described in WO 93/09222, WO 91/12650, WO 91/06667, U.S. 5,272,071, and U.S. 5,641,670. Briefly, specific
15 polynucleotide sequences corresponding to the polynucleotides or sequences proximal or distal to a gene are allowed to integrate into a host cell genome by homologous recombination where expression of the gene can be affected. In one embodiment, regulatory sequences are introduced that either increase or decrease expression of an endogenous sequence. Accordingly, a protein can be produced in a cell not normally
20 producing it. Alternatively, increased expression of a protein can be effected in a cell normally producing the protein at a specific level. Further, expression can be decreased or eliminated by introducing a specific regulatory sequence. The regulatory sequence can be heterologous to the protein sequence or can be a homologous sequence with a desired mutation that affects expression. Alternatively, the entire gene can be deleted.
25 The regulatory sequence can be specific to the host cell or capable of functioning in more than one cell type. Still further, specific mutations can be introduced into any desired region of the gene to produce mutant proteins of the invention. Such mutations could be introduced, for example, into the specific functional regions.

To create an homologous recombinant animal, a vector is prepared which
30 contains at least a portion of a nucleic acid of the invention into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, functionally disrupt, the endogenous gene. In one embodiment, the vector is designed such that, upon homologous recombination, the endogenous gene is functionally disrupted (*i.e.*, no

longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous gene is mutated or otherwise altered but still encodes functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous protein). In the homologous recombination vector, the altered portion of the gene is flanked at its 5' and 3' ends by additional nucleic acid of the gene to allow for homologous recombination to occur between the exogenous gene carried by the vector and an endogenous gene in an embryonic stem cell. The additional flanking nucleic acid is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the vector (see, e.g., Thomas and Capecchi (1987) *Cell* 51:503 for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced nucleic acid has homologously recombined with the endogenous gene are selected (see, e.g., Li *et al.* (1992) *Cell* 69:915). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see, e.g., Bradley in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination vectors and homologous recombinant animals are described further in Bradley (1991) *Current Opinion in Bio/Technology* 2:823-829 and in PCT Publication Nos. WO 90/11354, WO 91/01140, WO 92/0968, and WO 93/04169.

In another embodiment, transgenic non-human animals can be produced which contain selected systems that allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.* (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate

expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the
5 other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut *et al.* (1997) *Nature* 385:810-813 and PCT Publication Nos. WO 97/07668 and WO 97/07669.

10 III. Polypeptides

The present invention also provides isolated polypeptides and variants and fragments thereof that are encoded by the nucleic acid molecules of the invention, especially as shown in SEQ ID NOS: 1-6, 8, and 10. For example, as described above, the nucleotide sequences can be used to design primers to clone and express
15 cDNAs encoding the polypeptides of the invention. Further, the nucleotide sequences of the invention, *e.g.*, the sequences shown in SEQ ID NOS: 1-6, 8, and 10, can be analyzed using routine search algorithms (*e.g.*, BLAST, Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410; BLAZE, Brutlag *et al.* (1993) *Comp. Chem.* 17:203-207) to identify open reading frames (ORFs).

20 As used herein, a polypeptide is said to be "isolated" or "purified" when it is substantially free of cellular material when it is isolated from recombinant and non-recombinant cells, or free of chemical precursors or other chemicals when it is chemically synthesized. A polypeptide, however, can be joined to another polypeptide with which it is not normally associated in a cell and still be "isolated" or
25 "purified."

The polypeptides of the invention can be purified to homogeneity. It is understood, however, that preparations in which the polypeptide is not purified to homogeneity are useful and considered to contain an isolated form of the polypeptide. The critical feature is that the preparation allows for the desired function of the
30 polypeptide, even in the presence of considerable amounts of other components. Thus, the invention encompasses various degrees of purity. In one embodiment, the language "substantially free of cellular material" includes preparations of the polypeptide having less than about 30% (by dry weight) other proteins (*i.e.*,

contaminating protein), less than about 20% other proteins, less than about 10% other proteins, or less than about 5% other proteins.

When a polypeptide is recombinantly produced, it can also be substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, less than about 10%, or less than about 5% of the volume of the protein preparation. The language "substantially free of chemical precursors or other chemicals" includes preparations of the polypeptide in which it is separated from chemical precursors or other chemicals that are involved in its synthesis. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of the polypeptide having less than about 30% (by dry weight) chemical precursors or other chemicals, less than about 20% chemical precursors or other chemicals, less than about 10% chemical precursors or other chemicals, or less than about 5% chemical precursors or other chemicals.

In one embodiment, a polypeptide comprises an amino acid sequence encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. However, the invention also encompasses sequence variants. Variants include a substantially homologous protein encoded by the same genetic locus in an organism, *i.e.*, an allelic variant. Variants also encompass proteins derived from other genetic loci in an organism, but having substantial homology to a polypeptide encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. Variants also include proteins substantially homologous to these polypeptides but derived from another organism, *i.e.*, an ortholog. Variants also include proteins that are substantially homologous to these polypeptides that are produced by chemical synthesis. Variants also include proteins that are substantially homologous or identical to these polypeptides that are produced by recombinant methods.

As used herein, two proteins (or a region of the proteins) are substantially homologous or identical when the amino acid sequences are at least about 45-55%, typically at least about 70-75%, more typically at least about 80-85%, and most typically at least about 90, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical. A substantially homologous amino acid sequence, according to the present invention, will be encoded by a nucleic acid hybridizing to a nucleic acid

sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, or fragment thereof under stringent conditions as more described above.

To determine the percent similarity or identity of two amino acid sequences, or of two nucleic acids, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the sequence of one protein or nucleic acid for optimal alignment with the other protein or nucleic acid). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in one sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the other sequence, then the molecules are homologous at that position. As used herein, amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity". The percent homology between the two sequences is a function of the number of identical positions shared by the sequences (*i.e.*, per cent homology equals the number of identical positions/total number of positions times 100).

The invention also encompasses polypeptides having a lower degree of identity but having sufficient similarity so as to perform one or more of the same functions performed by a polypeptide encoded by a nucleic acid of the invention. Similarity is determined by conserved amino acid substitution. Such substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Conservative substitutions are likely to be phenotypically silent. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu, and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr. Guidance concerning which amino acid changes are likely to be phenotypically silent are found in Bowie *et al.* (1990) *Science* 247:1306-1310.

TABLE 1. Conservative Amino Acid Substitutions.

Aromatic	Phenylalanine
	Tryptophan
	Tyrosine
Hydrophobic	Leucine
	Isoleucine
	Valine
Polar	Glutamine
	Asparagine
Basic	Arginine
	Lysine
	Histidine
Acidic	Aspartic Acid
	Glutamic Acid
Small	Alanine
	Serine
	Threonine
	Methionine
	Glycine

- Both identity and similarity can be readily calculated (*Computational*
- 5 *Molecular Biology*, Lesk, A.M., ed., Oxford University Press, New York, 1988;
Biocomputing: Informatics and Genome Projects, Smith, D.W., ed., Academic Press,
New York, 1993; *Computer Analysis of Sequence Data, Part 1*, Griffin, A.M., and
Griffin, H.G., eds., Humana Press, New Jersey, 1994; *Sequence Analysis in Molecular*
Biology, von Heinje, G., Academic Press, 1987; and *Sequence Analysis Primer*,
- 10 Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991).

Preferred computer program methods to determine identity and similarity
between two sequences include, but are not limited to, GCG program package
(Devereux, J., *et al.* (1984) *Nucleic Acids Res.* 12(1):387), BLASTP, BLASTN,
FASTA (Atschul, S.F. *et al.* (1990) *J. Molec. Biol.* 215:403).

A variant polypeptide can differ in amino acid sequence by one or more substitutions, deletions, insertions, inversions, fusions, and truncations or a combination of any of these. Further, variant polypeptides can be fully functional or can lack function in one or more activities. Fully functional variants typically contain only conservative variation or variation in non-critical residues or in non-critical regions. Functional variants can also contain substitution of similar amino acids that result in no change or an insignificant change in function. Alternatively, such substitutions may positively or negatively affect function to some degree.

Non-functional variants typically contain one or more non-conservative amino acid substitutions, deletions, insertions, inversions, or truncation or a substitution, insertion, inversion, or deletion in a critical residue or critical region.

As indicated, variants can be naturally-occurring or can be made by recombinant means or chemical synthesis to provide useful and novel characteristics for the polypeptide. This includes preventing immunogenicity from pharmaceutical formulations by preventing protein aggregation.

Amino acids that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham *et al.* (1989) *Science* 244:1081-1085). The latter procedure introduces single alanine mutations at every residue in the molecule. The resulting mutant molecules are then tested for biological activity *in vitro*, or *in vitro* proliferative activity. Sites that are critical for polypeptide activity can also be determined by structural analysis such as crystallization, nuclear magnetic resonance or photoaffinity labeling (Smith *et al.* (1992) *J. Mol. Biol.* 224:899-904; de Vos *et al.* (1992) *Science* 255:306-312).

The invention also includes polypeptide fragments of the polypeptides of the invention. Fragments can be derived from a polypeptide encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements thereof. However, the invention also encompasses fragments of the variants of the polypeptides described herein.

As used herein, a fragment comprises at least 6 contiguous amino acids. Useful fragments include those that retain one or more of the biological activities of

the polypeptide as well as fragments that can be used as an immunogen to generate polypeptide specific antibodies.

Biologically active fragments (peptides which are, for example, 6, 9, 12, 15, 20, 30, 35, 36, 37, 38, 39, 40, 50, 100 or more amino acids in length) can comprise a domain, segment, or motif that has been identified by analysis of the polypeptide sequence using well-known methods, *e.g.*, signal peptides, extracellular domains, one or more transmembrane segments or loops, ligand binding regions, zinc finger domains, DNA binding domains, acylation sites, glycosylation sites, or phosphorylation sites.

The invention also provides fragments with immunogenic properties. These contain an epitope-bearing portion of the polypeptides and variants of the invention. These epitope-bearing peptides are useful to raise antibodies that bind specifically to a polypeptide or region or fragment. These peptides can contain at least 6, 7, 8, 9, 12, at least 14, or between at least about 15 to about 30 amino acids. The epitope-bearing peptide and polypeptides may be produced by any conventional means (Houghten (1985) *Proc. Natl. Acad. Sci. USA* 82:5131-5135). Simultaneous multiple peptide synthesis is described in U.S. Patent No. 4,631,211.

Fragments can be discrete (not fused to other amino acids or polypeptides) or can be within a larger polypeptide. Further, several fragments can be comprised within a single larger polypeptide. In one embodiment a fragment designed for expression in a host can have heterologous pre- and pro-polypeptide regions fused to the amino terminus of the polypeptide fragment and an additional region fused to the carboxyl terminus of the fragment.

The invention thus provides chimeric or fusion proteins. These comprise a polypeptide of the invention operatively linked to a heterologous protein having an amino acid sequence not substantially homologous to the polypeptide. "Operatively linked" indicates that the polypeptide protein and the heterologous protein are fused in-frame. The heterologous protein can be fused to the N-terminus or C-terminus of the polypeptide. In one embodiment the fusion protein does not affect function of the polypeptide *per se*. For example, the fusion protein can be a GST-fusion protein in which the polypeptide sequences are fused to the C-terminus of the GST sequences. Other types of fusion proteins include, but are not limited to, enzymatic fusion proteins, for example beta-galactosidase fusions, yeast two-hybrid GAL fusions,

poly-His fusions and Ig fusions. Such fusion proteins, particularly poly-His fusions, can facilitate the purification of recombinant polypeptide. In certain host cells (*e.g.*, mammalian host cells), expression and/or secretion of a protein can be increased by using a heterologous signal sequence. Therefore, in another embodiment, the fusion
5 protein contains a heterologous signal sequence at its N-terminus.

EP-A-O 464 533 discloses fusion proteins comprising various portions of immunoglobulin constant regions. The Fc is useful in therapy and diagnosis and thus results, for example, in improved pharmacokinetic properties (EP-A 0232 262). In drug discovery, for example, human proteins have been fused with Fc portions for the
10 purpose of high-throughput screening assays to identify antagonists. Bennett *et al.* (1995) *Journal of Molecular Recognition* 8:52-58 and Johanson *et al.* (1995) *The Journal of Biological Chemistry* 270,16:9459-9471. Thus, this invention also encompasses soluble fusion proteins containing a polypeptide of the invention and various portions of the constant regions of heavy or light chains of immunoglobulins
15 of various subclass (IgG, IgM, IgA, IgE). Preferred as immunoglobulin is the constant part of the heavy chain of human IgG, particularly IgG1, where fusion takes place at the hinge region. For some uses it is desirable to remove the Fc after the fusion protein has been used for its intended purpose, for example when the fusion protein is to be used as antigen for immunizations. In a particular embodiment, the Fc
20 part can be removed in a simple way by a cleavage sequence that is also incorporated and can be cleaved with factor Xa.

A chimeric or fusion protein can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different protein sequences are ligated together in-frame in accordance with conventional techniques. In another
25 embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of nucleic acid fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive nucleic acid fragments which can subsequently be annealed and re-amplified to generate a chimeric nucleic acid sequence (see
30 Ausubel *et al.*, *Current Protocols in Molecular Biology*, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST protein). A nucleic acid encoding a polypeptide of the invention can be

cloned into such an expression vector such that the fusion moiety is linked in-frame to the polypeptide protein.

The isolated polypeptide can be purified from cells that naturally express it, purified from cells that have been altered to express it (recombinant), or synthesized
5 using known protein synthesis methods.

In one embodiment, the protein is produced by recombinant DNA techniques. For example, a nucleic acid molecule encoding the polypeptide is cloned into an expression vector, the expression vector introduced into a host cell and the protein expressed in the host cell. The protein can then be isolated from the cells by an
10 appropriate purification scheme using standard protein purification techniques.

Polypeptides often contain amino acids other than the 20 amino acids commonly referred to as the 20 naturally-occurring amino acids. Further, many amino acids, including the terminal amino acids, may be modified by natural processes, such as processing and other post-translational modifications, or by
15 chemical modification techniques well known in the art. Common modifications that occur naturally in polypeptides are described in basic texts, detailed monographs, and the research literature, and they are well known to those of skill in the art.

Accordingly, the polypeptides also encompass derivatives or analogs in which a substituted amino acid residue is not one encoded by the genetic code, in which a
20 substituent group is included, in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence for purification of the mature polypeptide or a pro-protein sequence.

25 Known modifications include, but are not limited to, acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of
30 covalent crosslinks, formation of cystine, formation of pyroglutamate, formylation, gamma carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing,

phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

Such modifications are well-known to those of skill in the art and have been described in great detail in the scientific literature. Several particularly common
5 modifications, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic texts, such as *Proteins - Structure and Molecular Properties*, 2nd Ed., T.E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as by Wold, F., *Posttranslational*
10 *Covalent Modification of Proteins*, B.C. Johnson, Ed., Academic Press, New York 1-12 (1983); Seifter *et al.*, *Meth. Enzymol.* 182: 626-646 (1990) and Rattan *et al.* (1992) *Ann. N.Y. Acad. Sci.* 663:48-62.

As is also well known, polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination, and they may be
15 circular, with or without branching, generally as a result of post-translation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and branched circular polypeptides may be synthesized by non-translational natural processes and by synthetic methods.

Modifications can occur anywhere in a polypeptide, including the peptide
20 backbone, the amino acid side-chains and the amino or carboxyl termini. Blockage of the amino or carboxyl group in a polypeptide, or both, by a covalent modification, is common in naturally-occurring and synthetic polypeptides. For instance, the amino terminal residue of polypeptides made in *E. coli*, prior to proteolytic processing, almost invariably will be N-formylmethionine.

25 The modifications can be a function of how the protein is made. For recombinant polypeptides, for example, the modifications will be determined by the host cell posttranslational modification capacity and the modification signals in the polypeptide amino acid sequence. Accordingly, when glycosylation is desired, a polypeptide should be expressed in a glycosylating host, generally a eukaryotic cell.
30 Insect cells often carry out the same posttranslational glycosylations as mammalian cells and, for this reason, insect cell expression systems have been developed to efficiently express mammalian proteins having native patterns of glycosylation. Similar considerations apply to other modifications.

The same type of modification may be present in the same or varying degree at several sites in a given polypeptide. Also, a given polypeptide may contain more than one type of modification.

Uses of the polypeptides of the invention are described in detail below. In
5 general, polypeptides or proteins of the present invention can be used as a molecular weight marker on SDS-PAGE gels or on molecular sieve gel filtration columns using art-recognized methods. The polypeptides of the present invention can be used to raise antibodies or to elicit an immune response. The polypeptides can also be used as a reagent, *e.g.*, a labeled reagent, in assays to quantitatively determine levels of the
10 protein or a molecule to which it binds (*e.g.*, a receptor or a ligand) in biological fluids. The polypeptides can also be used as markers for tissues in which the corresponding protein is preferentially expressed, either constitutively, during tissue differentiation, or in a diseased state. The polypeptides can be used to isolate a corresponding binding partner, *e.g.*, receptor or ligand, such as, for example, in an
15 interaction trap assay, and to screen for peptide or small molecule antagonists or agonists of the binding interaction.

IV. Antibodies

In another aspect, the invention provides antibodies to the polypeptides and
20 polypeptide fragments of the invention, *e.g.*, having an amino acid encoded by a nucleic acid comprising all or a portion of a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen
25 binding site that specifically binds an antigen. A molecule that specifically binds to a polypeptide of the invention is a molecule that binds to that polypeptide or a fragment thereof, but does not substantially bind other molecules in a sample, *e.g.*, a biological sample, which naturally contains the polypeptide. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments
30 which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind to a polypeptide of the invention. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that

contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of a polypeptide of the invention. A monoclonal antibody composition thus typically displays a single binding affinity for a particular polypeptide of the invention with which it immunoreacts.

5 Polyclonal antibodies can be prepared as described above by immunizing a suitable subject with a desired immunogen, *e.g.*, polypeptide of the invention or fragment thereof. The antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized polypeptide. If desired, the antibody molecules directed
10 against the polypeptide can be isolated from the mammal (*e.g.*, from the blood) and further purified by well-known techniques, such as protein A chromatography to obtain the IgG fraction. At an appropriate time after immunization, *e.g.*, when the antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the
15 hybridoma technique originally described by Kohler and Milstein (1975) *Nature* 256:495-497, the human B cell hybridoma technique (Kozbor *et al.* (1983) *Immunol. Today* 4:72), the EBV-hybridoma technique (Cole *et al.* (1985), *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing hybridomas is well known (see generally *Current*
20 *Protocols in Immunology* (1994) Coligan *et al.* (eds.) John Wiley & Sons, Inc., New York, NY). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with an immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds a
25 polypeptide of the invention.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating a monoclonal antibody to a polypeptide of the invention (see, *e.g.*, *Current Protocols in Immunology, supra*; Galfre *et al.* (1977) *Nature* 266:55052; R.H. Kenneth, in
30 *Monoclonal Antibodies: A New Dimension In Biological Analyses*, Plenum Publishing Corp., New York, New York (1980); and Lerner (1981) *Yale J. Biol. Med.* 54:387-402. Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods that also would be useful. Typically, the immortal

cell line (*e.g.*, a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line, *e.g.*, a myeloma cell line that is sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a number of myeloma cell lines can be used as a fusion partner according to standard techniques, *e.g.*, the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind a polypeptide of the invention, *e.g.*, using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal antibody to a polypeptide of the invention can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (*e.g.*, an antibody phage display library) with the polypeptide to thereby isolate immunoglobulin library members that bind the polypeptide. Kits for generating and screening phage display libraries are commercially available (*e.g.*, the Pharmacia *Recombinant Phage Antibody System*, Catalog No. 27-9400-01; and the Stratagene *SurfZAP™* Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. Patent No. 5,223,409; PCT Publication No. WO 92/18619; PCT Publication No. WO 91/17271; PCT Publication No. WO 92/20791; PCT Publication No. WO 92/15679; PCT Publication No. WO 93/01288; PCT Publication No. WO 92/01047; PCT Publication No. WO 92/09690; PCT Publication No. WO 90/02809; Fuchs *et al.* (1991) *Bio/Technology* 9:1370-1372; Hay *et al.* (1992) *Hum. Antibod. Hybridomas* 3:81-85; Huse *et al.* (1989) *Science* 246:1275-1281; Griffiths *et al.* (1993) *EMBO J.* 12:725-734.

Additionally, recombinant antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can

- be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in PCT Publication No. WO 87/02671; European Patent Application 184,187;
- 5 European Patent Application 171,496; European Patent Application 173,494; PCT Publication No. WO 86/01533; U.S. Patent No. 4,816,567; European Patent Application 125,023; Better *et al.* (1988) *Science* 240:1041-1043; Liu *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:3439-3443; Liu *et al.* (1987) *J. Immunol.* 139:3521-3526; Sun *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:214-218; Nishimura
- 10 *et al.* (1987) *Canc. Res.* 47:999-1005; Wood *et al.* (1985) *Nature* 314:446-449; and Shaw *et al.* (1988) *J. Natl. Cancer Inst.* 80:1553-1559; Morrison (1985) *Science* 229:1202-1207; Oi *et al.* (1986) *Bio/Techniques* 4:214; U.S. Patent 5,225,539; Jones *et al.* (1986) *Nature* 321:552-525; Verhoeven *et al.* (1988) *Science* 239:1534; and Beidler *et al.* (1988) *J. Immunol.* 141:4053-4060.
- 15 Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Such antibodies can be produced using transgenic mice that are incapable of expressing endogenous immunoglobulin heavy and light chains genes, but which can express human heavy and light chain genes. The transgenic mice are immunized in the normal fashion with a selected antigen, *e.g.*, all or a
- 20 portion of a polypeptide of the invention. Monoclonal antibodies directed against the antigen can be obtained using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA
- 25 and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg and Huszar (1995) *Int. Rev. Immunol.* 13:65-93. For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, *e.g.*, U.S. Patent 5,625,126; U.S. Patent 5,633,425; U.S. Patent 5,569,825; U.S. Patent
- 30 5,661,016; and U.S. Patent 5,545,806. In addition, companies such as Abgenix, Inc. (Freemont, CA), can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

Completely human antibodies that recognize a selected epitope can be generated using a technique referred to as "guided selection." This technology is described, for example, in Jespers *et al.* (1994) *Bio/technology* 12:899-903).

Uses of the antibodies of the invention are described in detail below. In general, antibodies of the invention (*e.g.*, a monoclonal antibody) can be used to isolate a polypeptide of the invention by standard techniques, such as affinity chromatography or immunoprecipitation. A polypeptide specific antibody can facilitate the purification of natural polypeptide from cells and of recombinantly produced polypeptide expressed in host cells. Moreover, an antibody specific for a polypeptide of the invention can be used to detect the polypeptide (*e.g.*, in a cellular lysate, cell supernatant, or tissue sample) in order to evaluate the abundance and pattern of expression of the polypeptide. Antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, (-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H .

V. Computer Readable Means

The nucleotide or amino acid sequences of the invention are also provided in a variety of mediums to facilitate use thereof. As used herein, "provided" refers to a manufacture, other than an isolated nucleic acid or amino acid molecule, which contains a nucleotide or amino acid sequence of the present invention. Such a manufacture provides the nucleotide or amino acid sequences, or a subset thereof (*e.g.*, a subset of open reading frames (ORFs)) in a form which allows a skilled artisan

to examine the manufacture using means not directly applicable to examining the nucleotide or amino acid sequences, or a subset thereof, as they exists in nature or in purified form.

In one application of this embodiment, a nucleotide or amino acid sequence of the present invention can be recorded on computer readable media. As used herein, "computer readable media" refers to any medium that can be read and accessed directly by a computer. Such media include, but are not limited to: magnetic storage media, such as floppy discs, hard disc storage medium, and magnetic tape; optical storage media such as CD-ROM; electrical storage media such as RAM and ROM; and hybrids of these categories such as magnetic/optical storage media. The skilled artisan will readily appreciate how any of the presently known computer readable mediums can be used to create a manufacture comprising computer readable medium having recorded thereon a nucleotide or amino acid sequence of the present invention.

As used herein, "recorded" refers to a process for storing information on computer readable medium. The skilled artisan can readily adopt any of the presently known methods for recording information on computer readable medium to generate manufactures comprising the nucleotide or amino acid sequence information of the present invention.

A variety of data storage structures are available to a skilled artisan for creating a computer readable medium having recorded thereon a nucleotide or amino acid sequence of the present invention. The choice of the data storage structure will generally be based on the means chosen to access the stored information. In addition, a variety of data processor programs and formats can be used to store the nucleotide sequence information of the present invention on computer readable medium. The sequence information can be represented in a word processing text file, formatted in commercially-available software such as WordPerfect and MicroSoft Word, or represented in the form of an ASCII file, stored in a database application, such as DB2, Sybase, Oracle, or the like. The skilled artisan can readily adapt any number of dataprocessor structuring formats (e.g., text file or database) in order to obtain computer readable medium having recorded thereon the nucleotide sequence information of the present invention.

By providing the nucleotide or amino acid sequences of the invention in computer readable form, the skilled artisan can routinely access the sequence

information for a variety of purposes. For example, one skilled in the art can use the nucleotide or amino acid sequences of the invention in computer readable form to compare a target sequence or target structural motif with the sequence information stored within the data storage means. Search means are used to identify fragments or
5 regions of the sequences of the invention which match a particular target sequence or target motif.

As used herein, a "target sequence" can be any DNA or amino acid sequence of six or more nucleotides or two or more amino acids. A skilled artisan can readily recognize that the longer a target sequence is, the less likely a target sequence will be
10 present as a random occurrence in the database. The most preferred sequence length of a target sequence is from about 10 to 100 amino acids or from about 30 to 300 nucleotide residues. However, it is well recognized that commercially important fragments, such as sequence fragments involved in gene expression and protein processing, may be of shorter length.

As used herein, "a target structural motif," or "target motif," refers to any rationally selected sequence or combination of sequences in which the sequence(s) are
15 ~~chosen based on a three-dimensional configuration which is formed upon the folding~~ of the target motif. There are a variety of target motifs known in the art. Protein target motifs include, but are not limited to, enzyme active sites and signal sequences.
20 Nucleic acid target motifs include, but are not limited to, promoter sequences, hairpin structures and inducible expression elements (protein binding sequences).

Computer software is publicly available which allows a skilled artisan to access sequence information provided in a computer readable medium for analysis and comparison to other sequences. A variety of known algorithms are disclosed
25 publicly and a variety of commercially available software for conducting search means are and can be used in the computer-based systems of the present invention. Examples of such software includes, but is not limited to, MacPattern (EMBL), BLASTN and BLASTX (NCBIA).

For example, software which implements the BLAST (Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410) and BLAZE (Brutlag *et al.* (1993) *Comp. Chem.* 17:203-207)
30 search algorithms on a Sybase system can be used to identify open reading frames (ORFs) of the sequences of the invention which contain homology to ORFs or proteins from other libraries. Such ORFs are protein encoding fragments and are

useful in producing commercially important proteins such as enzymes used in various reactions and in the production of commercially useful metabolites.

VI. Detection Assays

5 Portions or fragments of the nucleotide sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue
10 typing); and (iii) aid in forensic identification of a biological sample. These applications are described in the subsections below.

1. Chromosome Mapping

 Once the nucleic acid (or a portion of the sequence) has been isolated, it can
15 be used to map the location of the gene on a chromosome. The mapping of the sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease. Briefly, genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the nucleic acid molecules described herein. Computer analysis of the sequences can be used to
20 predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the appropriate nucleotide sequences will yield an amplified fragment.

25 Somatic cell hybrids are prepared by fusing somatic cells from different mammals (*e.g.*, human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that
30 contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of

individual genes to specific human chromosomes. (D'Eustachio *et al.* (1983) *Science* 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

5 PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycle. Using the nucleic acid molecules of the invention to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can
10 similarly be used to map a specified sequence to its chromosome include *in situ* hybridization (described in Fan *et al.* (1990) *PNAS* 97:6223-27), pre-screening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a nucleotide sequence to a
15 metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each
20 chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a nucleotide sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good
25 results at a reasonable amount of time. for a review of this technique, see Verma *et al.*, *Human Chromosomes: A Manual of Basic Techniques* (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used
30 for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are preferred for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, *Medelian Inheritance in Man*, available on-line through Johns Hopkins University Welch Medical Library).

- 5 The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland *et al.* (1987) *Nature* 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with a specified gene, can be determined. If a
10 mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR
15 based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

2. Tissue Typing

20 The nucleotide sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield
25 unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an
30 alternative technique that determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the nucleic acid molecules described herein can be used to prepare two PCR primers from the 5' and 3' ends of

the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a
5 unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The nucleic acid molecules of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. It is
10 estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals. The
15 noncoding sequences of these sequences can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers which each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

20 If a panel of reagents from nucleic acid molecules described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

25

3. Use of Partial Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means of positively identifying, for example, a perpetrator
30 of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, *e.g.*, hair or skin, or body fluids, *e.g.*, blood, saliva, or semen found at a crime scene. The

amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

The sequences of the present invention can be used to provide polynucleotide reagents, *e.g.*, PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (*i.e.* another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Sequences targeted to noncoding regions of sequences described herein are particularly appropriate for this use, as greater numbers of polymorphisms occur in the noncoding regions, making it easier to differentiate individuals using this technique. Examples of polynucleotide reagents include the nucleic acid molecules or the invention, or portions thereof, *e.g.*, fragments having a length of at least 20 bases, preferably at least 30 bases.

The nucleic acid molecules described herein can further be used to provide polynucleotide reagents, *e.g.*, labeled or labelable probes which can be used in, or example, an *in situ* hybridization technique, to identify a specific tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, primers or probes can be used to screen tissue culture for contamination (*i.e.*, screen for the presence of a mixture of different types of cells in a culture).

VII. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining protein and/or nucleic acid expression as well as activity of proteins of the invention, in the context of a biological sample (*e.g.*, blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant expression or activity. The

invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with activity or expression of proteins or nucleic acids of the invention.

Disorders relating to programmed cell death are particularly relevant as
5 discussed in detail herein below.

For example, mutations in a specified gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby prophylactically treat an individual prior to the onset of a disorder characterized by or associated with expression or activity of nucleic acid molecules or proteins of the
10 invention.

Another aspect of the invention pertains to monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression or activity of proteins of the invention in clinical trials.

These and other agents are described in further detail in the following sections.
15

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of proteins or nucleic acids of the invention in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or
20 an agent capable of detecting the protein, or nucleic acid (*e.g.*, mRNA, genomic DNA) that encodes the protein, such that the presence of the protein or nucleic acid is detected in the biological sample. A preferred agent for detecting mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to mRNA or genomic DNA sequences described herein. The nucleic acid probe can be, for example, a
25 full-length nucleic acid, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to appropriate mRNA or genomic DNA. For example, the nucleic acid probe can be all or a portion of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, or the complement of the sequences shown in SEQ ID NOS: 1-6, 8, and
30 10, or a portion thereof. Other suitable probes for use in the diagnostic assays of the invention are described herein.

In one embodiment, the agent for detecting proteins of the invention is an antibody capable of binding to the protein, preferably an antibody with a detectable

label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (*e.g.*, Fab or F(ab')₂) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (*i.e.*, physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, calls and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect mRNA, protein, or genomic DNA of the invention in a biological sample *in vitro* as well as *in vivo*. For example, *in vitro* techniques for detection of mRNA include Northern hybridizations and *in situ* hybridizations. *In vitro* techniques for detection of protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of genomic DNA include Southern hybridizations. Furthermore, *in vivo* techniques for detection of protein include introducing into a subject a labeled anti-protein antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a serum sample or biopsy isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting protein, mRNA, or genomic DNA of the invention, such that the presence of protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of protein, mRNA or genomic DNA in the control sample with the presence of protein, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting the presence of proteins or nucleic acid molecules of the invention in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting protein or mRNA in a biological sample; means for determining the amount of in the sample; and means for
5 comparing the amount of in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect protein or nucleic acid.

2. Prognostic Assays

10 The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant expression or activity of proteins and nucleic acid molecules of the invention. Accordingly, the term "diagnostic" refers not only to ascertaining whether a subject has an active disease but also relates to ascertaining whether a subject is predisposed
15 to developing active disease as well as ascertaining the probability that treatment of active disease will be effective. For example, the assays described herein, such as the preceding diagnostic assays or the following assays can be utilized to identify a subject having or at risk of developing a disorder associated with protein or nucleic acid expression or activity such as a proliferative disorder, a differentiative or
20 developmental disorder, or a hematopoietic disorder. Alternatively, the prognostic assays can be utilized to identify a subject having or at risk for developing a differentiative or proliferative disease (e.g., cancer). Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant expression or activity of proteins or nucleic acid molecules of the invention, in which
25 a test sample is obtained from a subject and protein or nucleic acid (e.g., mRNA, genomic DNA) is detected, wherein the presence of protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant expression or activity of the protein or nucleic acid sequence of the invention. As used herein, a "test sample" refers to a biological sample
30 obtained from a subject of interest. For example, a test sample can be a biological fluid (e.g., serum), cell or tissue sample.

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, polypeptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant expression or activity of a protein or nucleic acid molecule of the invention. For example, such methods can be used to determine whether a subject can be effectively treated with an agent for a disorder, such as a proliferative disorder, a differentiative or a developmental disorder. Alternatively, such methods can be used to determine whether a subject can be effectively treated with an agent for a differentiative or proliferative disease (*e.g.*, cancer). Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant expression or activity of a protein or nucleic acid of the present invention, in which a test sample is obtained and protein or nucleic acid expression or activity is detected (*e.g.*, wherein the abundance of particular protein or nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant expression or activity.)

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein-below.

The methods of the invention can also be used to detect genetic alterations in genes or nucleic acid molecules of the present invention, thereby determining if a subject with the altered gene is at risk for a disorder characterized by aberrant development, aberrant cellular differentiation, aberrant cellular proliferation or an aberrant hematopoietic response. In certain embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding a particular protein, or the mis-expression of the gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of (1) a deletion of one or more nucleotides; (2) an addition of one or more nucleotides; (3) a substitution of one or more nucleotides, (4) a chromosomal rearrangement; (5) an alteration in the level of a messenger RNA transcript; (6) aberrant modification, such as of the methylation pattern of the genomic DNA; (7) the presence of a non-wild type splicing pattern of a messenger RNA transcript; (8) a non-wild type level; (9) allelic loss; and (10) inappropriate post-translational modification. As described

herein, there are a large number of assay techniques known in the art that can be used for detecting alterations in a particular gene. A preferred biological sample is a tissue or serum sample isolated by conventional means from a subject.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, *e.g.*, U.S. Patent Nos. 4,683,195 and 4,683,202), such an anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, *e.g.*, Landegran *et al.* (1988) *Science* 241:1077-1080; and Nakazawa *et al.* (1994) *PNAS* 91:360-364), the latter of which can be particularly useful for detecting point mutations (see Abravaya *et al.* (1995) *Nucleic Acids Res.* 23:675-682). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (*e.g.*, genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to the gene under conditions such that hybridization and amplification of the gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:1874-1878), transcriptional amplification system (Kwoh *et al.*, (1989) *Proc. Natl. Acad. Sci. USA* 86:1173-1177), Q-Beta Replicase (Lizardi *et al.* (1988) *Bio/Technology* 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in a given gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicate mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for sample, U.S. Patent No. 5,498,531) can be used

to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations can be identified by hybridizing a sample and control nucleic acids, *e.g.*, DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotide probes (Cronin *et al.* (1996) *Human Mutation* 7:244-255; Kozal *et al.* (1996) *Nature Medicine* 2:753-759). For example, genetic mutations can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. *et al. supra*. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the gene and detect mutations by comparing the sequence of the gene from the sample with the corresponding wild-type (control) gene sequence. Examples of sequencing reactions include those based on techniques developed by Maxim and Gilbert ((1997) *PNAS* 74:560) or Sanger ((1977) *PNAS* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, *e.g.*, PCT International Publication No. WO 94/16101; Cohen *et al.* (1996) *Adv. Chromatogr.* 36:127-162; and Griffin *et al.* (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

Other methods for detecting mutations include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers *et al.* (1985) *Science* 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wild-type sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-standard duplexes are treated with an agent that cleaves single-stranded regions of the duplex such as which

will exist due to base pair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with Rnase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digest the mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size
5 on denaturing polyacrylamide gels to determine the site of mutation. See, for example Cotton *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:4397; Saleeba *et al.* (1992) *Methods Enzymol.* 217:286-295. In certain embodiments, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or
10 more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994)
15 *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on a nucleotide sequence of the invention is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

20 In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:2766, see also Cotton (1993) *Mutat Res* 285:125-144; and Hayashi (1992) *Genet Anal. Tech.*
25 *Appl.* 9:73-79). Single-stranded DNA fragments of sample and control nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be
30 enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In one embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen *et al.* (1991) *Trends Genet.* 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers *et al.* (1985) *Nature* 313:495). When DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys. Chem.* 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki *et al.* (1986) *Nature* 324:163); Saiki *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:6320). Such allele-specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology that depends on selective PCR amplification may be used in conjunction with the instant invention. Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs *et al.* (1989) *Nucleic Acids Res.* 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner (1993) *Tibtech* 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini *et al.* (1992) *Mol. Cell Probes* 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) *Proc. Natl. Acad. Sci. USA* 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness

5 involving a gene of the present invention. Any cell type or tissue in which the gene is expressed may be utilized in the prognostic assays described herein.

3. Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression

10 or activity of nucleic acid molecules or proteins of the present invention (*e.g.*, modulation of cellular signal transduction, regulation of gene transcription in a cell involved in development or differentiation, regulation of cellular proliferation) can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to

15 increase gene expression, protein levels, or upregulate protein activity, can be monitored in clinical trials of subjects exhibiting decreased gene expression, protein levels, or downregulated protein activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease gene expression, protein levels, or downregulate protein activity, can be monitored in clinical trials of subjects exhibiting

20 increased gene expression, protein levels, or upregulated protein activity. In such clinical trials, the expression or activity of the specified gene and, preferably, other genes that have been implicated in, for example, a proliferative disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes that are modulated in cells

25 by treatment with an agent (*e.g.*, compound, drug or small molecule) which modulates protein activity (*e.g.*, identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on proliferative disorders, developmental or differentiative disorder, or hematopoietic disorder, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of

30 expression of the specified gene and other genes implicated in the proliferative disorder, developmental or differentiative disorder, or hematopoietic disorder, respectively. The levels of gene expression (*i.e.*, a gene expression pattern) can be quantified by Northern blot analysis or RT-PCR, as described herein, or alternatively

by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of the specified gene or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may
5 be determined before, and at various points during, treatment of the individual with the agent.

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

In one embodiment, the present invention provides a method for monitoring
10 the effectiveness of treatment of a subject with an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, polypeptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) comprising the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of a specified protein, mRNA, or genomic
15 DNA of the invention in the pre-administration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the protein, mRNA, or genomic DNA in the pre-administration sample with the protein, mRNA, or genomic DNA in
20 the post-administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of the protein or nucleic acid molecule to higher levels than detected, *i.e.*, to increase effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease
25 effectiveness of the agent. According to such an embodiment, protein or nucleic acid expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

VIII. Screening Assays

30 The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, antisense, polypeptides, peptidomimetics, small molecules or other drugs) which bind to nucleic acid molecules, polypeptides or proteins described herein or have a

stimulatory or inhibitory effect on, for example, expression or activity of the nucleic acid molecules, polypeptides or proteins of the invention.

As an example, apoptosis-specific assays may be used to identify modulators of any of the target nucleic acids or proteins of the present invention, which proteins
5 and/or nucleic acids are related to apoptosis. Accordingly, an agent that modulates the level or activity of any of these nucleic acids or proteins can be identified by means of apoptosis-specific assays. For example, high throughput screens exist to identify apoptotic cells by the use of chromatin or cytoplasmic-specific dyes. Thus, hallmarks of apoptosis, cytoplasmic condensation and chromosome fragmentation,
10 can be used as a marker to identify modulators of any of the genes related to programmed-cell death described herein. Other assays include, but are not limited to, the activation of specific endogenous proteases, loss of mitochondrial function, cytoskeletal disruption, cell shrinkage, membrane blebbing, and nuclear condensation due to degradation of DNA.

15 In one embodiment, the invention provides assays for screening candidate or test compounds that bind to or modulate the activity of protein or polypeptide described herein or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries;
20 spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to polypeptide libraries, while the other four approaches are applicable to polypeptide, non-peptide oligomer or small molecule libraries of
25 compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in DeWitt *et al.* (1993) *Proc. Natl. Acad. Sci. U.S.A.* 90:6909; Erb *et al.* (1994) *Proc. Natl. Acad. Sci. U.S.A.* 91:11422; Zuckermann *et al.* (1994). *J. Med. Chem.* 37:2678; Cho *et al.* (1993) *Science* 261:1303; Carell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.* 33:2059; Carell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.*
30 33:2061; and in Gallop *et al.* (1994) *J. Med. Chem.* 37:1233.

Libraries of compounds may be presented in solution (*e.g.*, Houghten (1992) *Biotechniques* 13:412-421), or on beads (Lam (1991) *Nature* 354:82-84), chips (Fodor

(1993) *Nature* 364:555-556), bacteria (Ladner U.S. Patent No. 5,223,409), spores (Ladner USP '409), plasmids (Cull *et al.* (1992) *Proc. Natl. Acad. Sci. U.S.A.* 89:1865-1869) or on phage (Scott and Smith (1990) *Science* 249:386-390); (Devlin (1990) *Science* 249:404-406); (Cwirla *et al.* (1990) *Proc. Natl. Acad. Sci.* 87:6378-6382); (Felici (1991) *J. Mol. Biol.* 222:301-310); (Ladner *supra*).

In one embodiment, an assay is a cell-based assay in which a cell that expresses an encoded polypeptide (*e.g.*, cell surface protein such as a receptor) is contacted with a test compound and the ability of the test compound to bind to the polypeptide is determined. The cell, for example, can be of mammalian origin, such as a keratinocyte. Determining the ability of the test compound to bind to the polypeptide can be accomplished, for example, by coupling the test compound with a radioisotope or enzymatic label such that binding of the test compound to the polypeptide can be determined by detecting the labeled with ^{125}I , ^{35}S , ^{14}C , or ^3H , either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, test compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a test compound to interact with the polypeptide without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of a test compound with the polypeptide without the labeling of either the test compound or the polypeptide. McConnell *et al.* (1992) *Science* 257:1906-1912. As used herein, a "microphysiometer" (*e.g.*, Cytosensor™) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between ligand and polypeptide.

In one embodiment, the assay comprises contacting a cell which expresses an encoded protein described herein on the cell surface (*e.g.*, a receptor) with a polypeptide ligand or biologically-active portion thereof, to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the polypeptide, wherein determining the ability of the test compound to interact with the polypeptide comprises determining the ability of

the test compound to preferentially bind to the polypeptide as compared to the ability of the ligand, or a biologically active portion thereof, to bind to the polypeptide.

In another embodiment, an assay is a cell-based assay comprising contacting a cell expressing a particular target molecule described herein with a test compound and
5 determining the ability of the test compound to modulate or alter (*e.g.* stimulate or inhibit) the activity of the target molecule. Determining the ability of the test compound to modulate the activity of the target molecule can be accomplished, for example, by determining the ability of a known ligand to bind to or interact with the target molecule. Determining the ability of the known ligand to bind to or interact
10 with the target molecule can be accomplished by one of the methods described above for determining direct binding. In one embodiment, determining the ability of the known ligand to bind to or interact with the target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (*e.g.*, intracellular Ca^{2+} , diacylglycerol, IP_3 , etc.), detecting
15 catalytic/enzymatic activity of the target an appropriate substrate, detecting the induction of a reporter gene (comprising a target-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase), or detecting a cellular response, for example, development, differentiation or rate of
20 proliferation.

In yet another embodiment, an assay of the present invention is a cell-free assay in which protein of the invention or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the protein or biologically active portion thereof is determined. Binding of the test
25 compound to the protein can be determined either directly or indirectly as described above. In one embodiment, the assay includes contacting the protein or biologically active portion thereof with a known compound which binds the protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the protein. Determining the ability of
30 the test compound to interact with the protein comprises determining the ability of the test compound to preferentially bind to the protein or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which a protein of the invention or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate or alter (*e.g.*, stimulate or inhibit) the activity of the protein or biologically active portion thereof is determined.

- 5 Determining the ability of the test compound to modulate the activity of the protein can be accomplished, for example, by determining the ability of the protein to bind to a known target molecule by one of the methods described above for determining direct binding. Determining the ability of the protein to bind to a target molecule can also be accomplished using a technology such as real-time Bimolecular Interaction
- 10 Analysis (BIA). Sjolander and Urbaniczky (1991) *Anal. Chem.* 63:2338-2345 and Szabo *et al.* (1995) *Curr. Opin. Struct. Biol.* 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (*e.g.*, BIAcore™). Changes in the optical phenomenon surface plasmon resonance (SPR) can be used as an indication of real-time reactions between
- 15 biological molecules.

- In an alternative embodiment, determining the ability of the test compound to modulate the activity of a protein of the invention can be accomplished by
- ~~determining the ability of the protein to further modulate the activity of a target~~
- 20 molecule. For example, the catalytic/enzymatic activity of the target molecule on an appropriate substrate can be determined as previously described.

- In yet another embodiment, the cell-free assay involves contacting a protein of the invention or biologically active portion thereof with a known compound which binds the protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the
- 25 protein, wherein determining the ability of the test compound to interact with the protein comprises determining the ability of the protein to preferentially bind to or modulate the activity of a target molecule.

- The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of isolated proteins. In the case of cell-free
- 30 assays in which a membrane-bound form an isolated protein is used it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the isolated protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside,

n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton®X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPS), 3-[(3-cholamidopropyl)dimethylamminio]-2-hydroxy-1-propane sulfonate (CHAPSO), or N-dodecyl-N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either the protein or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to the protein, or interaction of the protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or protein of the invention, and the mixture incubated under conditions conducive to complex formation (*e.g.*, at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either a protein of the invention or a target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated protein of the invention or target molecules can be prepared from biotin-NHS(N-hydroxy-succinimide) using techniques well known in the art (*e.g.*, biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with a protein of the invention or target molecules, but which do not interfere

with binding of the protein to its target molecule, can be derivatized to the wells of the plate, and unbound target or protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using
5 antibodies reactive with the protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the protein or target molecule.

In another embodiment, modulators of expression of nucleic acid molecules of the invention are identified in a method wherein a cell is contacted with a candidate
10 compound and the expression of appropriate mRNA or protein in the cell is determined. The level of expression of appropriate mRNA or protein in the presence of the candidate compound is compared to the level of expression of mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of expression based on this comparison. For example,
15 when expression of mRNA or protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator or enhancer of the mRNA or protein expression. Alternatively, when expression of the mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the
20 candidate compound is identified as an inhibitor of the mRNA or protein expression. The level of mRNA or protein expression in the cells can be determined by methods described herein for detecting mRNA or protein.

In yet another aspect of the invention, the proteins of the invention can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, *e.g.*, U.S. Patent
25 No. 5,283,317; Zervos *et al.* (1993) *Cell* 72:223-232; Madura *et al.* (1993) *J. Biol. Chem.* 268:12046-12054; Bartel *et al.* (1993) *Biotechniques* 14:920-924; Iwabuchi *et al.* (1993) *Oncogene* 8:1693-1696; and Brent WO94/10300), to identify other proteins (captured proteins) which bind to or interact with the proteins of the invention and modulate their activity. Such captured proteins are also likely to be involved in the
30 propagation of signals by the proteins of the invention as, for example, downstream elements of a protein-mediated signaling pathway. Alternatively, such captured proteins are likely to be cell-surface molecules associated with non-protein-expressing cells, wherein such captured proteins are involved in signal transduction.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (*e.g.*, a
5 modulating agent, an antisense nucleic acid molecule, a specific antibody, or a protein-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of
10 novel agents identified by the above-described screening assays for treatments as described herein.

IX. Methods of Treatment

The present invention provides for both prophylactic and therapeutic methods
15 of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant expression or activity of or related to proteins or nucleic acids of the invention. Methods of treatment involve modulating nucleic acid or polypeptide level or activity in a subject having a disorder that can be treated by such modulation. Accordingly, modulation can cause up regulation or down regulation of
20 the levels of expression or up regulation or down regulation of the activity of the nucleic acid or protein. Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

Expression of the nucleic acids of the invention has been shown for the following tissues: testes, brain, heart, kidney, skeletal muscle, spleen, lung, smooth
25 muscle, pancreas, and liver as shown in Figure 8. Accordingly, disorders to which the methods disclosed herein are particularly relevant include those involving these tissues.

Disorders involving the spleen include, but are not limited to, splenomegaly, including nonspecific acute splenitis, congestive splenomegaly, and splenic infarcts;
30 neoplasms, congenital anomalies, and rupture. Disorders associated with splenomegaly include infections, such as nonspecific splenitis, infectious mononucleosis, tuberculosis, typhoid fever, brucellosis, cytomegalovirus, syphilis, malaria, histoplasmosis, toxoplasmosis, kala-azar, trypanosomiasis, schistosomiasis,

leishmaniasis, and echinococcosis; congestive states related to partial hypertension, such as cirrhosis of the liver, portal or splenic vein thrombosis, and cardiac failure; lymphohematogenous disorders, such as Hodgkin disease, non-Hodgkin lymphomas/leukemia, multiple myeloma, myeloproliferative disorders, hemolytic anemias, and thrombocytopenic purpura; immunologic-inflammatory conditions, such as rheumatoid arthritis and systemic lupus erythematosus; storage diseases such as Gaucher disease, Niemann-Pick disease, and mucopolysaccharidoses; and other conditions, such as amyloidosis, primary neoplasms and cysts, and secondary neoplasms.

10 Disorders involving the lung include, but are not limited to, congenital anomalies; atelectasis; diseases of vascular origin, such as pulmonary congestion and edema, including hemodynamic pulmonary edema and edema caused by microvascular injury, adult respiratory distress syndrome (diffuse alveolar damage), pulmonary embolism, hemorrhage, and infarction, and pulmonary hypertension and
15 vascular sclerosis; chronic obstructive pulmonary disease, such as emphysema, chronic bronchitis, bronchial asthma, and bronchiectasis; diffuse interstitial (infiltrative, restrictive) diseases, such as pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia (pulmonary infiltration with eosinophilia),
20 *Bronchiolitis obliterans*-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, including Goodpasture syndrome, idiopathic pulmonary hemosiderosis and other hemorrhagic syndromes, pulmonary involvement in collagen vascular disorders, and pulmonary alveolar proteinosis; complications of therapies, such as drug-induced lung disease, radiation-induced lung disease, and lung transplantation;
25 tumors, such as bronchogenic carcinoma, including paraneoplastic syndromes, bronchioloalveolar carcinoma, neuroendocrine tumors, such as bronchial carcinoid, miscellaneous tumors, and metastatic tumors; pathologies of the pleura, including inflammatory pleural effusions, noninflammatory pleural effusions, pneumothorax, and pleural tumors, including solitary fibrous tumors (pleural fibroma) and malignant
30 mesothelioma.

Disorders involving the liver include, but are not limited to, hepatic injury; jaundice and cholestasis, such as bilirubin and bile formation; hepatic failure and cirrhosis, such as cirrhosis, portal hypertension, including ascites, portosystemic

shunts, and splenomegaly; infectious disorders, such as viral hepatitis, including hepatitis A-E infection and infection by other hepatitis viruses, clinicopathologic syndromes, such as the carrier state, asymptomatic infection, acute viral hepatitis, chronic viral hepatitis, and fulminant hepatitis; autoimmune hepatitis; drug- and toxin-induced liver disease, such as alcoholic liver disease; inborn errors of metabolism and pediatric liver disease, such as hemochromatosis, Wilson disease, α_1 -antitrypsin deficiency, and neonatal hepatitis; intrahepatic biliary tract disease, such as secondary biliary cirrhosis, primary biliary cirrhosis, primary sclerosing cholangitis, and anomalies of the biliary tree; circulatory disorders, such as impaired blood flow into the liver, including hepatic artery compromise and portal vein obstruction and thrombosis, impaired blood flow through the liver, including passive congestion and centrilobular necrosis and peliosis hepatis, hepatic vein outflow obstruction, including hepatic vein thrombosis (Budd-Chiari syndrome) and veno-occlusive disease; hepatic disease associated with pregnancy, such as preeclampsia and eclampsia, acute fatty liver of pregnancy, and intrahepatic cholestasis of pregnancy; hepatic complications of organ or bone marrow transplantation, such as drug toxicity after bone marrow transplantation, graft-versus-host disease and liver rejection, and nonimmunologic damage to liver allografts; tumors and tumorous conditions, such as nodular hyperplasias, adenomas, and malignant tumors, including primary carcinoma of the liver metastatic tumors, and liver fibrosis.

Disorders involving the brain include, but are not limited to, disorders involving neurons, and disorders involving glia, such as astrocytes, oligodendrocytes, ependymal cells, and microglia; cerebral edema, raised intracranial pressure and herniation, and hydrocephalus; malformations and developmental diseases, such as neural tube defects, forebrain anomalies, posterior fossa anomalies, and syringomyelia and hydromyelia; perinatal brain injury; cerebrovascular diseases, such as those related to hypoxia, ischemia, and infarction, including hypotension, hypoperfusion, and low-flow states--global cerebral ischemia and focal cerebral ischemia--infarction from obstruction of local blood supply, intracranial hemorrhage, including intracerebral (intraparenchymal) hemorrhage, subarachnoid hemorrhage and ruptured berry aneurysms, and vascular malformations, hypertensive cerebrovascular disease, including lacunar infarcts, slit hemorrhages, and hypertensive encephalopathy; infections, such as acute meningitis, including acute pyogenic (bacterial) meningitis

and acute aseptic (viral) meningitis, acute focal suppurative infections, including brain abscess, subdural empyema, and extradural abscess, chronic bacterial meningoencephalitis, including tuberculosis and mycobacterioses, neurosyphilis, and neuroborreliosis (Lyme disease), viral meningoencephalitis, including arthropod-

5 borne (Arbo) viral encephalitis, *Herpes simplex* virus Type 1, *Herpes simplex* virus Type 2, *Varicella-zoster* virus (*Herpes zoster*), cytomegalovirus, poliomyelitis, rabies, and human immunodeficiency virus 1, including HIV-1 meningoencephalitis (subacute encephalitis), vacuolar myelopathy, AIDS-associated myopathy, peripheral neuropathy, and AIDS in children, progressive multifocal leukoencephalopathy,

10 subacute sclerosing panencephalitis, fungal meningoencephalitis, other infectious diseases of the nervous system; transmissible spongiform encephalopathies (prion diseases); demyelinating diseases, including multiple sclerosis, multiple sclerosis variants, acute disseminated encephalomyelitis and acute necrotizing hemorrhagic encephalomyelitis, and other diseases with demyelination; degenerative diseases, such

15 as degenerative diseases affecting the cerebral cortex, including Alzheimer disease and Pick disease, degenerative diseases of basal ganglia and brain stem, including Parkinsonism, idiopathic Parkinson disease (paralysis agitans), progressive supranuclear palsy; corticobasal degeneration; multiple system atrophy, including striatonigral degeneration, Shy-Drager syndrome, and olivopontocerebellar atrophy,

20 and Huntington disease; spinocerebellar degenerations, including spinocerebellar ataxias, including Friedreich ataxia, and ataxia-telangiectasia, degenerative diseases affecting motor neurons, including amyotrophic lateral sclerosis (motor neuron disease), bulbospinal atrophy (Kennedy syndrome), and spinal muscular atrophy; inborn errors of metabolism, such as leukodystrophies, including Krabbe disease,

25 metachromatic leukodystrophy, adrenoleukodystrophy, Pelizaeus-Merzbacher disease, and Canavan disease, mitochondrial encephalomyopathies, including Leigh disease and other mitochondrial encephalomyopathies; toxic and acquired metabolic diseases, including vitamin deficiencies such as thiamine (vitamin B₁) deficiency and vitamin B₁₂ deficiency, neurologic sequelae of metabolic disturbances, including

30 hypoglycemia, hyperglycemia, and hepatic encephatopathy, toxic disorders, including carbon monoxide, methanol, ethanol, and radiation, including combined methotrexate and radiation-induced injury; tumors, such as gliomas, including astrocytoma, including fibrillary (diffuse) astrocytoma and glioblastoma multiforme, pilocytic

astrocytoma, pleomorphic xanthoastrocytoma, and brain stem glioma, oligodendroglioma, and ependymoma and related paraventricular mass lesions, neuronal tumors, poorly differentiated neoplasms, including medulloblastoma, other parenchymal tumors, including primary brain lymphoma, germ cell tumors, and
5 pineal parenchymal tumors, meningiomas, metastatic tumors, paraneoplastic syndromes, peripheral nerve sheath tumors, including schwannoma, neurofibroma, and malignant peripheral nerve sheath tumor (malignant schwannoma), and neurocutaneous syndromes (phakomatoses), including neurofibromatosis, including Type 1 neurofibromatosis (NF1) and TYPE 2 neurofibromatosis (NF2), tuberous
10 sclerosis, and Von Hippel-Lindau disease.

Disorders involving the heart, include but are not limited to, heart failure, including but not limited to, cardiac hypertrophy, left-sided heart failure, and right-sided heart failure; ischemic heart disease, including but not limited to angina pectoris, myocardial infarction, chronic ischemic heart disease, and sudden cardiac
15 death; hypertensive heart disease, including but not limited to, systemic (left-sided) hypertensive heart disease and pulmonary (right-sided) hypertensive heart disease; valvular heart disease, including but not limited to, valvular degeneration caused by calcification, such as calcific aortic stenosis, calcification of a congenitally bicuspid aortic valve, and mitral annular calcification, and myxomatous degeneration of the
20 mitral valve (mitral valve prolapse), rheumatic fever and rheumatic heart disease, infective endocarditis, and noninfected vegetations, such as nonbacterial thrombotic endocarditis and endocarditis of systemic lupus erythematosus (Libman-Sacks disease), carcinoid heart disease, and complications of artificial valves; myocardial disease, including but not limited to dilated cardiomyopathy, hypertrophic
25 cardiomyopathy, restrictive cardiomyopathy, and myocarditis; pericardial disease, including but not limited to, pericardial effusion and hemopericardium and pericarditis, including acute pericarditis and healed pericarditis, and rheumatoid heart disease; neoplastic heart disease, including but not limited to, primary cardiac tumors, such as myxoma, lipoma, papillary fibroelastoma, rhabdomyoma, and sarcoma, and
30 cardiac effects of noncardiac neoplasms; congenital heart disease, including but not limited to, left-to-right shunts--late cyanosis, such as atrial septal defect, ventricular septal defect, patent ductus arteriosus, and atrioventricular septal defect, right-to-left shunts--early cyanosis, such as tetralogy of fallot, transposition of great arteries,

truncus arteriosus, tricuspid atresia, and total anomalous pulmonary venous connection, obstructive congenital anomalies, such as coarctation of aorta, pulmonary stenosis and atresia, and aortic stenosis and atresia, and disorders involving cardiac transplantation.

- 5 Disorders involving the kidney include, but are not limited to, congenital anomalies including, but not limited to, cystic diseases of the kidney, that include but are not limited to, cystic renal dysplasia, autosomal dominant (adult) polycystic kidney disease, autosomal recessive (childhood) polycystic kidney disease, and cystic diseases of renal medulla, which include, but are not limited to, medullary sponge kidney, and
- 10 nephronophthisis-uremic medullary cystic disease complex, acquired (dialysis-associated) cystic disease, such as simple cysts; glomerular diseases including pathologies of glomerular injury that include, but are not limited to, in situ immune complex deposition, that includes, but is not limited to, anti-GBM nephritis, Heymann nephritis, and antibodies against planted antigens, circulating immune complex nephritis,
- 15 antibodies to glomerular cells, cell-mediated immunity in glomerulonephritis, activation of alternative complement pathway, epithelial cell injury, and pathologies involving mediators of glomerular injury including cellular and soluble mediators, acute
- glomerulonephritis, such as acute proliferative (poststreptococcal, postinfectious) glomerulonephritis, including but not limited to, poststreptococcal glomerulonephritis
- 20 and nonstreptococcal acute glomerulonephritis, rapidly progressive (crescentic) glomerulonephritis, nephrotic syndrome, membranous glomerulonephritis (membranous nephropathy), minimal change disease (lipoid nephrosis), focal segmental glomerulosclerosis, membranoproliferative glomerulonephritis, IgA nephropathy (Berger disease), focal proliferative and necrotizing glomerulonephritis (focal
- 25 glomerulonephritis), hereditary nephritis, including but not limited to, Alport syndrome and thin membrane disease (benign familial hematuria), chronic glomerulonephritis, glomerular lesions associated with systemic disease, including but not limited to, systemic lupus erythematosus, Henoch-Schönlein purpura, bacterial endocarditis, diabetic glomerulosclerosis, amyloidosis, fibrillary and immunotactoid
- 30 glomerulonephritis, and other systemic disorders; diseases affecting tubules and interstitium, including acute tubular necrosis and tubulointerstitial nephritis, including but not limited to, pyelonephritis and urinary tract infection, acute pyelonephritis, chronic pyelonephritis and reflux nephropathy, and tubulointerstitial nephritis induced

by drugs and toxins, including but not limited to, acute drug-induced interstitial nephritis, analgesic abuse nephropathy, nephropathy associated with nonsteroidal anti-inflammatory drugs, and other tubulointerstitial diseases including, but not limited to, urate nephropathy, hypercalcemia and nephrocalcinosis, and multiple myeloma; diseases
5 of blood vessels including benign nephrosclerosis, malignant hypertension and accelerated nephrosclerosis, renal artery stenosis, and thrombotic microangiopathies including, but not limited to, classic (childhood) hemolytic-uremic syndrome, adult hemolytic-uremic syndrome/thrombotic thrombocytopenic purpura, idiopathic HUS/TTP, and other vascular disorders including, but not limited to, atherosclerotic
10 ischemic renal disease, atheroembolic renal disease, sickle cell disease nephropathy, diffuse cortical necrosis, and renal infarcts; urinary tract obstruction (obstructive uropathy); urolithiasis (renal calculi, stones); and tumors of the kidney including, but not limited to, benign tumors, such as renal papillary adenoma, renal fibroma or hamartoma (renomedullary interstitial cell tumor), angiomyolipoma, and oncocytoma, and malignant
15 tumors, including renal cell carcinoma (hypernephroma, adenocarcinoma of kidney), which includes urothelial carcinomas of renal pelvis.

Disorders involving the testis and epididymis include, but are not limited to, congenital anomalies such as cryptorchidism, regressive changes such as atrophy, inflammations such as nonspecific epididymitis and orchitis, granulomatous
20 (autoimmune) orchitis, and specific inflammations including, but not limited to, gonorrhea, mumps, tuberculosis, and syphilis, vascular disturbances including torsion, testicular tumors including germ cell tumors that include, but are not limited to, seminoma, spermatocytic seminoma, embryonal carcinoma, yolk sac tumor choriocarcinoma, teratoma, and mixed tumors, tumors of sex cord-gonadal stroma
25 including, but not limited to, leydig (interstitial) cell tumors and sertoli cell tumors (androblastoma), and testicular lymphoma, and miscellaneous lesions of tunica vaginalis.

Disorders involving the skeletal muscle include tumors such as rhabdomyosarcoma.

Disorders involving the pancreas include those of the exocrine pancreas such as
30 congenital anomalies, including but not limited to, ectopic pancreas; pancreatitis, including but not limited to, acute pancreatitis; cysts, including but not limited to, pseudocysts; tumors, including but not limited to, cystic tumors and carcinoma of the pancreas; and disorders of the endocrine pancreas such as, diabetes mellitus; islet cell

tumors, including but not limited to, insulinomas, gastrinomas, and other rare islet cell tumors.

Preferred disorders include those involving the central nervous system and particularly the brain.

- 5 With regard to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics. "Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market.
- 10 More specifically, the term refers the study of how a patient's genes determine his or her response to a drug (*e.g.*, a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with the molecules of the present invention or modulators according to that individual's drug response genotype.
- 15 Pharmacogenomics allows a clinician or physician to target prophylactic or therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug related side effects.

1. Prophylactic Methods

- 20 In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with aberrant expression or activity of genes or proteins of the present invention, by administering to the subject an agent which modulates expression or at least one activity of a gene or protein of the invention. Subjects at risk for a disease that is caused or contributed to by aberrant gene
- 25 expression or protein activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of aberrancy, for example, an agonist or
- 30 antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating expression or activity of genes or proteins of the invention for therapeutic purposes. The modulatory method of the invention involves contacting a cell with an agent that modulates one or more of the activities of the specified protein associated with the cell. An agent that modulates protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of a protein described herein, a polypeptide, a peptidomimetic, or other small molecule. In one embodiment, the agent stimulates one or more protein activities. Examples of such stimulatory agents include active protein as well as a nucleic acid molecule encoding the protein that has been introduced into the cell. In another embodiment, the agent inhibits one or more protein activities. Examples of such inhibitory agents include antisense nucleic acid molecules and anti-protein antibodies. These modulatory methods can be performed *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of a protein or nucleic acid molecule of the invention. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) expression or activity of a gene or protein of the invention. In another embodiment, the method involves administering a protein or nucleic acid molecule of the invention as therapy to compensate for reduced or aberrant expression or activity of the protein or nucleic acid molecule.

Stimulation of protein activity is desirable in situations in which the protein is abnormally downregulated and/or in which increased protein activity is likely to have a beneficial effect. Likewise, inhibition of protein activity is desirable in situations in which the protein is abnormally upregulated and/or in which decreased protein activity is likely to have a beneficial effect. One example of such a situation is where a subject has a disorder characterized by aberrant development or cellular differentiation. Another example of such a situation is where the subject has a proliferative disease (e.g., cancer) or a disorder characterized by an aberrant

hematopoietic response. Yet another example of such a situation is where it is desirable to achieve tissue regeneration in a subject (*e.g.*, where a subject has undergone brain or spinal cord injury and it is desirable to regenerate neuronal tissue in a regulated manner).

5

Pharmaceutical Compositions

The nucleic acid molecules, protein modulators of the protein, and antibodies (also referred to herein as "active compounds") can be incorporated into pharmaceutical compositions suitable for administration to a subject, *e.g.*, a human. Such compositions typically comprise the nucleic acid molecule, protein, modulator, or antibody and a pharmaceutically acceptable carrier.

The term "administer" is used in its broadest sense and includes any method of introducing the compositions of the present invention into a subject. This includes producing polypeptides or polynucleotides *in vivo* as by transcription or translation, *in vivo*, of polynucleotides that have been exogenously introduced into a subject. Thus, polypeptides or nucleic acids produced in the subject from the exogenous compositions are encompassed in the term "administer."

As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, such media can be used in the compositions of the invention. Supplementary active compounds can also be incorporated into the compositions. A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as

ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampules, disposable syringes or multiple dose
5 vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water,
10 Cremophor EL™ (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for
15 example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and
20 antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example,
25 aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a ubiquitin protease protein or anti- ubiquitin protease antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions
30 are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields

a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For oral administration, the agent can be contained in enteric forms to survive the stomach or further coated or mixed to be released in a particular region of the GI tract by known methods. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser, which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. "Dosage unit form" as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (U.S. 5,328,470) or by stereotactic injection (see e.g., Chen *et al.* (1994) *PNAS* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, e.g. retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

As defined herein, a therapeutically effective amount of protein or polypeptide (i.e., an effective dosage) ranges from about 0.001 to 30 mg/kg body weight,

preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight.

The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments. In a preferred example, a subject is treated with antibody, protein, or polypeptide in the range of between about 0.1 to 20 mg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of antibody, protein, or polypeptide used for treatment may increase or decrease over the course of a particular treatment. Changes in dosage may result and become apparent from the results of diagnostic assays as described herein.

The present invention encompasses agents which modulate expression or activity. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (i.e., including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds.

It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon the identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if

applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention. Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein. When one or more of these small molecules is to be administered to an animal (e.g., a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, gender, and diet of the subject, the time of administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

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3. Pharmacogenomics

The molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on the protein activity (e.g., gene expression) as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) disorders (e.g., proliferative or developmental disorders) associated with aberrant protein activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer a molecule of the invention or modulator thereof,

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as well as tailoring the dosage and/or therapeutic regimen of treatment with such a molecule or modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See *e.g.*, Eichelbaum (1996) *Clin Exp. Pharmacol. Physiol.* 23(10-11):983-985 and Linder (1997) *Clin. Chem.* 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (*e.g.*, a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants). Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a "SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1,000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict drug response. According to this method, if a gene that encodes a drug's target is known (*e.g.*, a protein or a polypeptide of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (*e.g.*, N-acetyltransferase 2(NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme is the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (*e.g.*, a molecule or modulator of the present invention) can given an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with a

molecule or modulator of the invention, such as a modulator identified by one of the exemplary screening assays described herein.

Disorders which may be treated or diagnosed by methods described herein include, but are not limited to disorders involving apoptosis. Certain disorders are associated with an increased number of surviving cells, which are produced and continue to survive or proliferate when apoptosis is inhibited.

As used herein, "programmed cell death" refers to a genetically regulated process involved in the normal development of multicellular organisms. This process occurs in cells destined for removal in a variety of normal situations, including larval development of the nematode *C. elegans*, insect metamorphosis, development in mammalian embryos, including the nephrogenic zone in the developing kidney, and regression or atrophy (e.g., in the prostate after castration). Programmed cell death can occur following the withdrawal of growth and trophic factors in many cells, nutritional deprivation, hormone treatment, ultraviolet irradiation, and exposure to toxic and infectious agents including reactive oxygen species and phosphatase inhibitors, e.g., okadaic acid, calcium ionophores, and a number of cancer chemotherapeutic agents. See Wilson (1998) *Biochem. Cell Biol.* 76:573-582 and Hetts (1998) *JAMA* 279:300-307, the contents of which are incorporated herein by reference. Thus, the proteins of the invention, by being differentially expressed during programmed cell death, e.g., neuronal programmed cell death, can modulate a programmed cell death pathway activity and provide novel diagnostic targets and therapeutic agents for disorders characterized by deregulated programmed cell death, particularly in cells that express the protein.

As used herein, a "disorder characterized by deregulated programmed cell death" refers to a disorder, disease or condition which is characterized by a deregulation, e.g., an upregulation or a downregulation, of programmed cell death. Programmed cell death deregulation can lead to deregulation of cellular proliferation and/or cell cycle progression. Examples of disorders characterized by deregulated programmed cell death include, but are not limited to, neurodegenerative disorders, e.g., Alzheimer's disease, dementias related to Alzheimer's disease (such as Pick's disease), Parkinson's and other Lewy diffuse body diseases, multiple sclerosis, amyotrophic lateral sclerosis, progressive supranuclear palsy, epilepsy, Jakob-Creutzfeldt disease, or AIDS related dementias; myelodysplastic syndromes, e.g.,

aplastic anemia; ischemic injury, e.g., myocardial infarction, stroke, or reperfusion injury; autoimmune disorders, e.g., systemic lupus erythematosus, or immune-mediated glomerulonephritis; or proliferative disorders, e.g., cancer, such as follicular lymphomas, carcinomas with p53 mutations, or hormone-dependent tumors, e.g.,
5 breast cancer, prostate cancer, or ovarian cancer). Clinical manifestations of faulty apoptosis are also seen in stroke and in rheumatoid arthritis. Wilson (1998) *Biochem. Cell. Biol.* 76:573-582.

Failure to remove autoimmune cells that arise during development or that develop as a result of somatic mutation during an immune response can result in
10 autoimmune disease. One of the molecules that plays a critical role in regulating cell death in lymphocytes is the cell surface receptor for Fas.

Viral infections, such as those caused by herpesviruses, poxviruses, and adenoviruses, may result in aberrant apoptosis. Populations of cells are often depleted in the event of viral infection, with perhaps the most dramatic example being the cell
15 depletion caused by the human immunodeficiency virus (HIV). Most T cells that die during HIV infections do not appear to be infected with HIV. Stimulation of the CD4 receptor may result in the enhanced susceptibility of uninfected T cells to undergo apoptosis.

Many disorders can be classified based on whether they are associated with
20 abnormally high or abnormally low apoptosis. Thompson (1995) *Science* 267:1456-1462. Apoptosis may be involved in acute trauma, myocardial infarction, stroke, and infectious diseases, such as viral hepatitis and acquired immunodeficiency syndrome.

Primary apoptosis deficiencies include graft rejection. Accordingly, the invention is relevant to the identification of genes useful in inhibiting graft rejection.

25 Primary apoptosis deficiencies also include autoimmune diabetes. Accordingly, the invention is relevant to the identification of genes involved in autoimmune diabetes and accordingly, to the identification of agents that act on these targets to modulate the expression of these genes and hence, to treat or diagnose this disorder. Further, it has been suggested that all autoimmune disorders can be viewed
30 as primary deficiencies of apoptosis (Hetts, above). Accordingly, the invention is relevant for screening for gene expression and transcriptional profiling in any autoimmune disorder and for screening for agents that affect the expression or transcriptional profile of these genes.

Primary apoptosis deficiencies also include local self reactive disorder. This includes Hashimoto thyroiditis.

Primary apoptosis deficiencies also include lymphoproliferation and autoimmunity. This includes, but is not limited to, Canale-Smith syndrome.

- 5 Primary apoptosis deficiencies also include cancer. For example, p53 induces apoptosis by acting as a transcription factor that activates expression of various apoptosis-mediating genes or by upregulating apoptosis-mediating genes such as Bax.

- 10 Primary apoptosis excesses are associated with neurodegenerative disorders including Alzheimer's disease, Parkinson's disease, spinal muscular atrophy, and amyotrophic lateral sclerosis.

- 15 Primary apoptosis excesses are also associated with heart disease including idiopathic dilated cardiomyopathy, ischemic cardiomyopathy, and valvular heart disease. Evidence has also been shown of apoptosis in heart failure resulting from arrhythmogenic right ventricular dysplasia. For all these disorders, see Hetts, above.

- 20 Death receptors also include the TNF receptor-1 and hence, TNF acts as a death ligand.

- 25 A wide variety of neurological diseases are characterized by the gradual loss of specific sets of neurons. Such disorders include Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis (ALS) retinitis pigmentosa, spinal muscular atrophy, and various forms of cerebellar degeneration. The cell loss in these diseases does not induce an inflammatory response, and apoptosis appears to be the mechanism of cell death.

- 30 In addition, a number of hematologic diseases are associated with a decreased production of blood cells. These disorders include anemia associated with chronic disease, aplastic anemia, chronic neutropenia, and the myelodysplastic syndromes. Disorders of blood cell production, such as myelodysplastic syndrome and some forms of aplastic anemia, are associated with increased apoptotic cell death within the bone marrow.

- 35 These disorders could result from the activation of genes that promote apoptosis, acquired deficiencies in stromal cells or hematopoietic survival factors, or the direct effects of toxins and mediators of immune responses.

Two common disorders associated with cell death are myocardial infarctions and stroke. In both disorders, cells within the central area of ischemia, which is

produced in the event of acute loss of blood flow, appear to die rapidly as a result of necrosis. However, outside the central ischemic zone, cells die over a more protracted time period and morphologically appear to die by apoptosis.

The invention also pertains to disorders of the central nervous system (CNS).

5 These disorders include, but are not limited to cognitive and neurodegenerative disorders such as Alzheimer's disease, senile dementia, Huntington's disease, amyotrophic lateral sclerosis, and Parkinson's disease, as well as Gilles de la Tourette's syndrome, autonomic function disorders such as hypertension and sleep disorders, and neuropsychiatric disorders that include, but are not limited to
10 schizophrenia, schizoaffective disorder, attention deficit disorder, dysthymic disorder, major depressive disorder, mania, obsessive-compulsive disorder, psychoactive substance use disorders, anxiety, panic disorder, as well as bipolar affective disorder, e.g., severe bipolar affective (mood) disorder (BP-I), bipolar affective (mood) disorder with hypomania and major depression (BP-II). Further CNS-related
15 disorders include, for example, those listed in the American Psychiatric Association's Diagnostic and Statistical manual of Mental Disorders (DSM), the most current
— version of which is incorporated herein by reference in its entirety.

As used herein, "differential expression" or differentially expressed" includes both quantative and qualitative differences in the temporal and/or cellular expression
20 pattern of a gene, e.g., the programmed cell death genes disclosed herein, among, for example, normal cells and cells undergoing programmed cell death. Genes which are differentially expressed can be used as part of a prognostic or diagnostic marker for the evaluation of subjects at risk for developing a disorder characterized by deregulated programmed cell death. Depending on the expression level of the gene,
25 the progression state of the disorder can also be evaluated.

X. Arrays and Microarrays

The term "array" refers to a set of nucleic acid sequences that comprise at least one of SEQ ID NOS: 1-6, 8, and 10. Preferred arrays contain numerous genes. The
30 term can refer to all of the sequences in SEQ ID NOS: 1-6, 8, and 10 but could also include additional sequences, for example, sequences included as controls for specific biological processes. A "subarray" is also an array but is obtained by creating an array of less than all of the sequences in a starting array.

In one embodiment of the invention, the functional subarray comprises nucleic acid sequences expressed in programmed cell death as disclosed herein.

The array comprises not only the specific designated sequences but also variants of these sequences, as described herein. As described, variants include, 5 allelic variants, homologs from other loci in the same animal, orthologs, and sequences sufficiently similar such that they fulfill the requisites for sequence similarity/homology as described herein.

Further, the array not only comprises the specific designated sequences, but also comprises fragments thereof. As described herein, the range of fragments will 10 vary depending upon the specific sequence involved. Accordingly, the range of fragments is considerable, for example, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 125, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 etc. In no way, however, is a fragment to be construed as having a sequence identical to that which may be found in the prior art.

15 The array can be used to assay expression of one or more genes in the array.

In one embodiment, the array can be used to assay gene expression in a tissue to ascertain tissue specificity of genes in the array.

In addition to such qualitative determination, the invention allows the quantitation of gene expression. Thus, not only tissue specificity, but also the level of 20 expression of a battery of genes in the tissue is ascertainable. Thus, genes can be grouped on the basis of their tissue expression *per se* and level of expression in that tissue. This is useful, for example, in ascertaining the relationship of gene expression between or among tissues. Thus, one tissue can be perturbed and the effect on gene expression in a second tissue can be determined. In this context, the effect of one cell 25 type on another cell type in response to a biological stimulus can be determined. Such a determination is useful, for example, to know the effect of cell-cell interaction at the level of gene expression. If an agent is administered therapeutically to treat one cell type but has an undesirable effect on another cell type, the invention provides an assay to determine the molecular basis of the undesirable effect and thus provides the 30 opportunity to co-administer a counteracting agent or otherwise treat the undesired effect. Similarly, even within a single cell type, undesirable biological effects can be determined at the molecular level. Thus, the effects of an agent on expression of other than the target gene can be ascertained and counteracted.

In another embodiment, the array can be used to monitor the time course of expression of one or more genes in the array. This can occur in various biological contexts, as disclosed herein, for example development and differentiation, tumor progression, progression of other diseases, *in vitro* processes, such as cellular transformation and senescence, autonomic neural and neurological processes, such as, for example, pain and appetite, and cognitive functions, such as learning or memory.

The array is also useful for ascertaining the effect of the expression of a gene on the expression of other genes in the same cell or in different cells. This provides, for example, for a selection of alternate molecular targets for therapeutic intervention if the ultimate or downstream target cannot be regulated.

The array is also useful for ascertaining differential expression patterns of one or more genes in normal and abnormal cells. This provides a battery of genes that could serve as a molecular target for diagnosis or therapeutic intervention.

In one embodiment, the array, and particularly subarrays containing one or more of the nucleic acid sequences related to programmed cell death, are useful for diagnosing disease or predisposition to disease involving apoptosis. These disorders include, but are not limited to, those discussed in detail herein. In addition, the array or subarrays created therefrom are useful for diagnosing active disorders of the central nervous system or for predicting the tendency to develop such disorders. Disorders of the central nervous system include, but are not limited to, those disclosed in detail herein. Furthermore, the array and subarrays thereof are useful for diagnosing an active disorder or predicting the tendency to develop a disorder including, but not limited to, disorders involving secretion/synaptic vesicle release, cell proliferation, cytoskeletal reorganization, stress response/hormone response; and calcium signal transduction.

The array is also useful for ascertaining expression of one or more genes in model systems *in vitro* or *in vivo*. Various model systems have been developed to study normal and abnormal processes, including, but not limited to, apoptosis.

Apoptosis can be actively induced in animal cells by a diverse array of triggers that range from ionizing radiation to hypothermia to viral infections to immune reactions. Majno *et al.* (1995) *Amer. J. Pathol.* 146:3-15; Hockenberry *et al.* (1995) *Bio Essays* 17:631-638; Thompson *et al.* *Science* 267:1456-1462 (1995).

Transgenic mouse models have been developed for familial amyotrophic lateral sclerosis, familial Alzheimer's disease and Huntington's disease, reviewed in Price *et al.* (1998) *Science* 282:1079-1083. Amyotrophic lateral sclerosis is the most common adult onset motor neuron disease. Alzheimer's disease is the most common cause of dementia in adult life. It is associated with the damage of regions and neurocircuits critical for cognition and memory, including neurons in the neocortex, hippocampus, amygdala, basal forebrain cholinergic system, and brain stem monoaminergic nuclei. Neurological diseases that are associated with autosomal dominant trinucleotide repeat mutations include Huntington's disease, several spinal cerebellar ataxias and dentatorubral pallidoluysian atrophy. SCA-1 and SCA-3 or Machado-Joseph disease are characterized by ataxia and lack of coordination. In Huntington's disease, symptoms are related to degeneration of subsets of striatal and cortical neurons. Apoptosis is thought to play a role in the degeneration of these cells. In SCA-1, SCA-3, and in dentatorubral pallidoluysian atrophy, a variety of cell populations, and particularly cells in the cerebellum, have been shown to degenerate. See Price *et al.* above, which is incorporated by reference in its entirety for the teachings of model systems related to neurodegenerative diseases.

----- Mouse models have been developed for non-obese diabetic mice, to study disease progression for the treatment of autoimmune diabetes mellitus. Bellgrau *et al.* (1995) *Nature* 377:630-632. Models have also been developed in mice wherein the mice lack one or two copies of the p53 gene. Study of these mice has shown that apoptosis is involved in suppressing tumor development in vivo. Lozano *et al.* (1998) *Semin. Canc. Biol.* 8:337-344. Another animal model relevant to the study of apoptosis involves the targeted gene disruption of caspase genes creating caspase gene knockout mice. Colussi *et al.* (1999) *J. Immun. Cell. Biol.* 77:58-63. A further mouse model pertains to cold injury in mice, such injury inducing neuronal apoptosis. Murakami *et al.* (1999) *Prog. Neurobiol.* 57:289-299.

Knockout mice have been created for Apaf1. In these mice, defects are found in essentially all tissues whose development depends on cell death, including loss of interdigital webs, formation of the palate, control of neuron cell number, and development of the lens and retina. Cecconi *et al.* (1998) *Cell* 94:727-737.

Caspase knockout mice have also been achieved for caspase 1, 2, 3, and 9. Green (1998) *Cell* 94:695-698.

The array allows the simultaneous determination of a battery of genes involved in these processes and thus provides multiple candidates for *in vivo* verification and clinical testing. Because the array allows the determination of expression of multiple genes; it provides a powerful tool to ascertain coordinate gene expression, that is co-expression of two or more genes in a time and/or tissue-specific manner, both qualitatively and quantitatively. Thus, genes can be grouped on the basis of their expression *per se* and/or level of expression. This allows the classification of genes into functional categories even when the gene is completely uncharacterized with respect to function. Accordingly, if a first gene is expressed coordinately with a second gene whose function is known, a putative function can be assigned to that first gene. This first gene thus provides a new target for affecting that function in a diagnostic or therapeutic context. The larger the number of genes in an array, the greater is the probability that numerous known genes having the same or similar function will be expressed. In this case, the coordinate expression of one or more novel genes (with respect to function and/or structure) strongly allows discovery of genes in the same functional category as the known genes.

Accordingly, the array of the invention provides for "internal control" groups of genes whose functions are known and can thus be used to identify genes as being in the same functional category of the control group if they are coordinated expressed.

As an alternative to relying on such internal control groups, external control groups can be added to the array. The genes in such a group would have a known function. Genes coordinately expressed with these genes would thus be *prima facie* involved in the same function.

Therefore, the array provides a method not only for discovering novel genes having a specific function but also for assigning function to genes whose function is unknown or assigning to a known gene an additional function, previously unknown for that gene.

Accordingly, as disclosed and exemplified herein, previously characterized genes were grouped into new functional categories (i.e., previously the function was not known to be possessed by that gene). Furthermore, several uncharacterized genes could be functionally classified on the basis of coordinate expression with the "internal control group of genes". In a specific embodiment, disclosed and exemplified herein, genes related to programmed cell death in brain were selected.

The array could, accordingly be used to select for genes related to other important biological processes, such as those disclosed herein. Nucleic acid from any tissue in any biological process is hybridized to nucleic acid sequences in an array. The expression pattern of genes in the array allows for their classification into functional groups based on specific expression patterns. Internal or external control genes (i.e. genes known to be expressed in the specific tissue/biological process) provide verification to classify other genes in the specific category.

Just as the array was useful for identifying programmed cell death genes, other relevant normal biological models include differentiation programs and disorders such as those disclosed herein.

The array is also useful for drug discovery. Candidate compounds can be used to screen cells and tissues in any of the biological contexts disclosed herein, such as pathology, development, differentiation, etc. Thus the expression of one or more genes in the array can be monitored by using the array to screen for RNA expression in a cell or tissue exposed to a candidate compound. Compounds can be selected on the basis of the overall effect on gene expression, not necessarily on the basis of its effect on a single gene. Thus, for example, where a compound is desired that affects a particular first gene or genes but has no effect on a second gene or genes, the array provides a way to globally monitor the effect on gene expression of a compound.

Alternatively, it may be desirable to target more than one gene, i.e. to modulate the expression of more than one gene. The array provides a way to discover compounds that will modulate a set of genes. All genes of the set can be upregulated or downregulated. Alternatively, some of the genes may be upregulated and others downregulated by the same compound. Moreover, compounds are discoverable that modulate desired genes to desired degrees.

In the context of drug discovery, functional subarrays of genes are especially useful. Thus, using the methods disclosed herein and those routinely available, groups of genes can be assembled based on their relationships to a specific biological function. The expression of this group of genes can be used for diagnostic purposes and to discover compounds relevant to the biological function. Thus, the subarray can provide the basis for discovering drugs relevant to treatment and diagnosis of disease, for example those disclosed herein.

In the present case, the group of genes whose expression is correlated with programmed cell death can be used to discover compounds that affect programmed cell death, and especially disorders in which programmed cell death is involved. These include but are not limited to those disclosed herein.

5 Apoptosis can be triggered by the addition of apoptosis-promoting ligands to a cell in culture or in vivo. In one embodiment of the invention, therefore, the arrays and subarrays described herein are useful to identify genes that respond to apoptosis-promoting ligands and conversely to identify ligands that act on genes involved in apoptosis. Apoptosis can also be triggered by decreasing or removing an apoptosis-inhibiting or survival-promoting ligand. Accordingly, apoptosis is triggered in view
10 of the fact that the cell lacks a signal from a cell surface survival factor receptor. Ligands include, but are not limited to, FasL. Death-inhibiting ligands include, but are not limited to, IL-2. See Hetts *et al.* (1998) *JAMA* 279:300-307 (incorporated by reference in its entirety for teaching of ligands involved in active and passive
15 apoptosis pathways.) Central in the pathway, and also serving as potential molecules for inducing (or releasing from inhibition) apoptosis pathways include FADD, caspases, human CED4 homolog (also called apoptotic protease activating factor 1), the Bcl-2 family of genes including, but not limited to, apoptosis promoting (for example, Bax and Bad) and apoptosis inhibiting (for example, Bcl-2 and Bcl-x_l)
20 molecules. See Hetts *et al.*, above.

Multiple caspases upstream of caspase-3 can be inhibited by viral proteins such as cowpox, CrmA, and baculovirus, p35, synthetic tripeptides and tetrapeptides inhibit caspase-3 specifically (Hetts, above). Accordingly, the arrays and subarrays are useful for determining the modulation of gene expression in response to these
25 agents.

The array is also useful for obtaining a set of human (or other animal) orthologs that can be used for drug discovery, treatment, diagnosis, and the other uses disclosed herein. The subarrays can be used to specifically create a corresponding human (or other animal) subarray that is relevant to a specific biological function.
30 Accordingly, a method is provided for obtaining sets of genes from other organisms, which sets are correlated with, for example, disease or developmental disorders.

In a preferred embodiment of the invention, the arrays and subarrays disclosed herein are in a "microarray". The term "microarray" is intended to designate an array

of nucleic acid sequences on a chip. This includes *in situ* synthesis of desired nucleic acid sequences directly on the chip material, or affixing previously chemically synthesized nucleic acid sequences or nucleic acid sequences produced by recombinant DNA methodology onto the chip material. In the case of recombinant DNA methodology, nucleic acids can include whole vectors containing desired inserts, such as phages and plasmids, the desired inserts removed from the vector as by, PCR cloning, cDNA synthesized from mRNA, mRNA modified to avoid degradation, and the like.

A series of state-of-the-art reviews of the technology for production of nucleic acid microarrays in various formats and examples of their utilization to address biological problems is provided in *Nature Genetics*, 21 Supplement, January 1999. These topics include molecular interactions on microarrays, expression profiling using cDNA microarrays, making and reading microarrays, high density synthetic oligonucleotide arrays, sequencing and mutation analysis using oligonucleotide microarrays, the use of microarrays in drug discovery and development, gene expression informatics, and use of arrays in population genetics. Various microarray substrates, methods for processing the substrates to affix the nucleic acids onto the substrates, processes for hybridization of the nucleic acid on the substrate to an external nucleic acid sample, methods for detection, and methods for analyzing expression data using specific algorithms have been widely disclosed in the art. References disclosing various microarray technologies are listed below.

Lashkari *et al.* (1997) "Yeast Microarrays for Genome Wide Parallel Genetic and Gene Expression Analysis", *Proc. Natl. Acad. Sci.* 94:13057-13062; Ramsay (1998) "DNA Chips: State-of-the-Art", *Nature Biotechnology* 16:40-44; Marshall *et al.* (1998) "DNA Chips: An Array of Possibilities", *Nature Biotechnology* 16:27-31; Wodicka *et al.* (1997) "Genome-Wide Expression Monitoring In *Saccharomyces Cerevisiae*", *Nature Biotechnology* 15:1359-1367; Southern *et al.* (1999) "Molecular Interactions On Microarrays", *Nature Genetics* 21(1):5-9; Duggan, *et al.* (1999) *Nature Genetics* 21(1):10-14; Cheung *et al.* (1999) "Making and Reading Microarrays", *Nature Genetics* 21(1):15-19; Lipshutz *et al.* (1999) "High Density Synthetic Oligonucleotide Arrays", *Nature Genetics* 21(1):20-24; Bowtell (1999) *Nature Genetics* 21:25-32; Brown *et al.* (1999) "Exploring the New World of the Genome with DNA Microarrays" *Nature Genetics* 21(1):33-37; Cole *et al.* (1999)

- “The Genetics of Cancer--A 3D Model” *Nature Genetics* 21(1):38-41; Hacia (1999) “Resequencing and Mutational Analysis Using Oligonucleotide Microarrays”, *Nature Genetics* 21(1):42-47; Debouck *et al.* (1999) “DNA Microarrays in Drug Discovery and Development”, *Nature Genetics* 21(1):48-50; Bassett, Jr. *et al.* (1999) “Gene Expression Informatics--It’s All In Your Mine”, *Nature Genetics* 21(1):51-55;
- 5 Chakravarti (1999) “Population Genetic--Making Sense Out of Sequence”, *Nature Genetics* 21(1):56-60; Chee *et al.* (1996) “Accessing Genetic Information with High-Density DNA Arrays”, *Science* 274:610-614; Lockhart *et al.* (1996) “Expression Monitoring by Hybridization to High-Density Oligonucleotide Arrays”, *Nature*
- 10 *Biotechnology* 14:1675-1680; Tamayo *et al.* (1999) “Interpreting Patterns of Gene Expression with Self-Organizing Maps: Methods and Application to Hematopoietic Differentiation”, *Proc. Natl. Acad. Sci.* 96:2907-2912; Eisen *et al.* (1998) “Cluster Analysis and Display of Genome-Wide Expression Patterns”, *Proc. Natl. Acad. Sci.* 95:14863-14868; Wen *et al.* (1998) “Large-Scale Temporal Gene Expression
- 15 Mapping of Central Nervous System Development”, *Proc. Natl. Acad. Sci.* 95:334-339; Ermolaeva *et al.* (1998) “Data Management and Analysis for Gene Expression Arrays”, *Nature Genetics* 20:19-23; Wang *et al.* (1998) “A Strategy for Genome-Wide Gene Analysis: Integrated Procedure for Gene Identification”, *Proc. Natl. Acad. Sci.* 95:11909-11914; U.S. Patent No. 5,837,832; U.S. Patent No. 5,861,242;
- 20 WO 97/10363.

In the instant case, the microarray contains nucleic acid sequences on a Biotodyne B filter. However, any medium, including those that are well-known and available to the person of ordinary skill in the art, to which nucleic acids can be affixed in a manner suitable to allow hybridization, are encompassed by the invention.

25 This includes, but is not limited to, any of the membranes disclosed in the references above, which are incorporated herein for reference to those membranes, and other membranes that are commercially available, including but not limited to, nitrocellulose-1, supported nitrocellulose-1, and Biotodyne A, which is a neutrally-charged nylon membrane suitable for Southern transfer and dot blotting procedures.

30 (All are available from Life Technologies.)

EXAMPLE

Summary

Programmed cell death (PCD) in rat cerebellar granule neurons (CGNs)
5 induced by potassium (K^+) withdrawal has been shown to depend on *de novo* RNA
synthesis. The inventors characterized this transcriptional component of CGN
programmed cell death using a custom-built brain-biased cDNA array representing
over 7000 different rat genes. Consistent with carefully orchestrated mRNA
10 regulation, the profiles of 234 differentially expressed genes segregated into distinct
temporal groups (immediate early, early, middle, and late) encompassing genes
involved in distinct physiological responses including cell-cell signaling, nuclear
reorganization, apoptosis, and differentiation. A set of 64 genes, including 22 novel
genes, were regulated by both K^+ withdrawal and kainate treatment. Human
15 homologs were isolated for 8 of these novel regulated genes: The sequences of these
human homologs are shown in SEQ ID NOS:1 (human NARC 9B), 2 (human NARC
8B), 3 (human NARC 2A), 4 (human NARC 16B), 5 (human NARC 10C), 6 (human
NARC 1C), 8 (human NARC 1A), and 10 (human NARC 25).
Thus, array technology was used to broadly characterize physiological responses at
20 the transcriptional level and identify novel genes induced by multiple models of
programmed cell death.

Background

In neurons, programmed cell death is an essential component of neuronal
development (Jacobson *et al.* 1997; Pettmann and Henderson (1998); Pettmann and
25 Henderson (1998) *Neuron* 20:633-747) and has been associated with many forms of
neurodegeneration (Hetts (1998) *Journal of the American Medical Association*
279:300-307). In the cerebellum, granule cell development occurs postnatally. The
final number of neurons represents the combined effects of additive processes such as
cell division and subtractive processes such as target-related programmed cell death.
30 Depolarization due to high concentrations (25 mM) of extracellular potassium (K^+)
promotes the survival of cerebellar granule neurons (CGNs) *in vitro*. CGNs
maintained in serum containing medium with high K^+ will undergo programmed cell
death when switched to serum-free medium with low K^+ (5 mM) (D'Mello *et al.*

(1993) *Proc. Natl. Acad. Sci. USA* 90:10989-10993; Miller and Johnson (1996) *Journal of Neuroscience* 16:7487-7495). The resulting programmed cell death has a transcriptional component that can be blocked by inhibitors of new RNA synthesis (Galli *et al.* (1995) *Journal of Neuroscience* 15:1172-1179; and Schulz and Klockgether (1996) *Journal of Neuroscience* 16:4696-4706). Traditionally, the regulation of limited numbers of specific genes were characterized during CGN programmed cell death using Northern nucleic acid hybridization (*e.g.* PTZ-17, Roschier *et al.* (1998) *Biochemical and Biophysical Research Communications* 252:10-13), reverse transcription polymerase chain reaction (RT-PCR; *e.g.* *c-jun*, cyclophilin, cyclin D1, *c-fos* and caspase (Miller *et al.* (1997) *Journal of Cell Biology* 139:205-217), and *in situ* hybridization (*e.g.* RP-8; Owens *et al.* (1995) *Developmental Brain Research* 86:35-47).

High-density cDNA arrays have been successfully used to characterize genome-wide mRNA expression in yeast (Lashkari *et al.* (1997) *Proc. Natl. Acad. Sci. USA* 94:13057-13062; Wodicka *et al.* (1997) *Nature Biotechnology* 15:1997). In higher eukaryotes, the strategy has been to array as many sequences as possible from known genes, from expressed sequence tags (ESTs), or from uncharacterized cDNA clones from a library (Bowtell (1999) *Nature Genetics* 21:25-32; Duggan *et al.* (1999) *Nature Genetics* 21:10-14; Marshall and Hodgson (1998) *Nature Biotechnology* 16:27-31; and Ramsay (1998) *Nature Biotechnology* 16:40-44). Global RNA regulation during cellular processes including cell-cycle regulation (Cho *et al.* (1998) *Molecular Cell* 2:65-73, and Spellman *et al.* (1998) *Mol. Biol. Cell.* 95:14863-14868), fibroblast growth control (Iyer *et al.* (1999) *Science* 283:83-87), metabolic responses to growth medium (Derisi and Brown (1997) *Science* 278: 680-686), and germ cell development (Chu *et al.* (1998) *Science* 282:699-705) have been temporally monitored using arrays. The program of gene expression delineated in these studies demonstrated a correlation between common function and coordinate expression, and also provided a comprehensive, dynamic picture of the processes involved (Brown and Botstein (1999) *Nature Genetics* 21:33-37). For the cellular process of programmed cell death, a DNA chip has been used to identify twelve known genes as differentially expressed between two conditions, etoposide-treated and untreated cells (Wang *et al.* (1999) *FEBS Letters* 445:269-273).

A genome-wide approach for the comprehensive characterization of the transcriptional component of rat CGN programmed cell death and for identification of novel neuronal apoptosis genes requires an array consisting of both known and novel rat cDNAs. The inventors constructed a brain-biased and programmed cell death-enriched clone set by arraying ~7300 consolidated ESTs from two cDNA libraries cloned from rat frontal cortex and differentiated PC12 cells deprived of nerve growth factor (NGF), and >300 genes that are known markers for the central nervous system and/or programmed cell death. They reproducibly and simultaneously monitored the expression of the genes at 1, 3, 6, 12, and 24 hours after K^+ withdrawal. They then categorized the regulated genes by time course expression pattern to identify cellular processes mobilized by CGN programmed cell death at the RNA level. In particular they focused on the expression profiles of many known pro- and anti-apoptotic regulatory proteins, including transcription factors, Bcl-2 family members, caspases, cyclins, heat shock proteins (HSPs), inhibitors of apoptosis (IAPs), growth factors and receptors, other signal transduction molecules, p53, superoxide dismutases (SODs); and other stress response genes. Finally, they compared the time courses of regulated genes induced by K^+ withdrawal in the presence or absence of serum to those induced by glutamate toxicity. Thus, they identified a restricted set of relevant genes regulated by multiple models of programmed cell death in CGNs.

Results

Construction and validation of a brain-biased cDNA microarray

In order to characterize the transcriptional component of neuronal apoptosis in rat cerebellar granule neurons, the inventors constructed a cDNA array, called Smart Chip™ I, that contains primarily rat brain genes. Figure 1 shows a schematic representation of the construction of the microarray. Two cDNA libraries were cloned from rat frontal cortex and nerve growth factor-deprived rat PC12 cells to enrich for cDNAs expressed in the central nervous system and in one *in vitro* model of neuronal apoptosis. Expressed sequence tags (ESTs) from the 5'-end were identified for 8,304 clones in the cortical library and 5,680 in the PC12 library. These 13,984 ESTs were condensed into 7,399 unique sequence clusters by using the Basic Local Alignment Search Tool (BLAST) sequence comparison analysis (Altschul *et al.* 1990) to identify ESTs with overlapping sequence. One representative clone was

chosen from each of 7,296 of the unique sequence clusters and prepared for PCR amplification using a robotic sample processor. In addition to the ESTs, PCR templates were prepared for 289 known DNA sequences, including negative controls, genes with known function in the CNS and/or during programmed cell death, and genes previously identified as regulated by CGN programmed cell death using differential display (data not shown). To check the fidelity of the set of array elements, a robotic sample processor was used to randomly choose 212 clones for sequencing. Ten clones produced poor sequence. The remaining 202 matched their seed sequence (data not shown), implicating 100% fidelity in sample tracking.

A sample volume of 20 nl from each of the 7584 PCR products was arrayed onto nylon filters at a density of $\sim 64/\text{cm}^2$ using a pin robot. The arrayed DNA elements were denatured and covalently attached to the nylon filters for use in reverse Northern nucleic acid hybridization experiments. In a typical experiment, "radiolabeled RNA", 1 μg polyA RNA radiolabeled by ^{33}P -dCTP incorporation during cDNA synthesis, was hybridized to triplicate arrays following RNA hydrolysis. Subsequently, the filters were washed and exposed to phosphoimage screens. Gene expression was quantified for each array element by digitizing the phosphoimage-captured hybridization signal intensity. Figure 2 illustrates that the coefficient of variation between triplicate hybridizations averaged less than 0.2 for genes whose intensities were above a threshold of 30-40 units. From control experiments when *in vitro* transcribed RNAs were deliberately spiked into samples, this threshold amounted to a copy number of less than 1 in 100,000 (data not shown).

Tissue distribution of brain-biased Smart Chip ESTs

To characterize the brain-biased cDNA array and possibly identify brain-specific genes, radiolabeled RNA from ten different normal rat tissues was hybridized to Smart Chip. Compared to heart, kidney, liver, lung, pancreas, skeletal muscle, smooth muscle, spleen, and testes, radiolabeled rat brain RNA produced more hybridization signal intensity against most of the brain-biased array elements. After data normalization and averaging between replicates, the threshold of detection was determined for each experiment and the number of genes detected for each tissue was tabulated (Figure 6). Most (6127 out of 7296) but not all of the ESTs were detected in at least one of the tissues profiled. The number of genes detected in brain was the

highest. 582 genes appeared to be brain-specific, as defined by detection above threshold for brain but below threshold for any of the other nine tissues.

The physiology of CGN KCl/serum-withdrawal as characterized by transcription 5 profiling on Smart Chip

Using the brain-biased, programmed cell death nucleic acid-enriched Smart Chip, global mRNA expression was profiled throughout a time course of KCl/serum-withdrawal-induced cell death in primary cultures of CGNs. The transcription-dependent CGN programmed cell death was coordinated, resulting in less than 30%
10 survival at 24 hours post-withdrawal as quantified by cell counting (data not shown). RNA samples, designated "treated", were isolated at 1, 3, 6, 12, and 24 hours after switching post-natal day eight CGNs from medium containing 5% serum and 25 mM KCl to serum-free medium with 5 mM KCl. For controls, the 5% serum/25 mM KCl medium was replaced, and "sham" RNA at 1, 3, 6, 12, and 24 hours was isolated.

15 Since the average coefficient of variation for gene expression intensities between triplicate hybridizations was less than 0.2, genes regulated at least three-fold during the time course (790 out of 6818 detected; data not shown) were further addressed. Using hierarchical clustering algorithms (see Experimental Procedures), the regulated genes were ordered based on their gene expression pattern across the ten
20 experimental points (five time points, sham and treated (Figure 3)). The dendrogram in Figure 3 depicts the hierarchy of relatedness between gene expression profiles. The first major branch point segregated those genes regulated by sham treatment (first five columns), and those regulated by KCl/serum-withdrawal treatment only (last five columns). A majority of genes (556) were regulated by sham treatment. These genes
25 included *trk A*, PSD-95, SV 2A, and VAMP 1, and were most likely induced by serum-add-back in the sham since the medium was exchanged at t=0 with unconditioned medium.

Figure 3 shows the expression pattern of 234 programmed cell death-induced genes that were regulated by KCl/serum-withdrawal only, and were not regulated by
30 serum-add-back in the sham experiments. Their coefficient of variation in expression level throughout the five serum-add-back experiments was less than 20%. Since the serum-add-back experiments were non-discriminating for these genes, the serum-add-back data were averaged to generate a single control data set for clustering with the

KCl/serum withdrawal time course. Four apparent temporal regulation classes were designated immediate early (peaking at 1 hour followed by rapid decay), early (peaking at 3-6 hours), middle (peaking at 6-12 hours), and late (up-regulated at 24 hours). Almost all of the immediate early genes encoded proteins with known roles in regulating secretion and synaptic vesicle release including synaptotagmin, synaphin, NSG-1, calcium calmodulin-dependent kinase II, synapsin, complexin, LDL receptor, and fodrin (Figure 7). Histones 1, 2A, and 3 fell in the early class. Middle genes comprised several known genes induced by programmed cell death or stress, including caspase 3, the mammalian oxy R homolog, cytochrome c oxidase and protein phosphatase Wip-1. Functions encoded for by late genes could be effectors of survival mechanisms including inhibitory neurotransmission (GAD, GABA-A receptor, GABA transporter), cell adhesion (nexin, basement membrane protein 40, phosphacan, rat GRASP), down-regulation of excitatory neurotransmission (glutamate transporter, sodium-dependent glutamate/aspartate transporter), leukotriene metabolism (dithiolethione-induced NADP-dependent leukotriene B4 12-hydroxydehydrogenase, leukotriene A-4 hydrolase), protein stabilization (cysteine proteinase inhibitor cystatin C, N-alpha-acetyl transferase, CaBP2, elongation factor 1-gamma, APG-1), and ionic balance and cell volume (SLC12A integral membrane protein transporter). Based on four distinct waves of gene expression, the major transcriptional responses observed for KCl/serum-withdrawal included initial up-regulation of synaptic vesicle release/recycling, then, of histone biosynthesis, followed by various constituents of programmed cell death regulation and stress-response signaling, and finally, of multiple survival mechanisms. The apparent changes in transcription most likely also reflect changes in the relative cell populations, since late mRNAs may be markers of neurons and non-neuronal cells which have survived KCl/serum-withdrawal at 24 hours. Another contributing factor may be the presence of two populations of dying neurons that respond with different kinetics to serum versus KCl withdrawal, as has been described by other groups.

30 Neuronal apoptosis regulated candidates (NARCs) regulated by multiple models of programmed cell death

112 novel ESTs were significantly regulated by KCl/serum-withdrawal in rat CGNs (data not shown). Some exhibited similar expression profiles throughout

KCl/serum-withdrawal and serum-add-back to genes with known function during programmed cell death, such as caspase 3. The temporally-coupled expression of these novel genes may reflect related functionality with caspase 3, since they probably share common RNA regulatory elements, including those regulating initiation,
5 elongation, processing, and/or stability. Apparent coordinate transcriptional up-regulation of synaptic vesicle release/recycling possibly reflects a physiological response to near cessation of synaptic transmission that may or may not contribute to the programmed cell death pathway. To help further distinguish genes that are specifically regulated in response to programmed cell death, CGN programmed cell
10 death induced by glutamate (excitatory neurotransmitter) toxicity was studied. In addition, the effect of KCl-withdrawal alone on gene expression was examined. This was done under defined medium conditions to minimize the effect of serum on the sham and treated samples.

Rat CGNs from post-natal day seven pups were isolated as before and plated
15 into basal medium Eagle containing "high", 10% dialysed fetal bovine serum, and "high", 25 mM KCl. After two days in culture, the medium was replaced with neurobasal medium supplemented with "low", 0.5% serum, and high KCl. To initiate KCl-withdrawal on day eight, the KCl concentration was switched to 5 mM for the treated samples. The same low serum, high KCl, neurobasal medium was replaced in
20 the controls to minimize gene induction by high serum. For the glutamate toxicity experiment, the cells were treated for 30 min in sodium-free Locke's medium with or without 100 μ M kainate for treated samples and controls, respectively.

After isolation from treated and control samples at 1, 3, 6, and 12 hours after KCl-withdrawal and 2, 4, 6, 12 hours after kainate treatment, mRNA was subjected to
25 expression profiling analysis on Smart Chip I. Figure 4 illustrates the changes in gene expression that occur over time when CGNs are induced to undergo programmed cell death by KCl/serum-withdrawal, KCl-withdrawal alone, or kainate treatment. In the scatter plots, due to differential expression, large numbers of regulated genes migrated away from a line of slope one when withdrawn (W) or treated (T) samples were
30 compared to control (C). The sham treated cells for the KCl/serum-withdrawal clearly responded to basal medium serum-add-back, whereas shams for KCl-withdrawal alone and kainate treatment did not respond to conditioned neurobasal

medium add-back. Profiling across the mRNA levels of thousands of genes provided a clear index of changes in overall cell physiology.

In general, apparent changes in gene expression were less robust in the cells cultured on neurobasal medium. The number of genes detected above threshold was similar for all three paradigms, 6634, 7017, and 6818, respectively, for KCl-withdrawal, kainate treatment, and KCl/serum withdrawal (data not shown). Yet the number of genes regulated by at least three-fold during KCl-withdrawal and kainate treatment was only 156 and 167, respectively (data not shown), compared to the 790 discussed above for KCl/serum withdrawal.

A hierarchical clustering algorithm was used to order the regulated genes based on their gene expression pattern across all CGN programmed cell death paradigms investigated. Twenty-six individual profiling experiments in duplicate or triplicate were performed across the 7584 rat genes on Smart Chip I using mRNA isolated from 5 serum-add-back time points, 5 KCl/serum-withdrawal time points, 4 time points each for sham and KCl-withdrawal, and 4 time points each for sham and kainate treatment.

Figure 4 shows expression clusters generated by one hierarchical clustering algorithm. The inset shows a specific group of genes having similar expression patterns. This group includes genes known to be regulated in programmed cell death, for example caspase 3 and Wip 1, as well as other nucleic acid sequences on the array not previously known to be regulated. Those sequences meeting specific criteria were designated "neuronal apoptosis regulated candidate" (NARC). Criteria for designating such genes were based on specific expression criteria as shown in Figure 4. Nucleic acid sequences having an expression pattern similar to genes known to be involved in apoptosis were designated as NARC sequences.

Gene expression validation by RT-PCR

Although the reproducibility in transcription profiling experiments was quite high (average CV<0.2), the gene expression regulation of known and novel genes was validated by semi-quantitative RT-PCR. The rat CGN model system was used to independently validate the expression of several NARC genes that had shown expression (when hybridized with sequences on the chip) related to programmed cell death. Reverse transcriptase-assisted PCR was performed to assess expression of

NARC 1-7, 9, 12, 13, 15, and 16. Experimental samples received KCl withdrawal treatment. Control samples show cells receiving no treatment. The PCR reactions contained 10, 5, 2.5, 1.3, and 0.7 ng of total RNA each. The RT-PCR protocol is disclosed in the exemplary material herein. NARC 1, 2, 4, 5, 7, 9, 12, 13, 15 and 16 all showed significantly increased expression 3-6 hours after KCl withdrawal. The designation "N" above is an abbreviation of the acronym "NARC" which is an abbreviation of "neuronal apoptosis regulated candidate" as described in the Examples section.

NARC1 and NARC2 regulation *in vivo* during cerebellar development

Two novel neuronal apoptosis regulated candidates, NARC1 and NARC2, were validated by *in situ* hybridization and shown to be coordinately up-regulated with caspase 3 during postnatal development when increased apoptosis is associated with synapse consolidation in the cerebellum (not shown).

Experimental Procedures

BLAST sequence comparison analysis

ESTs determined for the 5'-end of cDNA clones picked from two cDNA libraries, rat frontal cortex (8,304 clones) and NGF-deprived differentiated PC12 cells (5,680 clones), ranged from 100-1000 nt in sequence length and averaged 500 nt (data not shown). Sequence comparisons were done using BLAST (Altschul et al. 1990). Contiguous matches defined a sequence cluster. Large clusters were checked by hand to eliminate apparent chimeras. From 13,984 sequences inputted, the analysis identified 5,779 singletons and 1,620 larger clusters (data not shown). The 5'-most clone was selected from the larger clusters. Because two 96-well microtiter plates of clones were missing, a total of 7,296 out of the 7,399 identified were selected for Smart Chip™ I.

cDNA microarray construction

Using a Genesis RSP 150 robotic sample processor (Tecan AG, Switzerland), bacterial cultures of individual EST clones from the two libraries were consolidated

from 13,792 clones spanning 144 96-well microtiter plates to 7296 Smart Chip I clones spanning 76 plates. To prepare templates for array elements, oligonucleotide primers specific for vector sequences up- and downstream of the cloning site were used to amplify the cDNA insert by PCR. Following ethanol precipitation and
5 concentration (to 1-10 mg/ml), the array element templates were resuspended in 3X SSC (1X SSC: 150 mM sodium chloride, 15 mM sodium citrate, pH 7.0). A sample volume of 20 nl from each template was arrayed onto nylon filters (Biodyne B, Gibco BRL Life Technologies, Gaithersburg, MD) at a density of $\sim 64/\text{cm}^2$ using a 96-well format pin robot (THOR). After the filters were dry, the arrayed DNA was denatured
10 in 0.4 M sodium hydroxide, neutralized in 0.1 M Tris-HCl, pH 7.5, rinsed in 2X SSC, and dried to completion.

Array hybridization

Rat poly A⁺ RNA was purchased from Clontech (Palo Alto, CA) for the organ
15 recital (Figure 8) or was isolated as total RNA from cultured CGNs using RNA STAT-60™ (Tel-Test, Inc., Friendswood, TX) and then prepared using Oligotex™ (Qiagen, Inc., Chatsworth, CA). Re-annealed 1 μg mRNA and 1 μg oligo(dT)₃₀ was incubated at 50°C for 30 min with SuperScript™ II as recommended by Gibco in the presence of 0.5 mM each deoxynucleotide dATP, dGTP, and dTTP, and 100 μCi $\alpha^{33}\text{P}$ -
20 dCTP (2000-4000 Ci/mmol; NEN™ Life Science Products, Boston, MA). After purification over Chroma Spin™ +TE-30 columns (Clontech), the labeled cDNA was annealed with 10 μg poly(dA)_{>200} and 10 μg rat Cot-1 DNA (prepared as described in Britten *et al.* (1974) *Methods in Enzymology* 29:263-418). At 2×10^6 cpm/ml, the annealed cDNA mixture was added to array filters in pre-annealing solution
25 containing 100 mg/ml sheared salmon sperm DNA in 7% SDS (sodium dodecyl sulfate), 0.25 M sodium phosphate, 1 mM ethylenediaminetetraacetic acid, and 10% formamide. Following over night hybridization at 65°C in a rotisserie-style incubator (Robbins Scientific, Sunnyvale, CA), the array filters were washed twice for 15 min at 22°C in 2X SSC, 1% SDS, twice for 30 min at 65°C in 0.2X SSC, 0.5% SDS, and
30 twice for 15 min at 22°C in 2X SSC. The array filters were then dried and exposed to phosphoimage screens for 48 h. The radioactive hybridization signals were captured with a Fuji BAS 2500 phosphoimager and quantified using Array Vision™ software (Imaging Research Inc., Canada). Array hybridizations for the organ recital, the CGN

KCl only-withdrawal, and the CGN kainate treatment experiments were performed in triplicate; for the CGN KCl/serum-withdrawal, they were performed in duplicate.

Transcription profiling data analysis

5 For replicate array hybridizations, the distribution of signal intensities across all rat genes was normalized to a median of 100. Replicate measurements were averaged and a coefficient of variation (CV; standard deviation/mean for triplicates or the absolute value of the difference/mean for duplicates) was determined for each gene. The detection threshold was chosen for each hybridization experiment by
10 graphing the moving average (with a window of 200) for CV versus mean gene expression intensity (Figure 2). The threshold was defined as the intensity at which lower intensities exhibited an average CV that was greater than 0.3. For most experiments, this threshold ranged from 10 to 40, and the number of genes detected above threshold ranged from 70% to 95%.

15

CGN cell culture

CGNs were prepared from seven day old rat pups as previously described (Johnson and Miller (1996) *Journal of Neuroscience* 16:74877-7495). Briefly, cerebella were isolated, and meningeal layers and blood vessels were removed under
20 a dissecting scope. Dissociated cells were plated at a density of 2.3×10^5 cells/cm² in basal medium Eagle (BME; Gibco) supplemented with 25 mM KCl, 10% dialyzed fetal bovine serum (Summit Biotechnology lot #04D35, Ft. Collins, CO), 100 U/ml penicillin, and 100 µg/ml streptomycin. Aphidicolin (Sigma, St. Louis, MO) was added to the cultures at 3.3 µg/ml, 24 hours after initial plating to reduce the number
25 of non-neuronal cells to less than 1-5%.

For KCl/serum-withdrawal experiments, after seven days in culture, the treated cells were switched to 5 mM KCl, BME, no serum, while the shams received a medium replacement. By 24 hours post-withdrawal, less than 30% of the cells were surviving as assayed by Hoechts cell counts (data not shown). This apparent cell
30 death could be rescued by actinomycin D at 2 µg/ml (data not shown).

For the KCl-withdrawal alone and kainate treatment experiments, on day two in culture, the medium was replaced with neurobasal medium (Gibco) supplemented with 25 mM KCl, 0.5% dialyzed fetal bovine serum, B27 supplement (Gibco), 0.5

mM L-glutamine (Gibco), 0.1 mg/ml AlbuMAX I (Gibco), 100 U/ml penicillin, 100 µg/ml streptomycin, and 3.3 µg/ml aphidicolin. On day seven, KCl-withdrawal was initiated by replacing the medium with 5 mM KCl while the shams received 25 mM. By 24 hours post-withdrawal, 40% of the cells were surviving as assayed by Hoechts cell counts (data not shown). As previously described, glutamate toxicity was induced by replacing the medium for 30 min with 5 mM KCl, 100 µM kainic acid (Sigma) in sodium free Locke's buffer, while the shams received no kainic acid (Coyle *et al.* (1996) *Neuroscience* 74:675-683). After 30 min, the supplemented neurobasal medium was replaced. By 12 hours post-withdrawal, 30% of the cells were surviving as assayed by Hoechts cell counts (data not shown). The KCl-withdrawal induced cell death was rescued by actinomycin D, whereas the kainate-induced was not.

Expression data clustering algorithms

After normalization and averaging of the KCl/serum-withdrawal data, 790 genes passed the following criteria over the 10 time points (5 treated, 5 sham) for input into heirarchical clustering analysis: 1. detection, maximum intensity greater than 30; 2. noise filter, the difference between maximum and minimum intensity greater than 30; and 3. regulation, fold induction between maximum and minimum intensity of at least 3 (data not shown). Hierarchical clusters were ordered based on Euclidian distances. 234 out of 790 genes that passed the significance filter described above were not regulated in the controls based on CV less than 0.2 for all five control time points (data not shown).

RT-PCR

Oligonucleotide primer sequences specific for each EST validated by RT-PCR were selected from quality sequence regions and designed to obtain a melting temperature of 55-60°C as predicted by PrimerSelect software (DNASTAR, Inc., Madison, WI) based on DNA stability measurements by (Breslauer *et al.* (1986) *Proc. Natl. Acad. Sci. USA* 83:3746-3750). The Stratagene Opti-Prime™ Kit (La Jolla, CA) was used to determine optimal RT-PCR amplification conditions for each primer pair. RT-PCR reactions on 2-fold serially diluted CGN programmed cell death cDNA were set up using the Genesis RSP 150 robotic sample processor and

incorporating the optimal buffer conditions for each primer pair. Every robot run included primers specific for housekeeping genes to control for day to day differences in cDNA template dilutions. The number of cycles was adjusted to obtain a linear range of amplification by comparing the amount of product made from the serially
5 diluted templates as assessed by agarose gel electrophoresis.

Preparation of Array on Nylon

10 I. Procedure for Generating Labeled First Strand cDNA Using Superscript II Reverse Transcriptase

1. 10 mL (100 mCi) 33P α -dCTP was dried down by SpeedVac.
2. In a separate tube, the following components were mixed:
1.0 ug Poly A+ RNA or 10 ug Total RNA
15 1 uL 1 ug/uL oligo-dT(30)
x uL DEPC-H₂O, to 10 uL

The above sample was heated at 70°C for 4 minutes and then placed on ice.

3. ~~8uL from the oligo/RNA mixture (#2)~~ was removed and used to resuspend the dried 3P3. The following components were added to the reaction:

- 20 4 uL 5X First Strand Buffer (comes with Superscript II RT)
- 2 uL 100 mM DTT
- 1 uL 10 mM dAGT-TPs
- 1 uL 0.1 mM cold dCTP
- 1 uL Rnase Inhibitor
- 25 1 uL Superscript II RT

The reaction was incubated for 30 minutes at 50°C.

4. After incubation, 2 uL 0.5 M NaOH, and 2 uL 10 mM EDTA were added. The reaction was heated at 65°C, for 10 minutes to degrade RNA template.

- 30 5. The volume was brought to 50 uL (i.e., add 26 uL H₂O).
6. One Choma-Spin +TE 30 column (Clontech, #K1321) was prepared for every probe made.

a. Air bubbles were removed from the column.

b. The break-away end of the column was removed and the column placed in an empty 2 mL tube and spun for 5 minutes at 700g (in Eppendorf 5415C "3.5").

5 c. The column was removed and the flow-through discarded. The column was placed in clean tube. The probe was added slowly to the center of the column bed without disturbing the matrix so that the liquid did not touch the side of the column and flow down the edge of the column wall.

d. The probe was eluted by spinning the column as above.

10 II. Hybridization

1. The hybridization chamber was preheated to 65°C.

2. 10 mL of 10% Formamide Church Buffer was added. This was placed in the hybridization chamber for around 15 minutes.

15 3. Sheared salmon sperm DNA was denatured at 95°C for 5 minutes, placed on ice, and then added to the hybridization mixture at a final concentration of 100 ug/mL. Prehybridization was for 1.5 hours.

4. The amount of probe was calculated necessary to achieve 2×10^6 cpm/mL for 10 mL.

5. The Cot Annealing Reactions (per bottle) were as follows:

20 Rat probe with Rat Filters:

10ug Poly dA (>200nt)

10ug Rat Cot 10 DNA

25uL 20 x SSC

probe + water to 100uL

25 Mouse probe with Rat Filters:

10ug Poly dA (>200nt)

10ug Mouse Cot 1 DNA

25uL 20 x SSC

probe + water to 100uL

30 Also added 5ug Rat Cot 10 DNA to the prehybridization.

Human probe with Human Filters:

10ug Poly dA (>200nt)

10ug Human Cot 1 DNA

25uL 20 x SSC

probe + water to 100uL

5 The probe was heated to 95°C, and then probe was allowed to preanneal at 65°C, for 1.5 hours.

6. The probe was added to prehybridizing filters (directly to the solution and not onto the filters) and hybridization was for approximately 20 hours.

10 III. Washing

1. Probe was removed.

2. Three quick washes were performed with preheated 2 x SSC/1% SDS, 65°C (washes could be done in roller bottles).

3. Two washes were performed for 15 minutes each with preheated high stringency wash buffer:

15 0.5 x SSC, 0.1% SDS for cross species washes

0.5 x SSC, 0.1% SDS for normal washes

0.1 x SSC, 0.1% SDS for very high stringency washes

20 4. After the high stringency washes, the filters were rinsed in a large square petri dish in 2 x SSC, no SDS. For experiments in which many filters are used, the 2 x SSC is frequently changed so there is no residual SDS left on the filters.

25 5. The filters were removed from the 2 x SSC and placed on Whatman filter paper. Filters were baked at 85°C for 1 hour or longer. Screens were protected against any moisture. Filters were placed on a blank phosphorimager screen. No yellowed phosphorimager screens were used since they may not respond to exposure linearly. Screens had been erased on a light box for no less than 20 minutes.

6. Blots were exposed to the screen at least 48 hours or as necessary.

30 IV. Scanning Filters on Fuji Phosphorimager

Gradation 16 bit, Resolution 50m, Dynamic Range S4000, select Read and Launch Image Gauge. Image was saved on the hard drive.

APPENDIX I:

10% Formamide-Church Buffer:

- 59.6mL water
- 70mL 20% SDS
- 50mL 2M NaPO₄ pH 7.2
- 5 20mL Ultrapure Formamide
- 0.4mL 0.5M EDTA pH 8.0

The above components were added to water, mixed, and filtered through a 0.2 um filter.

10 RT-PCR Protocol

I. For one PCR reaction mix, the following components were used:

- 28ul 5X First Strand Buffer
- 14ul 0.1M DTT
- 4ul dNTPs (20 mM)
- 15 7ul Rnase Inhibitor
- 7ul Superscript II

This buffer can be stored at -80°C for 3 months.

II. Total RNA was reversed transcribed as follows:

- 20 1.4ug Total RNA (DNAsed)
- 14ul Random Primers (50ng/ul--Gibco)

Water was added to 60ul. The mixture was incubated at 70°C for 10 minutes and then placed on ice for 2 minutes. 60ul of the RT Reaction Mix was added. Incubation was at room temperature for 10 minutes, then 50°C for 30 minutes, then 90°C for 10
25 minutes. The sample was diluted with 480ul water to result in 10ng per 5ul.

III. The PCR reaction was performed with the following ingredients:

- 5ul 4x PCR Buffer
- 5ul cDNA (at 10ng/5ul)
- 30 5ul 1uM Primer Pair
- 5ul Enzyme Cocktail (0.2ul Hot Start Taq, 1ul 2mM
dNTPs, 3.8ul water)

IV. Cycling was as follows:

- 5 95°C 15 minutes
 94°C 30 seconds
 52°C 30 seconds
 72°C 1 minute
 Cycle 26-30 times
 72°C 10 minutes
 4°C Hold

10 Cerebellar granule cell isolation was performed according to the method disclosed in Johnson *et al.* (1996) *J. Neurosci.* 16:74877-7495.

 The induction of apoptosis in neurites induced by kainate is described in *Neurosci.* 75:675-683 (1996). The procedure shown in this reference was followed.

 The following parameters were checked:

- 15 (1) Cerebellum granule neuron viability following potassium and serum withdrawal at time points corresponding to PCR-based methods for differential gene expression (Hoechst stain).
- (2) Effects of 2 ug/ml actinomycin D on potassium and serum withdrawal at 24 hours on cerebellar granule neurons; viability by Hoeschst stained cell counts.
- 20 (3) Time course of kainate-induced cell death for parallel analysis of PCR-based method for differential gene expression of CGN Poly A mRNA.
- (4) Time course of kainate-induced (30 minute exposure) apoptosis in CGNs; analysis by Hoechst cell counts.
- (5) Time course of potassium withdrawal apoptosis in CGNs in defined
- 25 media for PCR-based method for differential gene expression of analysis by Hoechst counts.

 While this invention has been particularly shown and described with reference

30 to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the spirit and scope of the invention as defined by the appended claims.

THAT WHICH IS CLAIMED:

1. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:
 - (a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, and
 - (b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
5 6, 8, and 10.
2. An isolated nucleic acid molecule consisting of a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - 10 b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
6, 8, and 10.
3. An isolated nucleic acid molecule consisting of a fragment of a nucleotide sequence selected from the group consisting of:
 - 15 a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
6, 8, and 10;wherein said fragment is at least 15 nucleotides in length.
- 20 4. A nucleic acid molecule comprising a nucleotide sequence which is at least 60% identical to a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
6, 8, and 10.
- 25 5. A nucleic acid molecule which hybridizes under high stringency conditions to a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
30 6, 8, and 10.

6. A vector comprising a nucleotide sequence selected from the group consisting of:
- a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10,
 - b) a nucleotide sequence which is at least 60% identical to a nucleotide sequence shown in SEQ ID NOS: 1-6, 8, and 10
 - c) a nucleotide sequence which hybridizes under high stringency conditions to a nucleotide sequence shown in SEQ ID NOS: 1-6, 8, and 10.
 - d) a complement of a nucleotide sequence of a, b, or c.
7. The vector of claim 6, wherein the isolated nucleic acid molecule is operably linked to at least one expression control element.
8. A host cell comprising the vector of claim 7.
9. A method for preparing a polypeptide comprising culturing the host cell of claim 8 under conditions in which the nucleic acid molecule is expressed.
10. An isolated polypeptide encoded by the nucleic acid molecule of claim 1.
11. An isolated polypeptide encoded by the nucleic acid molecule of claim 4.
12. An isolated polypeptide encoded by the nucleic acid molecule of claim 5.
13. An antibody which selectively binds to the polypeptide of claim 10.
14. An antibody which selectively binds to the polypeptide of claim 11.
15. An antibody which selectively binds to the polypeptide of claim 12.
16. A method for assaying for the presence of a nucleic acid molecule in a sample, comprising the steps of
- (a) contacting said sample with a nucleic acid probe that selectively hybridizes to the nucleic acid molecule, wherein said nucleic acid probe is selected from

the group consisting of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10; fragments of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, wherein said fragments are at least 15 nucleotides in length; and fragments of the complements of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, wherein said fragments of the complements are at least 15 nucleotides in length; and

(b) determining whether the nucleic acid probe binds to a nucleic acid molecule in the sample

10 17. A method for detecting a polypeptide of claim 10 in a sample, comprising the steps of:

(a) contacting the sample with an antibody that binds to a polypeptide of claim 10, and

(b) determining whether the compound binds to the polypeptide in the sample.

15 18. A method for modulating the activity of a polypeptide of claim 10, said method comprising contacting the polypeptide of claim 10 with an agent under conditions that allow the agent to modulate the activity of the polypeptide.

20

19. The method of claim 18, wherein said agent is an antibody that binds to said polypeptide

20. The method of claim 18, wherein said polypeptide is in a cell derived from the central nervous system.

25

21. The method of claim 18, wherein said cell derived from the central nervous system is undergoing aberrant apoptosis.

22. The method of claim 18, wherein said activity is modulated in a subject having or predisposed to having a disorder involving the central nervous system.

30

23. The method of claim 18, wherein said activity is modulated in a subject having or predisposed to having a disorder involving aberrant apoptosis.

24. A method for treating a disorder involving the central nervous system comprising administering any of the polypeptides of claim 10 to a subject having or at risk of developing said disorder.

25. A method for treating a disorder involving aberrant apoptosis comprising administering any of the polypeptides of claim 10 to a subject having or at risk of developing said disorder.

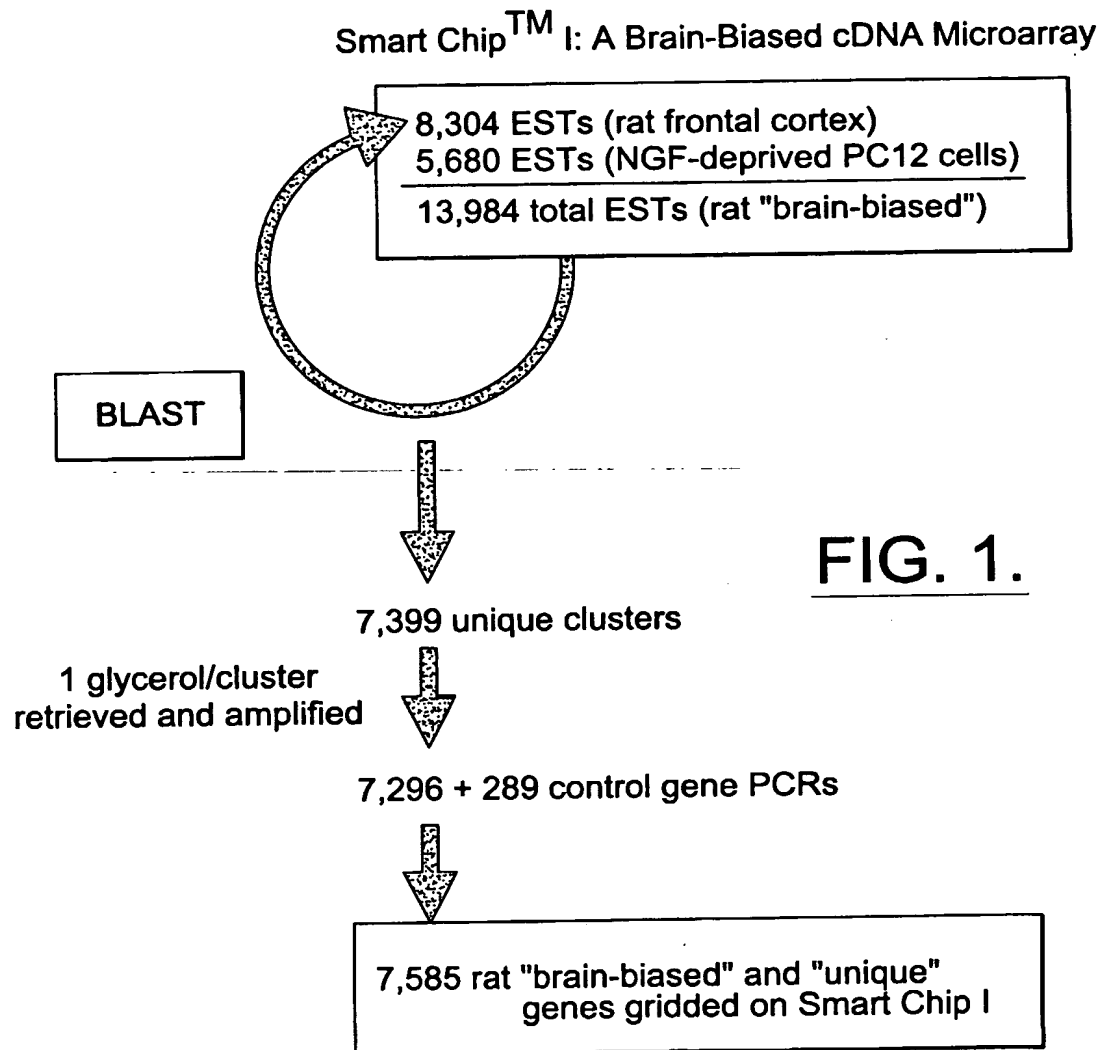
26. A kit comprising a nucleic acid probe which hybridizes to a nucleotide sequence of claim 1 and instructions for use.

27. A kit comprising an agent which binds to a polypeptide of claim 10 and instructions for use.

28. The kit of claim 35, wherein said agent is an antibody.

20

1/31



2/31

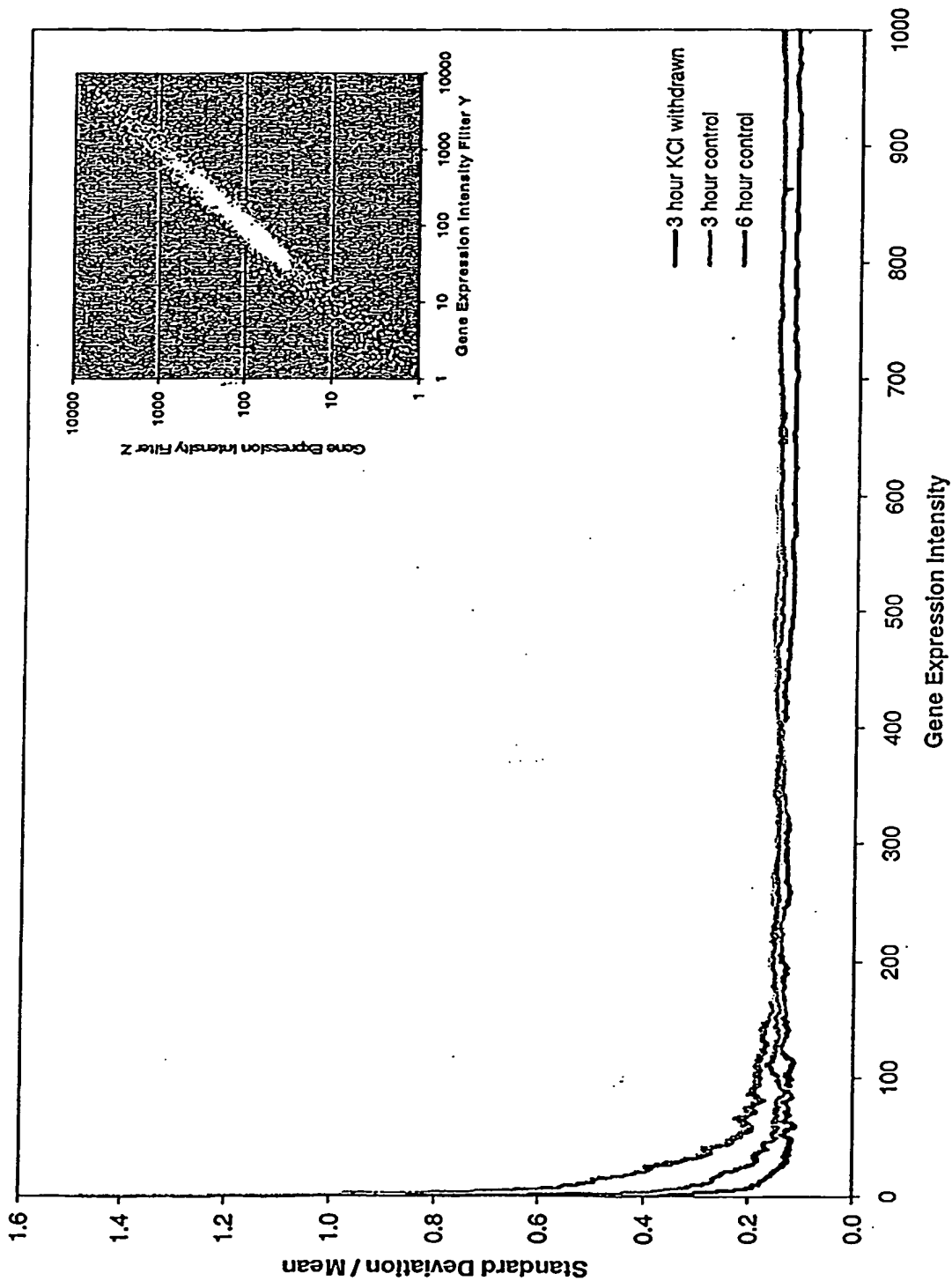


FIGURE 2

A.

3/31

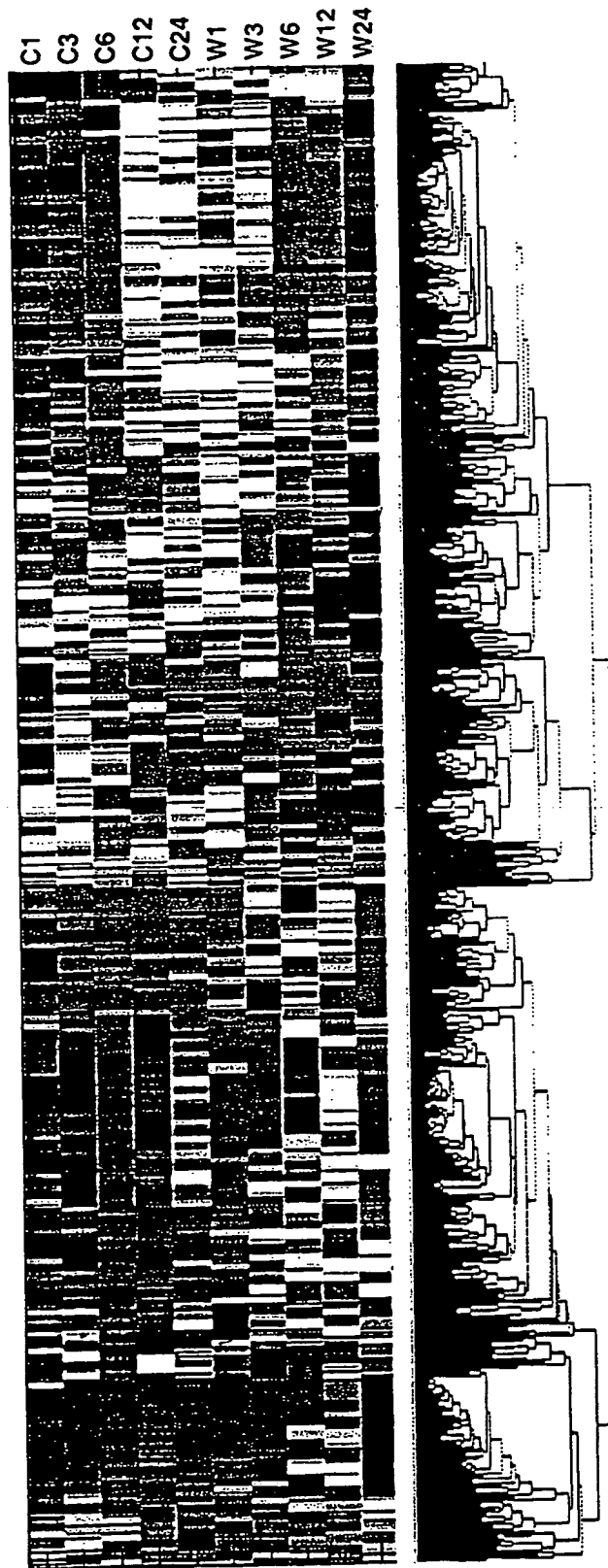


FIGURE 3A

B.

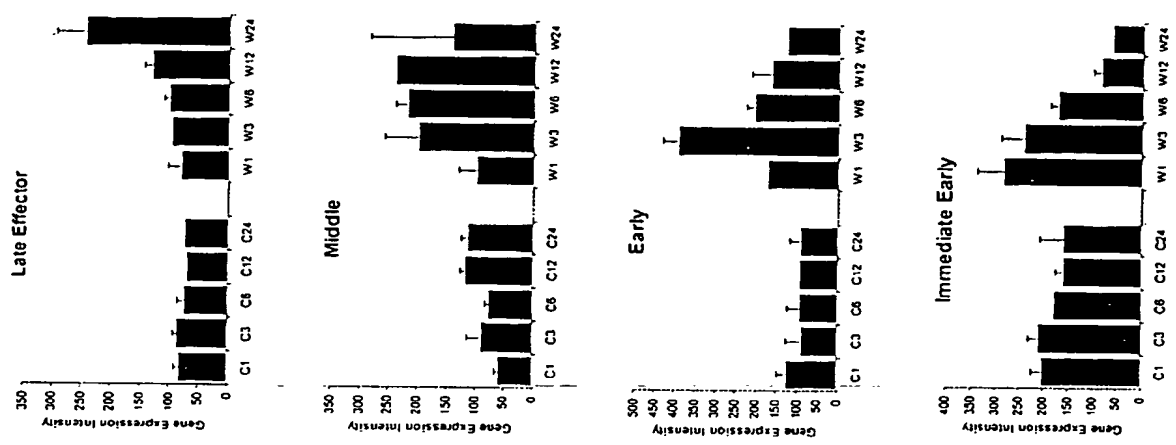
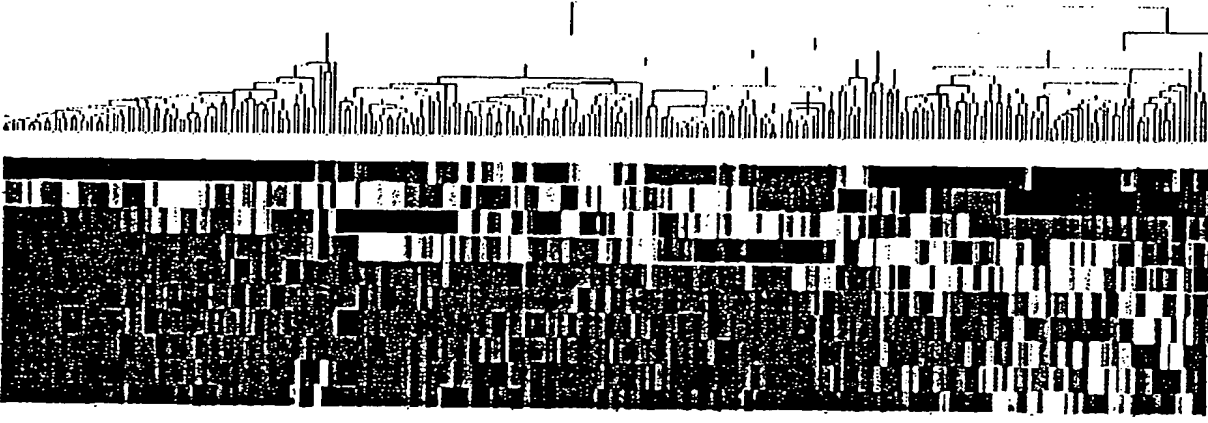
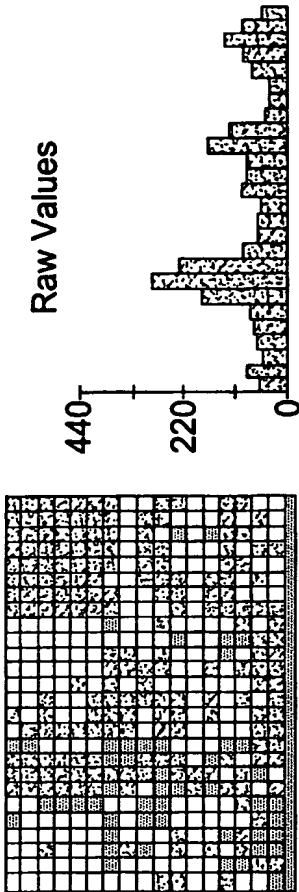
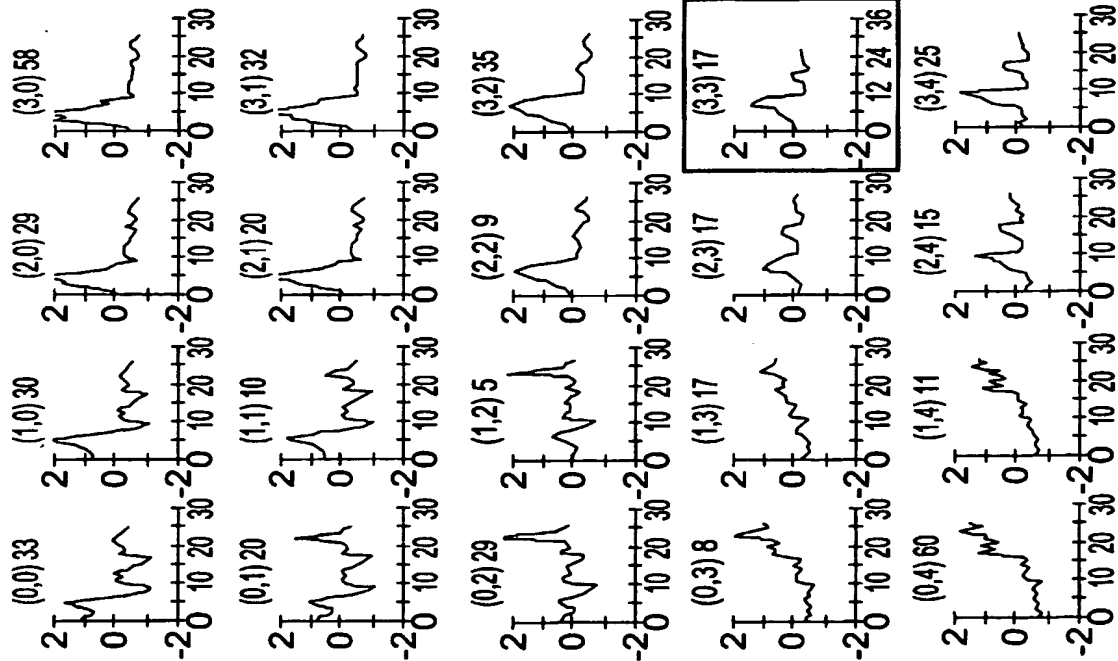


FIGURE 3B

SOM expression clusters.



Raw Values

440
220
0

Distance

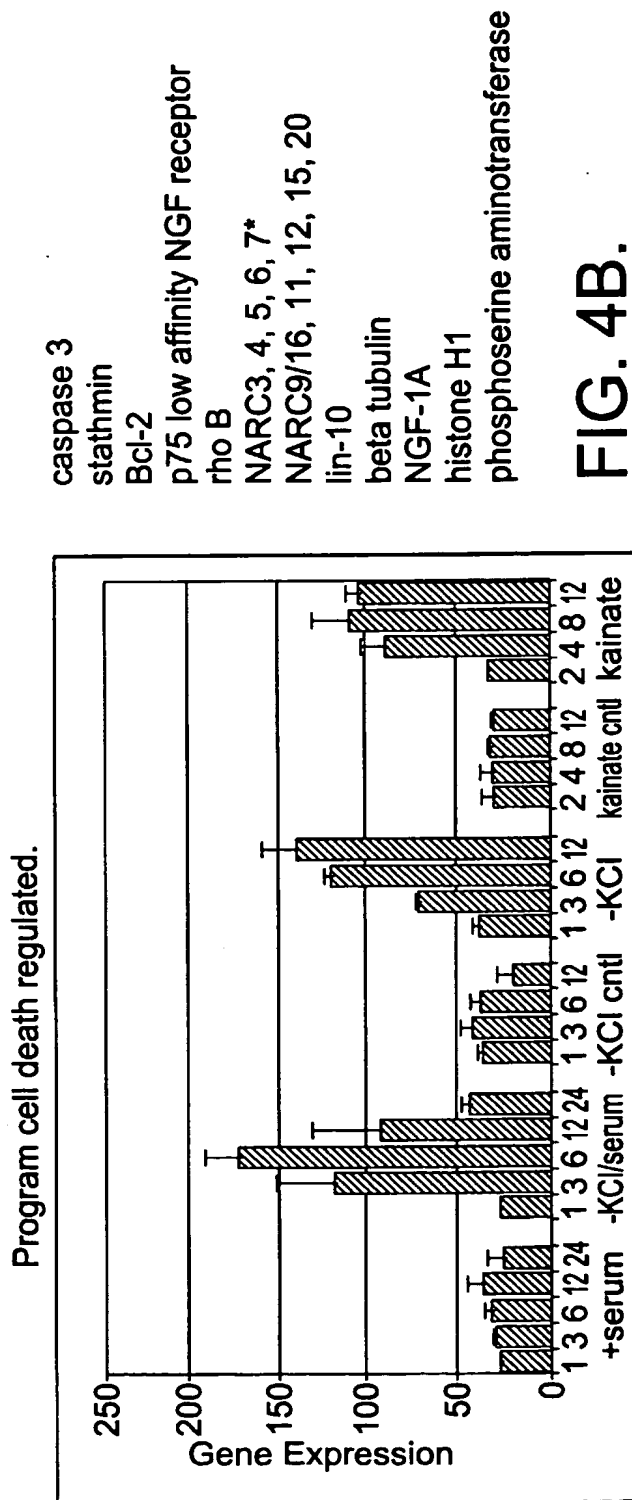
2.395
2.470
2.626
2.635
2.649
2.784
3.013
3.257
3.273
3.509
3.509
3.654
3.712
3.779
3.872
4.368
4.492

Array element

Wip 1
NARC2
NARC1
NARC1
NARC1
NARC1
NARC1
jtrba021c10t1
NARC2
jtrba049c04t1
jlrxa057d03t1
caspase 3*
histone H1
caspase 3
histone H1
jtrba033c06t3
jlrxa026e08t1
jtrba080a08t1

FIG. 4A.

6/31



SUBSTITUTE SHEET (RULE 26)

7/31

Regulated by KCl withdrawal only.

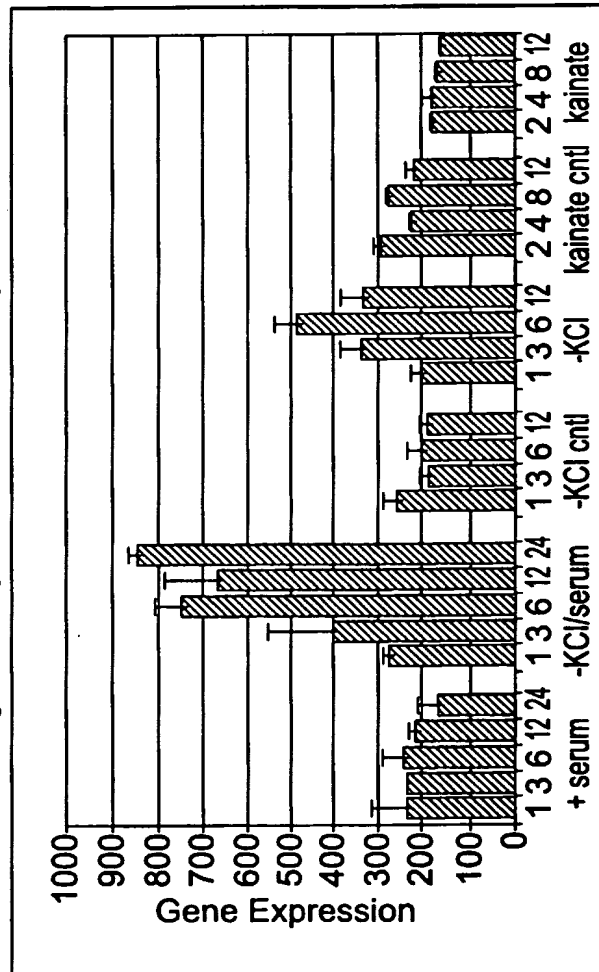


FIG. 4C.

8/31

Immediate early genes.



FIG. 4D.

9/31

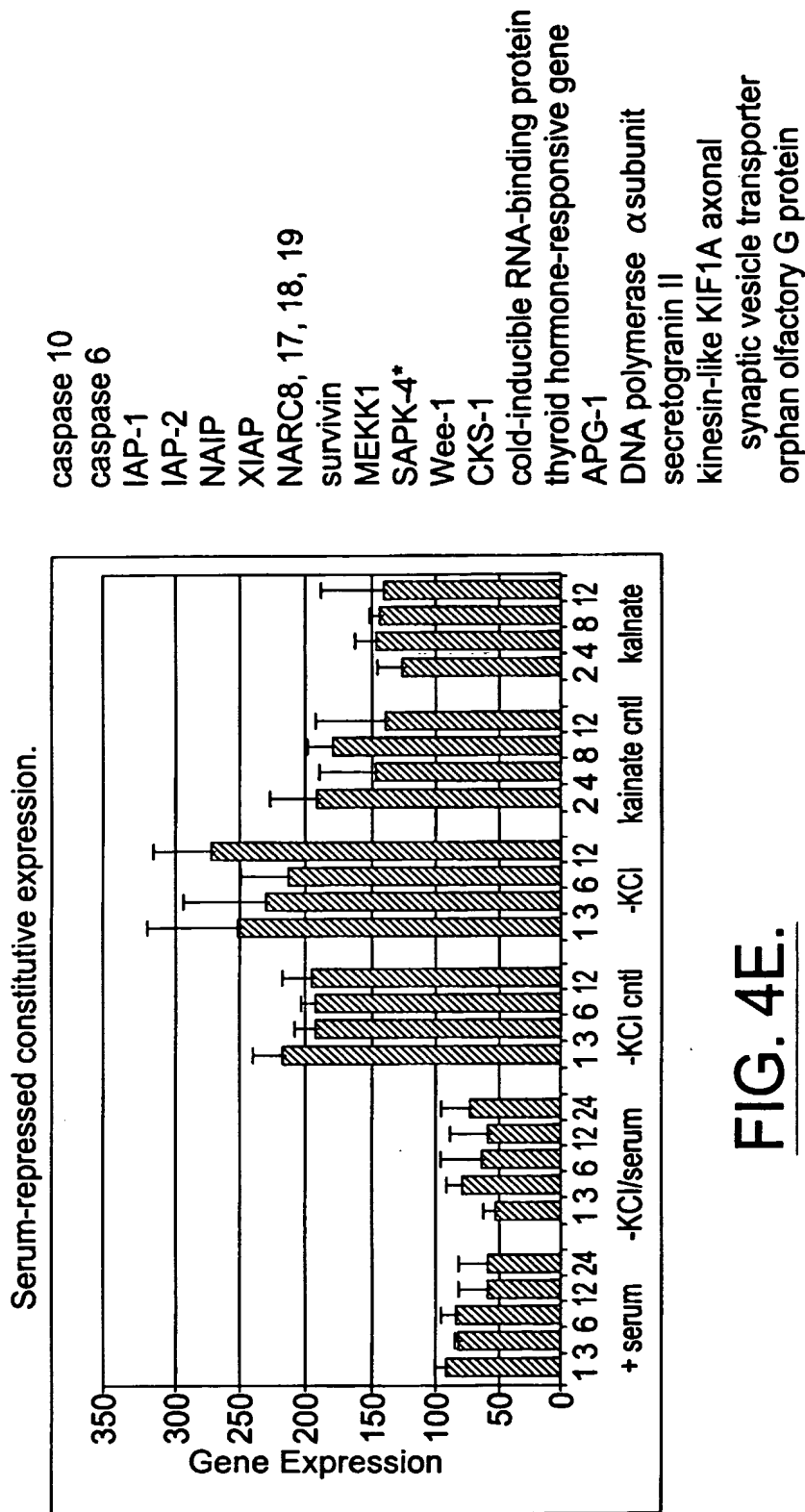


FIG. 4E.

10/31

NARC summary		Gene Expression Class	MTN	original EST Clone
Gene	Sequence info			
NARC1	Rat <i>frb</i> NARC1: 1057 nt 3'-UTR.	KCl-regulated	3.4 kb liver testes	rtx020f06b1
NARC2	Rat <i>AlxNARCxc1</i> : 2019 nt; >516 aa CRF that extends upstream of 5' end of sequence. Human <i>flhsNARC2A</i> : 1664 nt; >310 aa ORF that extends upstream of 5' end of sequence. Novel with no apparent homology to any known protein.	KCl-regulated	2.3 kb ubiquitous	rtx022h07b1
NARC3	Rat <i>frb</i> NARC3: 637 nt; >94 aa ORF that extends upstream of 5' end of sequence. Novel with no apparent homology to any known protein.	PCD early		rtx024h09a1
NARC4	Rat <i>frb</i> NARC4: 1247 nt 3'UTR.	PCD intermediate	2.2, 1.8, 1.1 kb ubiquitous	jtrxa012h0411
NARC5	Rat <i>frb</i> NARC5: 1837 nt; 129 aa ORF. Novel protein in a family including NARC6 containing coil-coil prot-prot domain and transcription factor motif. Homology with NARC6 extends upstream of apparent first ATG suggesting that the cDNA we have sequenced contains a nonsense mutation upstream of the apparent first ATG.	derepressed in low serum PCD intermediate	1.35 & 1.1 kb testes	jtrba034g08h3
NARC6	Rat <i>frb</i> NARC6: 1042 nt; 221 aa ORF. Novel protein in a family including NARC5 containing good coil-coil prot-prot domain and transcription factor motif.	derepressed in low serum PCD intermediate; ischemia 16/06=3.1	1.35 & 1.1 kb testes+++ brain+	jtrba118a12h1
NARC7	Rat <i>frb</i> NARC7: 2809nt.	PCO intermediate	3.2 kb in brain kidney testes: 1.35 kb ubiquitous	jtrxa027h0511
NARC8	Rat <i>frb</i> NARC8: 1475 nt; 298 aa ORF. Human <i>flhb</i> NARC8B: 1390 nt; >378 aa ORF that extends upstream of 5' end of sequence. Novel splice variant of rat nuclear receptor binding factor 1 that produces a truncated form of the protein.	derepressed in low serum	3.0 kb in heart brain skeletal: 1.35 kb in testes	jtrba022a0511
NARC9	Rat <i>frb</i> NARC9: 2393 nt; 249 aa ORF. Novel protein in family with NARC16 that has 50% identity, 65% similarity to C.elegans glycerophosphoryl diester phosphodiesterase and yeast Pho85.	PCD early	7.0 kb in brain: 5.0 kb in brain heart spleen lung skeletal testes (liver-may be bubble artifact)	jtrba022a0511
NARC10	Rat <i>frb</i> NARC10A: 1791 nt; 155 aa ORF. Human <i>find</i> NARD10C: 2034 nt; 183 aa ORF of human homolog. Novel protein with 50% identity, 60% similarity to neuron-specific nucleosome assembly protein	KCl-regulated	2.4 & 2.0 kb brain +++ heart-	jtrba033c06h3

TO FIG. 5B.

FIG. 5A.

11/31

FROM FIG. 5A.

NARC11	Rat <i>frb</i> NARC11: 1175 nt; ORF that extends upstream of 5' end of sequence with structure proposed in aa sequence figure. Novel splice variant of tyrosine phosphatase.	PCD early	3.5 kb in heart lung liver, 2.0 in spleen testes	<i>jtrba</i> 124g04t1
NARC12	Rat <i>frb</i> NARC12: 2250 nt. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	PCD early (less robust in defined medium)	4.2 kb brain	<i>jtrba</i> 031b03t1
NARC13	Rat <i>frb</i> NARC13: 2046NT; 151 aa ORF. Novel with no apparent homology to any known protein.	KCl regulated		<i>jtrba</i> 031h11t1
NARC14	Rat <i>frb</i> NARC14A: 863 nt; >74 aa ORF that extends upstream of 5' end of sequence. Novel protein similar to human AA316883.	KCl-regulated	1.6 kb in brain; 1.2 ubiquitous	<i>jtrba</i> 011c02t1
NARC15	Rat <i>frb</i> NARC15: 2485 nt 3'UTR. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	PCD down-regulated		<i>jtrba</i> 018g08t1
NARC16	Rat <i>frb</i> NARC16: 3381 nt; 626 aa ORF. Human <i>frb</i> NARC168: 3206 nt; 673 aa ORF. Novel protein that is similar to C. elegans glycerophosphoryl diester phosphodiesterase and is larger splice variant of NARC9; in particular, homology is to starch binding motif.	PCD early	4.6 & 3.4 kb skeletal heart brain testis	<i>jtrba</i> 043g05t1
NARC17	Rat <i>frb</i> NARC17: 988 nt 3'UTR.	derepressed in low serum		<i>jtrba</i> 045c08t1
NARC18	Rat <i>jtrba</i> 013h07t1: 263 nt. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	derepressed in low serum	6.3, 4.6, 3.0, 2.4 kb in brain; 1.2 kb in testes; 1.0 kb in liver testes	<i>jtrba</i> 013h07t1
NARC19	Rat <i>frb</i> NARC19: 1596 nt; 150 aa ORF. Human homolog is epididymal secretory protein.	derepressed in low serum	1.35 kb ubiquitous	<i>jtrba</i> 078d11t1
NARC20	Rat <i>frb</i> NARC20: 3934 nt. Pre-mRNA of myelin basic protein (probably intronic sequence followed by 50 nt of MBP coding region and 3' UTR).	derepressed in low serum; PCD intermediate	2.8 kb brain	<i>jtrba</i> 119f08t1
NARC21	Rat <i>jtrba</i> 025e06t1: 1435 nt rat contig encodes KIAA0863, 46% identity, 71% similarity to mouse Activity-Dependent Neurotrophic Factor.	KCl/serum-regulated only	ND	<i>jtrba</i> 025e06t1
NARC22	Rat <i>jtrba</i> 109f02t1: 1749 nt 3'UTR detected in rat config.	serum-regulated		<i>jtrba</i> 109f02t1
NARC23	Rat <i>jtrba</i> 087f11t1: 724 nt 3'UTR.	PCD intermediate (very low expression)		<i>jtrba</i> 087f11t1

TO FIG. 5C.

FIG. 5B.

12/31

FROM FIG. 5B.

NARC24	Rat jtrxa009d0811: >385aa ORF in rat config (that extends past 5' end. 65% identity, 78% similarity to novel C.elegans ORF.	PCD early	2.5 kb brain	jtrxa009d0811
NARC25	Rat ftrbNARC25: 1053 nt; >49 aa ORF that extends upstream of 5' end of sequence. Novel protein that is similar to synaptotagmin and inositol polyphosphate binding protein.	FN modulated ischemia-induced	4.8 kb brain	rtx017g03b1 rtx017h03b1 rtx017e03b1
NARC26	Rat ftrbNARC26: 1788 nt; 393 aa ORF. Novel with no apparent homology to any known protein.	FN modulated ischemia-induced	4.4 kb	jtrba057f0911
NARC27	Rat ftrbNARC27: 1403 nt; 429 aa ORF. Similar to "human secreted protein"; however, clone has much larger ORF with GTPase activation domain; longer Orf does not have signal peptide sequence; homology to centaurin.	FN modulated ischemia-induced	2.6 & 2.0 kb ubiquitous	jtrba094b041
NARC28	Rat ftrbNARC28: 1298 nt; >80 aa ORF that extends upstream of 5' end of sequence. Novel with no apparent homology to any known protein.	FN modulated ischemia-induced	6.4 kb in brain heart kidney; 4.5 kb in heart brain lung kidney testes	jtrba111f05t
NARC30	Rat ftrbNARC30: 4015 nt; >1212 aa ORF that extends upstream of 5' end of sequence. Similar to cell-death regulated secreted protein CW976_1; clone has a much larger ORF.	PCD early	4.2 kb brain heart lung skeletal kidney testes	jtrba057e0111
NARC31			2.3 & 1.8 kb	jtrxa001g0812

FIG. 5C.

13/31

Clone	Comment	Expression pattern
jtrb073h10t1	activin receptor type II	induced by KCl-withdrawal
jtrba001e02t1	ApoE, apolipoprotein E	induced by serum-withdrawal
jtrxa057f10t1	A-Raf proto-oncogene serine/threonine-protein kinase	constitutive
jtrxa055h12t1,	BAD	constitutive
mBcl-2	Bcl-2, mouse	induced by PCD
jthtb050a01	BclZ-2, human	basal expression repressed by serum-free conditions
jtrba012d09t1	BDNF	constitutive
jtrxa049d04t1	BMP-2, bone morphogenetic protein 2	constitutive
jtrba041d08t1,	BRCA2	constitutive
jthma018f01	caspase 10, human	basal expression derepressed by serum-free conditions
jtrba114c12t1,	caspase 6	basal expression derepressed by serum-free conditions
c-fos r,	c-fos	immediate early expression in all treatments
c-jun r	c-jun cpg	immediate early expression after KCl+serum withdrawal only
jfrxa024e05t1	CRES-RP, CREB related protein	induced by serum-addback, down-regulated by serum-withdrawal
jtrxa006e04t1	cyclin A	basal expression repressed by serum-free conditions
jtrxa012d04t1	cyclin B2	induced by kainate treatment
jtrxa039e08t1,	cyclin D-interacting myb-like protein	induced by kainate treatment
jtrba053h10t1,	(Dmp1)	
jtrxa006c09t1,	cyclin G, P53-activated	constitutive
jtrba106b06t1,	cyclin-dependent kinase 5 regulatory	constitutive
jtrba004g05t1	subunit p35 (Cdk5r)	
jtrba029b06t1	cyclin-dependent kinases regulatory subunit 1 (CKS-1)	induced by serum-withdrawal, down-regulated by serum-addback; basal expression derepressed by serum-free conditions
GAP43f.l.r	GAP 43	
jtrxa029h09t2,	Hsp 27	constitutive
jtrba086f03t1,	Hsp 60	constitutive
Hsp70cpg271	Hsp 70 cpg	immediate early expression after kainate treatment only; basal expression repressed by serum-free conditions
jtrxa001d09t2	Hsp 86, chaperonin	constitutive
jthqb017f10	IAP 1, human	basal expression derepressed by serum-free conditions
jthdc128h09t2	IAP-2, human	2 basal expression derepressed by serum-free conditions

FIG. 6A.

SUBSTITUTE SHEET (RULE 26)

14/31

Clone	Comment	Expression pattern
IGF	IGF, human	down-regulated by KCl-withdrawal; completely absent in kainate experiment
jtrxa006b09t1	IGF-II receptor	constitutive
jtrba071b09t1,	IL-6 receptor	constitutive
jtrba032a06t3	insulin receptor tyrosine kinase 53 kDa substrate	basal expression repressed by serum-free conditions
jtrxa001c01t2	insulin-induced growth response protein (CL-6)	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by
jtrba109c03t1	insulin-like growth factor II	constitutive
jtrba054b04t1	MAP kinase kinase	constitutive
jtrba096f01t1	MAP kinase kinase kinase 1 (MEKK1)	basal expression derepressed by serum-free conditions
erk2 h,	MAP kinase, human erk 2, p38	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by serum-free conditions
jtrba089d11t1	MAPKAP kinase, mitogen activated protein kinase activated protein kinase-3	constitutive
jtrxa027g07t1	MDM-2, p53 associated protein	constitutive
jthua018h08	NAIP	basal expression derepressed by serum-free conditions
p75 r	NGF receptor p75, low affinity	immediate early expression after induction of PCD
NGF m	NGF, mouse	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by serum-free conditions
jtrxa005d10t1	NGF-receptor, fast	constitutive
jtrxa001c03t2	p53	induced by serum-addback, down-regulated by
jtrba062e03t1	ras inhibitor	constitutive
jtrba002a07t1,	ras protein p21	constitutive
jtrba117b02t1	RB protein binding protein	constitutive
jfrxa024h04t1	RBP, retinoblastoma-binding protein (RbAp46)	constitutive
jtrba103c03t2	RBP1, retinoblastoma binding protein 1	
jtrxa047a06t1	RBP2, retinoblastoma binding protein 2	basal expression repressed by serum-free conditions
jtrxa007b03t1,	rho	constitutive
jtrba125h09t1	Rho GDP-dissociation inhibitor 1	induced by serum-withdrawal, down-regulated by serum-addback

FIG. 6B.

SUBSTITUTE SHEET (RULE 26)

15/31

Clone	Comment	Expression pattern
jtrxa027d05t1	RIP, nucleoporin-like protein (Rev interacting protein, Rev/Rex activation domain binding protein)	constitutive
jtrxa005c04t1	SKP1, cell-cycle control	
jtrxa038f09t1	SOD-1	constitutive
jtrba041d05t1	SOD-2	constitutive
jtrxa044d10t1	Ste-20 like kinase YSK-1	constitutive
jtrxa025c05t1	Ste20-like kinase, 54% identity, 73% identity to human Ste20-like kinase	constitutive
jtrxa013g03t1	stress-activated protein kinase 4	basal expression derepressed by serum-free conditions
jthta102e01	survivin, human	basal expression derepressed by serum-free conditions
jtrxa001d05t2	trk A	induced by serum-addback, down-regulated by serum-withdrawal
trkB r 2	trk B	basal expression repressed by serum-free conditions
jtrba002b11t1	trk C	constitutive
jtrxa018h09t2	wee 1 tyrosine kinase	basal expression derepressed by serum-free conditions
jchrb018f03	XIAP, human	basal expression derepressed by serum-free conditions

FIG. 6C.

Clone	Comments	16/31	SOM Id
	<u>transient down-regulation by serum, down-regulated by KCl withdrawal (secretion/synaptic vesicle release)</u>		
jtrxa046c07t1	synaptotagmin synaptic vesicle protein		(0,0)
jtrxa040b12t1	novel		(0,0)
jtrba016d09t1			(0,0)
jtrba010a11t1	rat inositol 1,4,5-triphosphate receptor		(0,0)
jtrba083g04t1	CDC10 homologue involved in cytokinesis		(0,0)
jtrba086f01t1	kynein intermediate chain		(0,0)
jtrxa011g07t1	fodrin (cytoskeletal component of Ca ⁺⁺ -secretion)		(0,0)
jtrxa060g03t1	(NAD ⁺) isocitrate dehydrogenase in mitochondria.		(0,0)
jtrxa055g08t1	neuroendocrine secretory protein 55 chromogranin family		(0,0)
jtrba028h11t1			(0,0)
jtrba005a01t1			(0,0)
jtrba061f02t1			(0,0)
jtrba039g01t1	Na ⁺ , K ⁺ -ATPase beta subunit		(0,0)
jtrba049a02t1	rat inositol 1,4,5-trisphosphate receptor		(0,0)
jtrba027c04t1			(0,0)
jtrba021f01t1	14-3-3 protein tau multifunctional regulator of CaMKII and PKC		(0,0)
jtrxa041a03t1	secretogranin neurendocrine secretory granule protein chromogranin family		(0,0)
jtrba010e10t1	succinyl-coa Sketoacid-coenzyme in mitochondria		(0,0)
jtrba035a02t1	synaphin 2 associated with docking/fusion complex		(0,0)
jtrxa038g09t1	novel		(0,0)
jtrba032c01t3	complexin cytosolic proteins that regulate SNAP receptor function		(0,0)
jtrxa032h06t1	cytochrome c in mitochondria		(0,0)
secretograninc	secretogranin		(0,0)
jtrba021e08t1			(0,0)
jtrba067h01t1			(0,0)
jtrba015a11t1	RIG (down-Regulated in Glioma)		(0,0)
jtrba012g02t1	alpha-tubulin-1		(0,0)
jtrxa027b05t1			(0,0)
p75 r	low affinity NGF receptor p75		(0,0)
jtrba043g04t1	DAP-5, a novel homolog of eukaryotic translation initiation factor 4G isolated as a putative modulator of gamma interferon-induced cell death		(0,0)
jtrba090c02t1			(0,0)
jtrba020h04t1			(0,0)
jtrxa027f11t1	novel		(0,0)
	<u>transient down-regulation by serum, down-regulated by KCl withdrawal and kainate treatment</u>		
rtrX024a09a1			(0,1)
rtrX024b09a1			(0,1)
jfrxa006g12t1	secretogranin I chromogranin family		(0,1)
jtrxa044c10t1			(0,1)
rtrX024c09a1			(0,1)
rtrX024d09a1			(0,1)
rtrX024h09a1			(0,1)

FIG. 7A.

SUBSTITUTE SHEET (RULE 26)

17/31

rtrX024f09a1		(0,1)
rtrX024g09a1		(0,1)
jtrba089e08t1		(0,1)
rtrS024e09a1		(0,1)
jtrba035d02t1	neurocalcin calcium-binding protein	(0,1)
jtrba009b09t2	novel	(0,1)
jtrba122c04t1	voltage-gated potassium channel KV1.4	(0,1)
jtrba025e08t1		(0,1)
rtrX023e08b1		(0,1)
jtrba078f10t1		(0,1)
jtrba050b02t1		(0,1)
jtrba090c03t1		(0,1)
jtrba125h10t1	novel	(0,1)
jtrba045f03t1		(0,2)
jtrxa052b09t1		(0,2)
jtrba049h12t1		(0,2)
jtrxa046g07t1		(0,2)
jtrba069a03t2		(0,2)
jtrxa044d10t1		(0,2)
jtrba034c03t3		(0,2)
jtrba069h07t2		(0,2)
jtrba032h02t3		(0,2)
jtrba024h09t1		(0,2)
jtrba076f12t2		(0,2)
jtrba034h07t3		(0,2)
jtrba029a05t1		(0,1)
jtrxa050b11t1		(0,2)
jtrxa031c06t1		(0,2)
jtrxa054e11t1		(0,2)
jtrba007g02t1		(0,2)
jtrba063b08t1		(0,2)
jtrxa046g08t1		(0,2)
jtrba041b03t1		(0,2)
jtrba005c05t1		(0,2)
jtrba083a05t1		(0,2)
jtrba054b06t1		(0,2)
jtrba011b09t1		(0,2)
	<u>immediate early effect after any perturbation</u>	
jtrba005h05t1	navel	(0,2)
jtrba111e08t2	c-fos	(0,2)
jtrxa055b02t1		(0,2)
c-fos r	c-fos	(0,2)
jtrba018g08t1	novel EST from PC12 NGF-deprived cDNA library (1996, PNAS 92:8303)	(0,2)
	<u>serum-repressed genes</u>	
jtrba060c09t1	(cell proliferation)	(0,3)
jtrxa033d10t1		(0,3)
jtrba093g06t1		(0,3)

FIG. 7B.

SUBSTITUTE SHEET (RULE 26)

18/31

jtrba103b07t2	transferrin transports iron from sites of absorption and heme degradation to those of storage and utilization; serum tranferrin may have a role in stimulating cell proliferation	(0,3)
jtrxa035d07t1		(0,3)
jtrxa015g08t1	novel	(0,3)
jtrxa045g12t1		(0,3)
jtrba005d06t1		(0,3)
jtrba046c05t1	<u>serum and KCl regulated</u> (secretion/synaptic residue released cytoskeletal reorganization) calcium-dependent actin-binding protein; activator protein for secretion/e)exocytosis	(1,0)
jtrxa030e12t1	microtubule-associated protein (MAP1B)	(1,0)
jtrba013g10t1	novel	(1,0)
jtrxa010h04t1	novel	(1,0)
jtrba032f05t3	Vap-33, vesicle-associated membrane binding protein; synaptobrev n binding protein	(1,0)
jtrba057g11t1	GABA-A receptor delta subunit	(1,0)
jtrxa022g12t2	novel	(1,0)
jtrba063e11t1		(1,0)
jtrba033b07t4	novel	(1,0)
jtrxa058h01t1	63% identical, 74% similar to mouse N-myo-downstream gene Ndr1	(1,0)
jtrxa005c02t1	novel	(1,0)
jtrba104d03t1	novel	(1,0)
jtrxa035f07t1	microtubule associated protein MAP-1b	(1,0)
jtrba057d04t1	plasma membrane Ca2+ATPase-isoform 2	(1,0)
jtrxa049g12t1	novel	(1,0)
jtrba006e10t1	CaMKIIbeta	(1,0)
jtrba009d10t2		(1,0)
jtrba036h03t2	myosin heavy chain	(1,0)
jtrba041h10t1	synapsin Ib	(1,0)
jtrba025h11t1		(1,0)
jtrba035g12t2		(1,0)
jtrba001a08t1	bovine cytochrome b-560, succinate-ubiquinone reductase OPs1	(1,0)
jtrxa056g12t1		(1,0)
jtrxa058a10t3	alpha tubulin	(1,0)
jtrxa058a10t3	alpha tubulin	(1,0)
jtrxa024f05t1	phosphofuctokinase C	(1,0)

FIG. 7C.

SUBSTITUTE SHEET (RULE 26)

19/31

jtrxa038e03t1	novel	(1,0)
jtrxa024b07t1	alpha tubulin	(1,0)
jtrxa024g09t1		(1,0)
	<u>cell death regulated</u>	
jtrba027g06t4	vesicle-associated calmodulin-binding kinase-like protein	(1,1)
jtrba040h07t1	beta-synuclein	(1,1)
jtrxa037f09t1	DNA replication licensing factor required for the entry into S phase and for cell division	(1,1)
jtrba044e03t1		(1,1)
jtrba057e01t1		(1,1)
jtrxa049c07t1	mammalian homolog of Drosophila circadian Perigene	(1,1)
jtrba109f02t1		(1,1)
jtrxa009d08t1		(1,1)
GluR5 r	GluR5	(1,1)
jtrba116b12t1	Hsp60, involved in mitochondrial protein import and macromolecular assembly; may prevent misfolding and promote refolding and proper assembly under stress conditions in the mitochondrial matrix	(1,1)
jtrba031h09t3		
jtrba009h10t2		
jtrxa039e01t1		
jtrba036e10t1		
jtrba035f12t2		
	<u>derepressed genes after growth in defined medium</u>	
jtrba119f08t1	novel (stress response/hormone response)	(1,3)
jtrba116h12t1	olfactory G protein involved in visual transduction and in mediating the effect of one or mor hormones/neurotransmitters	(1,3)
jtrba023d09t1	DNA polymerase alpha subunit	(1,3)
jtrxa040h01t1	cold-inducible RNA-binding protein mediating cold-inducible suppression (of mammalian cell growth; expressed transiently in developing neural tissue; LV-inducible-cDNA	(1,3)
jthma018t01	caspase 10	(1,3)
jtrxa009f03t1	novel	(1,3)
jtrba078d11t1		(1,3)
jtrxa045c08t1		(1,3)
jtrba016g02t1	thyroid hormone-responsive gene; gene from the Down syndrome optical region resembling a DNA binding protein with an SH3 domain	(1,3)
jtrba002f04t1		(1,3)
jtrba029e02t1		(1,3)
jtrba114c07t1	secretogranin II	(1,3)
jtrba124f02t1	GABA-A receptor alpha-5 subunit	(1,3)
jtrba013h07t1		(1,3)
jtrxa059d02t1		(1,3)
jtrxa053a03t1	lin-10	(1,3)
jtrba075f03t1		(1,3)
	<u>derepressed genes after growth in defined medium</u>	
jtrba034g06t3	novel (stress response/hormone response)	(1,4)

FIG. 7D.

SUBSTITUTE SHEET (RULE 26)

20/31

jtrba109c06t1		(1,4)
jtrba053b09t1		(1,4)
jtrxa032g02t1	novel	(1,4)
jtrba018b01t1	Hsp70-related protein APG-1, induction by hyperosmolar salt stres:	(1,4)
jtrxa026f02t1		(1,4)
jfrxa044a11t1		(1,4)
jtrba118a12t1	novel	(1,4)
jtrxa006b04t1	novel	(1,4)
jtrba068f01t1	progesterone-induced messenger RNA resembling phosphoserine amino transferase required in major phosphorylated pathway of serine and pyridaxine biosynthesis	(1,4)
jtrxa013f01t1	progesterone-induced messenger RNA resembling phosphoserine amino transferase required in major phosphorylated pathway of serine and pyridaxine biosynthesis	(1,4)
<u>Serum and KCl regulated</u>		
jtrba105c11t1	(calcium signal transduction)	(2,0)
SV2a r 1	SV2a	(2,0)
jtrba017f06t1	cytosolic aspartate aminotransferase	(2,0)
jtrba001d01t1	novel (jtrba005f12t1)	(2,0)
jtrba077g09t1		(2,0)
CaMKIIb r	CaMKII beta	(2,0)
SV2a r 2	SV2a	(2,0)
CaMKIIdC r	CaMKII delta C	(2,0)
jtrba005f12t1	novel (jtrba001d01t1)	(2,0)
jtrxa045c04t1	14-3-3 protein gamma-subtype; regulation of protein kinase C	(2,0)
jtrba045c03t1	citrate synthase in mitochondrial matrix	(2,0)
jtrba027h11t4	novel SH3 containing protein	(2,0)
jtrba022g04t1	novel	(2,0)
jtrba075h01t1	vacuolar adenosine triphosphatase subunit responsible for acidifying a variety of intracellular compartments	(2,0)
jtrba073h12t1	14-3-3 protein gamma-subtype; regulation of protein kinase C	(2,0)
jtrba073h12t1	calcium-independent alpha-latrotoxin receptor (secretin GPCR family:	(2,0)
jtrba115h04t1	synapsin 1	(2,0)
jtrxa001e02t2		(2,0)
jtrba091c06t1		(2,0)
jtrba087e08t1	novel	(2,0)
NGFIB r	NGF-lb	(2,0)
jtrba013b12t1	FUSE binding protein that activates the far upstream element of c-my:	(2,0)
jtrba029a12t1	novel	(2,0)
jtrxa059c04t1	testosterone induced splicing factor SF1	(2,0)
jtrxa019a09t2	TCP-1 chaperonin	(2,0)

FIG. 7E.

21/31

jtrba028g11t1	Stathmin, a ubiquitous phosphoprotein acting as an intracellular re ay of proliferation and differentiation	(2,3)
jtrxa021g01t1	rhoB ras-like immediate early gene	(2,3)
jtrba043g05t1	novel	(2,3)
jtrba022a05t1		(2,3)
jtrba018b03t1		(2,3)
jtrba014b01t1		(2,3)
jtrba043c06t1		(2,3)
jtrba124g12t1	beta tubulin-like protein	(2,3)
jtrba087f11t1		(2,3)
jtrba115d02t1	kinesin-like KIF1A axonal transporter of synaptic vesicles	(2,3)
	Stable transcripts	
<i>jtrba034e12t3</i>	<i>(survival factors)</i>	(2,4)
<i>jtrxa054d03t1</i>		(2,4)
<i>jtrxa001e10t2</i>		(2,4)
<i>jtrxa006h05t1</i>		(2,4)
<i>jtrba108b01t1</i>	<i>vimentin</i>	(2,4)
<i>jtrba081c02t1</i>		(2,4)
<i>jtrba113b04t1</i>		(2,4)
<i>jtrba112a12t1</i>		(2,4)
<i>jtrba086b12t1</i>		(2,4)
<i>jtrba023d12t1</i>		(2,4)
<i>jtrba005c11t1</i>		(2,4)
<i>jtrba117c06t1</i>		(2,4)
<i>jtrba005f07t1</i>		(2,4)
<i>jtrxa013g09t1</i>		(2,4)
<i>jtrxa045f01t1</i>		(2,4)
<i>Synaptophysin</i>	<i>synaptophysin</i>	(3,0)
<i>jtrxa045h03t1</i>	<i>synaptotagmin XI</i>	(3,0)
<i>p39</i>	<i>p39</i>	(3,0)
<i>jtrba002e06t1</i>	<i>clathrin-coated vesicle/synaptic vesicle proton pump 116 subunit vaccolar proton pump subunit 1</i>	(3,0)
<i>jtrba084a04t1</i>	<i>novel</i>	(3,0)
<i>N-sec 1 r</i>	<i>syntaxin-binding protein (n-sec1)</i>	(3,0)
<i>jtrba031d04t3</i>	<i>novel</i>	(3,0)
<i>jtrba060c03t1</i>	<i>sodium/potassium-transporting ATPase alpha-3 chain</i>	(3,0)
<i>fth286t</i>		(3,0)
<i>jtrba109d02t1</i>		(3,0)
<i>jtrxa001d05t2</i>	<i>trk A</i>	(3,0)
<i>rlrX019c05a1</i>		(3,0)
<i>jtrba037a07t1</i>	<i>sodium/potassium-transporting ATPase alpha-3 chain</i>	(3,0)
<i>jtrba057f09t1</i>		(3,0)
<i>jtrxa007f11t1</i>	<i>syntaxin-binding protein (n-sec1)</i>	(3,0)
<i>jtrxa044b01t1</i>	<i>novel</i>	(3,0)
<i>jtrba030g12t1</i>	<i>novel</i>	(3,0)
<i>jtrba048f02t1</i>	<i>novel</i>	(3,0)
<i>PSD-95 r</i>	<i>PSD-95</i>	(3,0)
<i>jtrxa054b03t1</i>	<i>novel</i>	(3,0)
<i>jtrba013a08t1</i>		(3,0)

FIG. 7F.

SUBSTITUTE SHEET (RULE 26)

22/31

<i>jtrba036a02t2</i>	(3,0)
<i>jtrba041a02t1</i>	(3,0)
<i>jtrba083f09t1</i>	(3,0)
<i>jtrxa056e08t1</i>	(3,0)
<i>jtrxa033f03t1</i>	(3,0)
<i>jtrba053d01t1</i>	(3,0)
<i>jtrba017b08t1</i>	(3,0)
<i>jtrxa046e09t1</i>	(3,0)
<i>jtrba055g03t1</i>	(3,0)
<i>jtrba049b09t1</i>	(3,0)
<i>jtrba094b12t1</i>	(3,0)
<i>jtrxa034e05t1</i>	(3,0)
<i>jtrba109c11t1</i>	(3,0)
<i>jtrba038h12t1</i>	(3,0)
<i>rtrX017e03b1</i>	(3,0)
<i>jtrba086f08t1</i>	(3,0)
<i>jtrxa004b09t1</i>	(3,0)
<i>jtrba019b02t1</i>	(3,0)
<i>jtrba019g02t1</i>	(3,0)
<i>jtrba019f12t1</i>	(3,0)
<i>jtrba117h02t1</i>	(3,0)
<i>rtrX017c03b1</i>	(3,0)
<i>rtrX017f03b1</i>	(3,0)
<i>jtrxa008c04t1</i>	(3,0)
<i>rtrX017g03b1</i>	(3,0)
<i>jtrxa005h09t1</i>	(3,0)
<i>jtrba029d07t1</i>	(3,0)
<i>rtrX015b01a1</i>	(3,0)
<i>jtrxa050h03t1</i>	(3,0)
<i>rtrX015a01a1</i>	(3,0)
<i>jtrba118c02t1</i>	(3,0)
<i>jtrba109f08t1</i>	(3,0)
<i>jtrba083c03t1</i>	(3,0)
<i>jtrba035h01t1</i>	(3,0)
<i>jtrba095g06t1</i>	(3,0)
<i>rfrX015g01a1</i>	(3,0)
<i>jtrba038a08t1</i>	(3,0)
<i>jtrba084b04t1</i>	(3,1)
<i>jtrba084h08t1</i>	(3,1)
<i>jtrba013a09t1</i>	(3,1)
<i>rGFAP 5'1/10 GFAP</i>	(3,0)
<i>RCSP r RGSP</i>	(3,1)
<i>jtrba084b09t1</i>	(3,1)
<i>jtrba045g09t1</i>	(3,0)
<i>syntaxin2 r</i>	(3,1)
<i>r Actin 5' beta actin</i>	(3,1)
<i>jtrxa011f11t1</i>	(3,1)
<i>jfrxa001c12t2</i>	(3,1)
<i>jtrxa038f02t1</i>	(3,1)

FIG. 7G.

SUBSTITUTE SHEET (RULE 26)

23/31

<i>rGFAP 5' RT-1 GFAP</i>	(3,1)
<i>r Actin 5' 1/10 beta actin</i>	(3,1)
<i>jtrba069b12t2</i>	(3,1)
<i>jtrba108b06t1</i>	(3,1)
<i>r GFAP 5' GFAP</i>	(3,1)
<i>jtrba046g08t1</i>	(3,1)
<i>jtrba022f09t1</i>	(3,1)
<i>jtrba085g11t1</i>	(3,1)
<i>jtrba073d05t1</i>	(3,1)
<i>jtrba054g03t1</i>	(3,1)
<i>r GAPDH glyceraldehyde-3-phosphate-dehydrogenase</i>	(3,1)
<i>jtrba063h10t1</i>	(3,1)
<i>erk2 h erk 2</i>	(3,1)
<i>jtrxa043b04t1</i>	(3,1)
<i>jtrxa050f07t1</i>	(3,1)
<i>jtrba021f10t1</i>	(3,1)
<i>rtrX017h03b1</i>	(3,1)
<i>jtrba038f01t1</i>	(3,1)
<i>jtrba078f09t1</i>	(3,1)
<i>r GFAP M RT GFAP</i>	(3,2)
<i>jtrba025g01t1</i>	(3,2)
<i>rGFAP M 1/10 GFAP</i>	(3,2)
<i>jtrxa005b01t1</i>	(3,2)
<i>jtrba100e12t1</i>	(3,2)
<i>rGFAP M GFAP</i>	(3,2)
<i>rtrX018h04a1</i>	(3,2)
<i>r Actin M 1/10 beta actin</i>	(3,2)
<i>jtrxa004d06t1</i>	(3,2)
<i>jtrxa030b02t1</i>	(3,2)
<i>jtrba112h05t1</i>	(3,2)
<i>jtrba120f04t1</i>	(3,2)
<i>jtrba032e04t3</i>	(3,2)
<i>jtrba118d04t1</i>	(3,2)
<i>r Actin M beta actin</i>	(3,2)
<i>jtrxa052f02t1</i>	(3,2)
<i>jtrba112h06t1</i>	(3,2)
<i>jtrba007a11t1</i>	(3,2)
<i>jtrba125b04t1</i>	(3,2)
<i>r 18S rRNA 18S rRNA</i>	(3,2)
<i>jtrba057e05t1</i>	(3,2)
<i>r nEnolase neuronal enolase</i>	(3,2)
<i>jtrxa045d02t1</i>	(3,2)
<i>jtrxa033a12t1</i>	(3,2)
<i>r GFAP 3' GFAP</i>	(3,2)
<i>jtrxa011c11t1</i>	(3,2)
<i>jtrba045d09t1</i>	(3,2)
<i>rGFAP 3' 1/10 GFAP</i>	(3,2)
<i>rGFAP 3' RT-1 GFAP</i>	(3,2)
<i>rGFAP 3'-1/10 GFAP</i>	(3,2)
<i>jtrxa007b04t1</i>	(3,2)

FIG. 7H.

24/31

<i>PC12-PTP1 r PC12-PTP1</i>	(3,2)
<i>jtrba087b01t1</i>	(3,2)
<i>jtrba061c01t1</i>	(3,2)
<i>jtrba079h08t2</i>	(3,2)
<i>KCl withdrawal unregulated</i>	
<i>rtrX019b05a1</i> <i>protein phosphatase Wip-1 (apoptosis)</i>	(3,3)
<i>rtrX022h07b1</i>	(3,3)
<i>rtrX020b06b1</i>	(3,3)
<i>rtrX020f06b1</i>	(3,3)
<i>rtrX020e06b1</i>	(3,3)
<i>rtrX020c06b1</i>	(3,3)
<i>jtrba021c10t1</i> <i>novel</i>	(3,3)
<i>rtrX022f07b1</i>	(3,3)
<i>jtrba049c04t1</i> <i>nucleolar ribosomal RNA precursor external transcribed spacer 1 (ETS1)</i>	(3,3)
<i>jtrxa057d03t1</i>	(3,3)
<i>rtrX030f10a1</i> <i>caspase 3</i>	(3,3)
<i>jtrxa057a03t1</i> <i>histone H1</i>	(3,3)
<i>rtrX030b10a1</i> <i>caspase 3</i>	(3,3)
<i>rtrX015h01a1</i>	(3,3)
<i>jtrba033c06t3</i> <i>novel</i>	(3,3)
<i>jtrxa026e08t1</i> <i>novel</i>	(3,3)
<i>jtrba080a08t1</i>	(3,3)
<i>Stable transcripts</i>	
<i>(survival factors)</i>	
<i>jtrba011c02t1</i>	(3,4)
<i>jtrba035f05t1</i>	(3,4)
<i>jtrxa004c06t1</i>	(3,4)
<i>jtrba076h01t2</i>	(3,4)
<i>jtrxa010b09t1</i>	(3,4)
<i>jtrba119b09t1</i>	(3,4)
<i>jtrxa001f09t2</i>	(3,4)
<i>jtrxa015d09t1</i>	(3,4)
<i>jtrba066f05t2</i>	(3,4)
<i>jtrxa004f04t1</i>	(3,4)
<i>jtrba068e03t1</i>	(3,4)
<i>jtrxa006f10t1</i>	(3,4)
<i>jtrxa005e05t1</i>	(3,4)
<i>jtrba065f07t1</i>	(3,4)
<i>jtrba032b05t3</i>	(3,4)
<i>jtrxa041d11t1</i>	(3,4)
<i>jtrxa004e01t1</i>	(3,4)
<i>jtrba098h06t1</i>	(3,4)
<i>jtrba019f07t1</i>	(3,4)
<i>jtrxa005h03t1</i>	(3,4)
<i>jtrxa003a01t1</i>	(3,4)
<i>jtrba120d07t1</i>	(3,4)
<i>jtrba011b04t1</i>	(3,4)
<i>jtrba115b01t1</i>	(3,4)
<i>jtrba009a07t2</i>	(3,4)

FIG. 7I.

SUBSTITUTE SHEET (RULE 26)

25/31

Tissue distribution of rat brain EST array elements.

Category/Tissue	# Array elements detected
Total rat brain ESTs arrayed	7296
Any tissue	6127
Brain	5329
Heart	4525
Kidney	2686
Liver	1658
Lung	2620
Pancreas	1365
Skeletal muscle	1861
Smooth muscle	2714
Spleen	3921
Testes	3545
Brain not heart	1000
Brain only	582
All tissues	1169

FIG. 8.

26/31

Table 2. Smart Chip Genes regulated by KCl/Serum-withdrawal.

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
nexin 1	Late	glia-derived protease that is a neurite-promoting factor	P07092	jtrba019f07t1
SPARC	Late	Secreted Protein Acidic and Rich in Cysteine (regulates cell growth through interactions with the extracellular matrix and cytokines)	P16975	jtrba034c12t3
β -2-microglobulin	Late	precursor of major histocompatibility complex class I molecules (Fc receptor)	P07151	jlrxa006h05t1
GAD	Late	glutamic acid decarboxylase (catalyzes the production of inhibitory neurotransmitter GABA).	P18088	jtrba120d01t1
PAF A	Late	platelet activating factor acetylhydrolase (modulates action of PAF)	Q60963	jtrba028a03t1
intestinal membrane A4 protein	Late	possible differentiation role in intestinal epithelium (gene localizes near synaptophysin locus)	Q04941	jlrxa003a01t1
glutathione S-transferase Yb-1	Late	conjugation of reduced glutathione to exogenous and	M11719	jlrxa001f09t2
myoblast cell surface antigen	Late	myoblast cell surface antigen	X16850	jlrxa004e01t1
Na ⁺ -dep. glutamate/aspartate transporter	Late	essential for terminating the postsynaptic action of excitatory neurotransmission by rapid removal of glutamate from the synaptic cleft	S75687	jtrba014e07t3
ATP citrate-lyase	Late	primary enzyme for synthesis of cytosolic acetyl-coa and <i>de novo</i> lipid synthesis in neurons	P16638 P16638	jlrxa005e05t1 jlrxa005e05t1
vimentin	Late	vimentin cytoskeletal protein	X62952	jtrba108b01t1
phosphacan	Late	chondroitin sulfate proteoglycan of brain that interacts with neurons and neural cell-adhesion molecules and is an extracellular variant of receptor-type protein tyrosine phosphatase	U04998	jtrba011f03t1
NARC19	Late	epididymal secretory protein		jtrba078d11t1
L4BD NADP-dependent oxidoreductase	Late	catalyses the key step of metabolic inactivation of leukotriene B4 in tissues other than leukocytes	U66322	jlrxa015a05t1
S100 beta	Late	Ca ²⁺ binding protein		
GABA _A receptor α -1 subunit	Late	inhibitory neurotransmitter GABA receptor	M63436	jtrba011b04t1
OXY B	Late	oxysterol-binding protein that may play a role in regulation of sterol metabolism	P22059	jlrxa048f03t1
cystatin C	Late	cysteine proteinase inhibitor that regulates cysteine protease activity including cathepsin B, H, and L	P14841	jtrba005h01t1
P0	Late	60S acidic ribosomal protein	P19945	jtrba076a01t2
Na ⁺ /Cl ⁻ -dependent GABA transporter protein 1	Late	terminates the inhibitory action of GABA by its high affinity Na ⁺ -dependent reuptake into presynaptic terminals	P31648	jtrba078h11t1
Apo E	Late	apolipoprotein E transcription factor	J02582	jtrba119b10t1

FIG. 9A.

SUBSTITUTE SHEET (RULE 26)

27/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
T-cell receptor γ -like protein	Late	novel protein with some homology to retroviral GAG and murine T-cell receptor γ .	M18247	jlrxa004c06t1
gelsolin-like protein	Late	novel protein with some homology to gelsolin involved in Ca^{2+} -regulated, actin-modulation; promotes assembly of monomers into filaments, as well as sever filaments already formed; binds fibronectin	P20305	jlrxa019e05t2
NAD ⁺ -dep. MTHFDH cyclohydrolase	Late	mitochondrial NAD-dependent bifunctional methylenetetrahydrofolate dehydrogenase/cyclohydrolase	P18155	jlrxa042c04t1
leukotriene A-4 hydrolase	Late	hydrolyzes an epoxide moiety of LTA-4 to form LTB-4; also has some peptidase activity; 3rd step in the biosynthesis of leukotrienes; belongs to zinc metallopeptidase family	M63848	jlrxa049a09t1
SKR6	Late	CB1 cannabinoid G protein coupled receptor	X55812	jtrba035f05t1
PRCD	Late	proteasome delta chain precursor for a multicatalytic proteinase complex involved in ATP/ubiquitin-dependent non-lysosomal proteolytic pathway	P28073	jlrxa030g11t1
N- α -acetyl transferase-like protein	Late	novel with some homology to N- α -acetyl transferase (acetylation of proteins and peptides, used to stabilise pharmaceuticals or induce herbicide resistance in plants)	P12945	jlrxa030g11t1
Na^+ - K^+ -2 Cl^- cotransporter	Late	electrically silent basolateral cotransporter which mediates Na^+ / Cl^- reabsorption required for the regulation of ionic balance and cell volume	P55012	jtrba066f05t2
SNK-like protein	Late	novel protein with some homology to SNK.	P53351	jlrxa060b04t1
transferrin	Late	serum/phorbol ester induced serine/threonine kinase transports iron from sites of absorption and heme degradation to those of storage and utilization; may have a role in stimulating cell proliferation	D38380	jtrba103b07t2
testican	Late	testican	X92864	jtrba18g01t1
CaBP2	Late	Ca^{2+} -binding protein with protein disulfide isomerase activity involved in rearrangement of intrachain & interchain disulfide bonds in proteins to form native structures in the ER	P38659	jlrxa054d03t1
PIP 4	Late	deoxyuridine 5'-triphosphate nucleotidohydrolase PPAR-interacting protein 4 (decreases intracellular dUTP such that uracil cannot be incorporated into DNA; binds and prevents PPAR dimerization with retinoid X receptor by inducing PPAR translocation to the nucleus	P70583	jlrxa001b10t2
DM-GRASP	Late	Ig superfamily axonal surface protein that supports neurite extension	L25274	jtrba009d09t2
testican-agrin-like protein	Late	novel protein with homology to testican and agrin	X92864	jtrba120c06t1
Transketolase	Late	implicated in Wernicke-Korsakoff syndrome caused brain damage	U09256	jlrxa044b08t1
EF1G	Late	elongation factor 1-gamma that may anchor mRNA elongation complex to other cellular components	P26641	jlrxa020c04t2
APG-1 synaptophysin	Late Middle	Hsp70-related protein induced by hyperosmolar salt stress synaptophysin synaptic vesicle protein	AB001926 P07825	jtrba018b01t1 jlrxa004f07t1

FIG. 9B.

SUBSTITUTE SHEET (RULE 26)

28/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
NADH dehydrogenase subunit 4	Middle	mitochondrial gene that shares regions of homology with NADH dehydrogenase subunit 4	U40063	jlrxa020h01t2
cytochrome c oxidase	Middle	mitochondrial cytochrome c oxidase	X14848	jtrba053b12t1
MU-crystallin homologue	Middle	NADP-regulated thyroid-hormone binding protein involved in the regulation of the free intracellular concentration of triiodothyronine and access to its nuclear receptors	Q14894	jtrba087g05t1
ubiquinone oxidoreductase-like protein	Middle	novel protein with domain homology to scavenger-receptor protein NADH-ubiquinone oxidoreductase chain 4	P05508	jtrba126d05t1
zinc finger protein	Middle	zinc finger protein	L03386	jtrba065c11t1
NARC4	Middle	novel	W57419	jlrxa012g04t1
L-proline transporter	Middle	high affinity L-proline transporter	M88111	jtrba030a12t1
NARC5	Middle	novel	X53581	jtrba034g08t3
NARC7	Middle	novel		jlrxa027h05t1
NARC6	Middle	novel		jtrba118a12t1
caspase 3	Middle	PCD	P55213	jlrxc015b03t1
Zfh-2	Middle	zinc finger homeodomain enhancer-binding protein-2 that binds T3-response elements	U51583	jlrxa032a03t1
heparan sulfate proteoglycan core-like protein	Middle	novel protein with domain homology to basement membrane heparan sulfate proteoglycan core cell adhesion	M85289	jtrba107h07t1
RasGAP	Middle	RasGAP		rlrX019a05a1
4F2 bc	Middle	splice variant of type II membrane glycoprotein, a cell growth antigen required for expression of system L-like neutral amino acid-transport activity in C6-Bu-1 rat glioma	X89225	jtrba085b10t2
BCR 1	Middle	breakpoint cluster region protein (GTPase-activating protein for RAC1 and CDC42; chromosomal translocation produces BCR-ABL oncogene)	U07000	jtrba085b10t2
hippocalcin	Middle	neuron specific calcium-binding protein of the recoverin family expressed in hippocampus	D12573	jlrxa054b09t1
RT1.Aw3	Middle	MHC class 1 protein	L40363	jtrba019c12t1
Y-box binding protein	Middle	mammalian equivalents of the bacterial stress-inducible oxyR gene	L35599	jlrxa013c05t1
p130	Middle	cyclin-dependent kinase homologue that is a phosphotyrosine-independent SH2 ligand		jtrba108f08t1
PITSLRE	Middle	novel	U59235	jtrba119f08t1
NARC20	Middle	apoptosis regulated protein phosphatase		rlrX019b05a1
Wip-1	Middle	pyruvate dehydrogenase kinase 2 subunit	U10357	jtrba088c07t1
p45	Middle	serine/threonine-protein kinase in the CDC2/CDKX subfamily	Q00537	jtrba038e02t1
PCTAIRE-2	Middle	novel	AA182996	jtrba011c02t1
NARC14	Middle	novel protein with similarity to acetyltransferase		jtrba073f01t1
acetyltransferase-like protein	Middle			

FIG. 9C.

SUBSTITUTE SHEET (RULE 26)

29/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
hTOM34-like protein	Middle	novel protein with domain homology to outer mitochondrial membrane 34 kDa translocase hTOM34	U58970	jtrba032b11t3
scavenger-receptor-like protein	Middle	novel protein with domain homology to scavenger-receptor protein	X99336	jtrba058c09t1
CAT-1	Middle	cationic amino acid transporter-1	U70476	jtrba043f05t1
carbonic anhydrase-like protein	Middle	novel protein with some homology to carbonic anhydrase	Y07785	jtrba002g03t1
NET-1-like protein	Middle	novel protein with some homology to guanine nucleotide regulatory protein NET-1 neuroepithelium oncogene	U02081	jlrxa043f01t1
stathmin	Middle	ubiquitous phosphoprotein which acts as an intracellular relay of proliferation and differentiation	Q09004	jtrba028g11t1
DIF-1-like protein	Middle	novel protein similar to C. elegans mitochondrial carrier protein DIF-1	Z75532	jtrba044d04t1
pentraxin	Early	neuronal pentraxin	U18772	jtrba049h01t2
NARC1	Early	novel		rlrX020c06b1
NARC2	Early	novel		rlrX022f07b1
apoptosis associated tyrosine kinase	Early	apoptosis associated tyrosine kinase	AF011908	jlrxa055h07t1
NARC30	Early	cell-death regulated secreted protein		jtrba057e05t1
Na ⁺ channel β -1 subunit	Early	Na ⁺ channel β -1 subunit, neurotransmission	Q00954	jtrba041g04t1
rhoB	Early	ras-like immediate early gene	M74295	jlrxa021g01t1
NARC13	Early	novel	D80004	jlrxa031h11t1
histone H2A	Early	histone H2A	X05862	jtrba124h04t1
PC3	Early	NGF-inducible anti-proliferative protein involved in cell cycle regulation, in particular, growth arrest and differentiation of neuronal precursor cells.	P27049	jlrxa027g01t1
histone H1	Early	histone H1	X72624	jtrba010c12t1
NARC9	Early	similar to C.elegans glycerophosphoryl diester phosphodiesterase	Q10003	jtra022a05t1
histone H3	Early	histone H3	M17876	jtrba019a12t2
HAP1-A-like protein	Late	similar to huntingtin-associated protein HAP1-A (except missing expanded polyglutamine repeat)	U38373	jtrba022f11t1
TAI	Late	a highly conserved oncofetal mRNA from rat hepatoma that encodes an integral membrane protein associated with liver development, carcinogenesis, and cell activation		jtrba026f06t3
DP5-like protein	Middle	similar to mRNA for polypeptide DP5 induced during programmed neuronal death	D83697	jtrba007a11t1
Nel	Middle	protein kinase C-binding protein Nel	U48245	jtrba016c05t1
CaMKII γ B	Imm. Early	CaMKII γ B involved in neuronal plasticity	U48245	jtrba016c05t1
dual specificity protein phosphatase 3	Imm. Early	dual specificity (tyrosine-, serine-) non-receptor protein phosphatase 3 involved in neuronal plasticity	P51452	jtrba073b11t1

FIG. 9D.

SUBSTITUTE SHEET (RULE 26)

30/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
LDL receptor	Imm. Early	low-density lipoprotein receptor that binds LDL, the major cholesterol-carrying lipoprotein of plasma and transports it into cells by endocytosis after clustering in clathrin-coated pits	P35952	jlrxa012e10t1
NGF-In	Imm. Early	NGF responsive transcription factor		
ZO-2	Imm. Early	zonula occludens 2 protein, a tight junction protein homologous to the Drosophila discs-large tumor suppressor protein (contains guanylate kinase homology and three PDZ domains)		rlrX017b03b1
GluR5	Imm. Early	excitatory neurotransmitter glutamate receptor		
vesicle-associated calmodulin-binding kinase-like protein	Imm. Early	vesicle-associated calmodulin-binding kinase-like protein	L22557	jtrba027g06t4
Neuro-endocrine secretory protein 55	Imm. Early	neuroendocrine secretory protein 55 in the chromagranin family	U77614	jlrxa055g08t1
NARC24	Imm. Early	novel		jlrxa009d08t1
β -synuclein	Imm. Early	β -synuclein	S69965	jtrba040h07t1
NARC21	Imm. Early	novel	AA442186	jlrxa025e06t1
14-3-3 protein tau	Imm. Early	multifunctional regulator of CaMKII and PKC involved in neuronal plasticity	P35216	jtrba021f01t1
synapsin Ib	Imm. Early	synapsin Ib	M27924	jtrba041h10t1
NSG-1 P19 protein	Imm. Early	expressed in the Golgi apparatus of neural/neuroendocrine cells	P47759	jlrxa011c11t1
NARC12	Imm. Early	novel	H35593	jlrxa031b03t1
synaphin 2	Imm. Early	Protein associated with docking/fusion complex for synaptic vesicle release		jtrba035a02t1
Ca ²⁺ -dependent action-binding protein	Imm. Early	activator protein for secretion/exocytosis	U16802	jtrba046c05t1
ribonucleoprotein L	Imm. Early	heterogeneous nuclear ribonucleoprotein L	X16135	jtrba025h11t1
TCP-1	Imm. Early	chaperonin	P80317	jlrxa019a09t2
MCM2	Imm. Early	DNA replication licensing factor required for the entry into S phase and for cell division	P97310	jtrxa037f09t1
cytochrome b-560	Imm. Early	succinate-ubiquinone reductase	U31241	jtrba001a08t1
α -tubulin-1	Down-reg.	α -tubulin-1 cytoskeletal protein		jtrba012g02t1
vacuolar H ⁺ -ATPase	Down-reg.	active proton transport	M88690	jlrxa011g01t1
Ribonuclear particle U-like protein	Down-reg.	novel protein with some domain homology to heterogeneous ribonuclear particle protein U	D14048	jlrxa034f09t1
spindlin	Down-reg.	major maternal transcript expressed in the mouse during the transition from oocyte to embryo	U48972	jlrxa037b02t1
fodrin	Imm. Early	cytoskeletal component of Ca ²⁺ -secretion		jlrxa011g07t1

FIG. 9E.

SUBSTITUTE SHEET (RULE 26)

31/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
MAP-IB	Imm. Early	microtubule associated cytoskeletal protein	A56577	jlrxa035f07t1
Synaptotagmin 1	Imm. Early	synaptic vesicle protein that binds to neurexins, syntaxin, and AP2 in Ca ²⁺ -independent fashion	P48018	jlrxa046c07t1
thyroid hormone-responsive gene	Imm. Early	gene from the Down syndrome critical region resembling a DNA binding protein with an SH3 domain	D83407	jtrba016g02t1
neurocalcin	Imm. Early	calcium-binding protein	D10884	jtrba035d02t1
NAD ⁺ -isocitrate dehydrogenase	Imm. Early	mitochondrial (NAD ⁺) isocitrate dehydrogenase	P50213	jlrxa060g03t1
NF-H	Imm. Early	heavy neurofilament cytoskeletal protein	M37227	jtrba073g11t1
K ⁺ channel β -subunit	Imm. Early	K ⁺ channel β -subunit, neurotransmission	L47665	jtrba022g06t1
NADH-ubiquinone oxidoreductase	Imm. Early	transfers electrons from NADH to ubiquinone in the respiratory chain	P34942	jlrxa026e02t1

(Table 2 footnotes)

^aThe gene identified as the most similar to the array element EST by BLAST. Genes are listed in the same order of hierarchical clustering as in Figure 4B.

^bGene expression class identified based on hierarchical clustering (Figure 4B). Following KCl/serum-withdrawal, Imm. Early, Early, Middle, and Late genes peaked at 1 hr., 3 or 6 hr., 12 hr., and 24 hr., respectively.

^cDescription of the gene identified by BLAST. The description was obtained from the Medline annotation for each sequence with indicated accession number. Red font highlights possible function physiologically relevant to programmed cell death or survival.

^dGenBank accession number.

^eSmart Chip EST identification number.

FIG. 9F.

SEQUENCE LISTING

<110> Chiang, Lillian Wei-Ming

<120> Nucleic Acid Molecules Derived from
Rat Brain and Programmed Cell Death Models

<130> 35800/205244 (5800-37-1)

<160> 11

<170> FastSEQ for Windows Version 3.0

<210> 1

<211> 2738

<212> DNA

<213> Homo sapiens

<400> 1

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cgatggaacc	agataaacctg	gaactaatct	ttgatttttt	cgaagaagat	ctcagtgcgc	180
acgtagtcca	gggtgatgcc	cttcctggac	atgtgggtac	agcttgtctc	ttatcatcca	240
ccattgtctga	gagtggaaaag	agtgtctggaa	ttcttactct	tcccatcatg	agcagaaatt	300
cccggaaaaac	aataggcaaaa	gtgagagttg	actatataat	tattaagcca	ttaccaggat	360
acagttgtga	catgaaatct	tcattttcca	agtattggaa	gccaagaata	ccattggatg	420
ttggccatcg	aggtgcagga	aactctacaa	caactgccca	gctggctaaa	gttcaagaaa	480
atactattgc	ttctttaaga	aatgctgcta	gtcatgggtc	agcctttgta	gaatttgacg	540
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Ser Ile His Ala Ser Cys Cys His Ala Pro Gly Leu Glu Cys Lys Val
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His Val Leu Gly Ala Tyr Ala Val Asp Asn Thr Cys Val Val Arg Ser
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cgg gac gtc agc act aca ggc agc acc agc gaa gag gcc gtg aca gcc      2254
Arg Asp Val Ser Thr Thr Gly Ser Thr Ser Glu Glu Ala Val Thr Ala
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Val Ala Ile Cys Cys Arg Ser Arg His Leu Ala Gln Ala Ser Gln Glu
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Leu Gln

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His Arg Cys Ala Lys Asp Pro Trp Arg Leu Pro Gly Thr Tyr Val Val
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Val Leu Lys Glu Glu Thr His Leu Ser Gln Ser Glu Arg Thr Ala Arg
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 Glu Asp Ser Ser Val Phe Ala Gln Ser Ile Pro Trp Asn Leu Glu Arg
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 Ile Thr Pro Pro Arg Tyr Arg Ala Asp Glu Tyr Gln Pro Pro Asp Gly
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 Gly Ser Leu Val Glu Val Tyr Leu Leu Asp Thr Ser Ile Gln Ser Asp
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 His Arg Glu Ile Glu Gly Arg Val Met Val Thr Asp Phe Glu Asn Val
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 Pro Glu Glu Asp Gly Thr Arg Phe His Arg Gln Ala Ser Lys Cys Asp
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 Ser His Gly Thr His Leu Ala Gly Val Val Ser Gly Arg Asp Ala Gly
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 Lys Ser Gln Leu Val Gln Pro Val Gly Pro Leu Val Val Leu Leu Pro
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 Ile Gly Ala Ser Ser Asp Cys Ser Thr Cys Phe Val Ser Gln Ser Gly
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 Thr Ser Gln Ala Ala Ala His Val Ala Gly Ile Ala Ala Met Met Leu
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 Ser Ala Glu Pro Glu Leu Thr Leu Ala Glu Leu Arg Gln Arg Leu Ile
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 His Phe Ser Ala Lys Asp Val Ile Asn Glu Ala Trp Phe Pro Glu Asp
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 Gln Arg Val Leu Thr Pro Asn Leu Val Ala Ala Leu Pro Pro Ser Thr
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 His Gly Ala Gly Trp Gln Leu Phe Cys Arg Thr Val Trp Ser Ala His
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 Ser Gly Pro Thr Arg Met Ala Thr Ala Ile Ala Arg Cys Ala Pro Asp
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 Glu Glu Leu Leu Ser Cys Ser Ser Phe Ser Arg Ser Gly Lys Arg Arg
 485 490 495
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 Asn Ala Phe Gly Gly Glu Gly Val Tyr Ala Ile Ala Arg Cys Cys Leu
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 Ser Met Gly Thr Arg Val His Cys His Gln Gln Gly His Val Leu Thr
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 Pro Val Leu Arg Pro Arg Gly Gln Pro Asn Gln Cys Val Gly His Arg
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 Glu Ala Ser Ile His Ala Ser Cys Cys His Ala Pro Gly Leu Glu Cys
 595 600 605
 Lys Val Lys Glu His Gly Ile Pro Ala Pro Gln Glu Gln Val Thr Val
 610 615 620
 Ala Cys Glu Glu Gly Trp Thr Leu Thr Gly Cys Ser Ala Leu Pro Gly

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Arg Ser Arg Asp	Val Ser Thr Thr Gly Ser Thr Ser Glu Glu Ala Val					
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Leu Pro Gly Phe Leu Val Lys Met Ser Gly Asp Leu Leu Glu Leu Ala	
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Leu Lys Leu Pro His Val Asp Tyr Ile Glu Glu Asp Ser Ser Val Phe	
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Ala Gln Ser Ile Pro Trp Asn Leu Glu Arg Ile Thr Pro Pro Arg Tyr	
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Arg Ala Asp Glu Tyr Gln Pro Pro Asp Gly Gly Ser Leu Val Glu Val	

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 His Arg Cys Ala Lys Asp Pro Trp Arg Leu Pro Gly Thr Tyr Val Val
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 Val Leu Lys Glu Glu Thr His Leu Ser Gln Ser Glu Arg Thr Ala Arg
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 Arg Leu Gln Ala Gln Ala Ala Arg Arg Gly Tyr Leu Thr Lys Ile Leu
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 Pro Glu Glu Asp Gly Thr Arg Phe His Arg Gln Ala Ser Lys Cys Asp
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International Bureau



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MA 02138 (US).

(21) International Application Number: PCT/US00/29132

(74) Agents: **COULTER, Kathryn, L.** et al.: Alston & Bird
LLP, Bank of America Plaza, 101 South Tryon Street, Suite
4000, Charlotte, NC 28280-4000 (US).

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(utility model), AU, AZ, BA, BB, BG, BR, BY, BZ, CA,
CH, CN, CR, CU, CZ, CZ (utility model), DE, DE (utility
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model), ES, FI, FI (utility model), GB, GD, GE, GH, GM,
HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK,
LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX,
MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SK
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(71) Applicant (*for all designated States except US*): **MIL-
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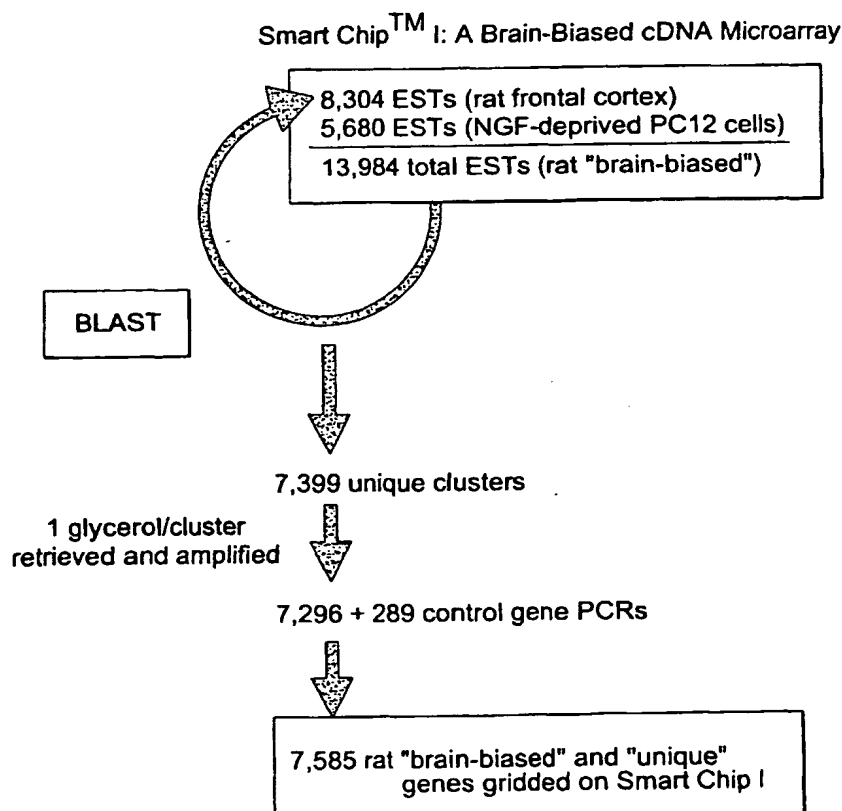
(84) Designated States (*regional*): ARIPO patent (GH, GM,
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European

(72) Inventor; and

(75) Inventor/Applicant (*for US only*): **CHIANG, Lillian,**

[Continued on next page]

(54) Title: NUCLEIC ACID MOLECULES DERIVED FROM RAT BRAIN AND PROGRAMMED CELL DEATH MODELS



(57) Abstract: The invention is directed to human homologs of nucleic acid molecules derived from rat brain and programmed cell death expression libraries. These molecules can constitute microarrays of expressed sequences useful for analyzing gene expression in various biological contexts, including development, differentiation, and disease, both *in vivo* and *in vitro*. The nucleic acid molecules are useful for diagnosis, treatment, and drug discovery. The nucleic acid molecules are useful for creating microarrays for transcriptional profiling. The invention further provides peptides encoded by the nucleic acid molecules, useful for methods of diagnosis, treatment, and drug discovery. The invention specifically relates to nucleic acid molecules involved in programmed cell death.

WO 01/31007 A3



patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,
IT, LU, MC, NL, PT, SE). OAPI patent (BF, BJ, CF, CG,
CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(88) Date of publication of the international search report:
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

INTERNATIONAL SEARCH REPORT

International Application No

PC., US 00/29132

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 98 49297 A (AMERICAN HOME PROD) 5 November 1998 (1998-11-05) abstract; claims 1-7 ---	1-17,26, 28
A	AOKI TOMOKAZU ET AL: "Rat TAFII-31 gene is induced upon programmed cell death in differentiated PC12 cells deprived of NGF." BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 234, no. 1, 1997, pages 230-234, XP002163760 ISSN: 0006-291X abstract; figures 4,5 ---	1-17,26, 28
A	RICKMAN DENNIS W ET AL: "Characterization of the cell death promoter, Bad, in the developing rat retina and forebrain." DEVELOPMENTAL BRAIN RESEARCH, vol. 115, no. 1, 8 June 1999 (1999-06-08), pages 41-47, XP000990537 ISSN: 0165-3806 abstract ---	1-17,26, 28
A	HEMACHANDRA REDDY P ET AL: "Transgenic mice expressing mutated full-length HD cDNA: A paradigm for locomotor changes and selective neuronal loss in Huntington's disease." PHILOSOPHICAL TRANSACTIONS OF THE ROYAL SOCIETY OF LONDON B BIOLOGICAL, vol. 354, no. 1386, June 1999 (1999-06), pages 1035-1045, XP000990516 June, 1999 ISSN: 0962-8436 abstract -----	1-17,26, 28

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 00/29132

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 22-25 and claims 18-21, as far as they concern an in vivo method, are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☒ Claims Nos.: 27
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims: it is covered by claims Nos.:

see subject 1. on extra sheet

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box 1.2

Claims Nos.: 27

Claim 27 relating to binding agents to the polypeptide of claim 10 could not be searched as its subject-matter was insufficiently disclosed .

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.1 or 4 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

2. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.2 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

3. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.3 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

4. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.5 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

5. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.6 or 8 and corresponding polypeptides , vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe.

6. Claims: 1-28 partly

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

nucleic acids with nucleotide sequence with seq.id.10 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

formation on patent family members

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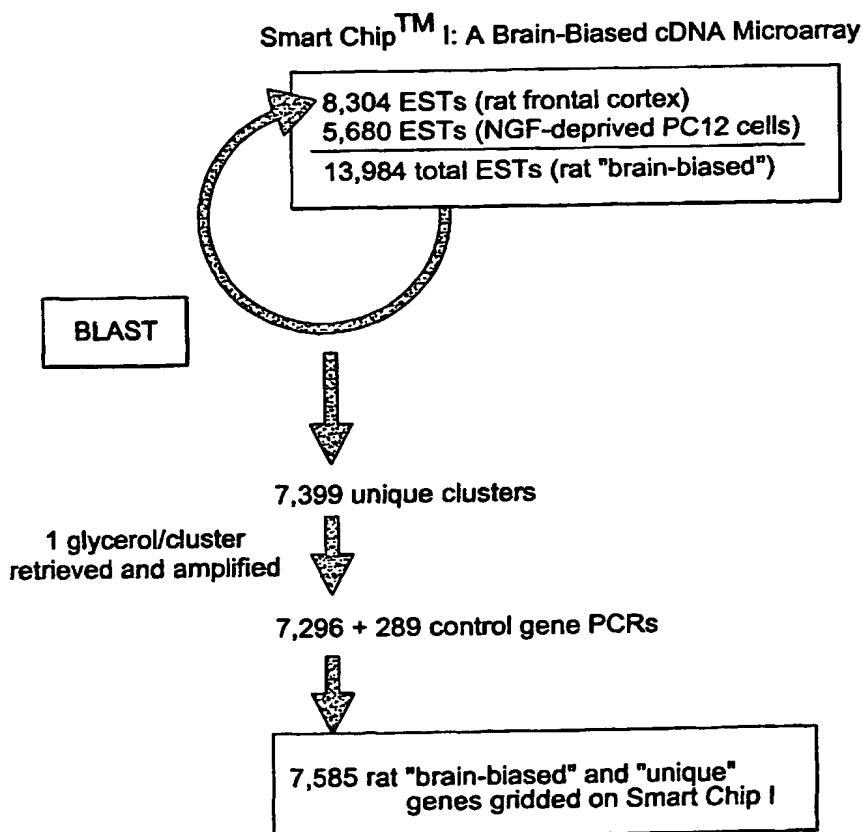
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- (51) International Patent Classification⁷: C12N 15/12, C07K 14/47, 16/18, C12Q 1/68, G01N 33/53, A61K 38/17
- (72) Inventor; and
(75) Inventor/Applicant (for US only): CHIANG, Lillian, Wei-Ming [US/US]; 38 Shepard Street #1, Cambridge, MA 02138 (US).
- (21) International Application Number: PCT/US00/29132
- (22) International Filing Date: 20 October 2000 (20.10.2000)
- (74) Agents: COULTER, Kathryn, L. et al.; Alston & Bird LLP, Bank of America Plaza, 101 South Tryon Street, Suite 4000, Charlotte, NC 28280-4000 (US).
- (25) Filing Language: English
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- (30) Priority Data:
60/161,188 22 October 1999 (22.10.1999) US
- (71) Applicant (for all designated States except US): MILLENNIUM PHARMACEUTICALS, INC. [US/US]; 75 Sidney Street, Cambridge, MA 02139 (US).

[Continued on next page]

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(utility model), SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

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NUCLEIC ACID MOLECULES DERIVED FROM RAT BRAIN AND PROGRAMMED CELL DEATH MODELS

FIELD OF THE INVENTION

The invention relates to nucleic acid molecules derived from rat brain and programmed cell death expression libraries. Also provided are vectors, host cells, and
5 methods for making and using the novel molecules of the invention.

BACKGROUND OF THE INVENTION

A great deal of effort has been expended by the modern scientific research community to identify and sequence genes, particularly human genes. The
10 identification of genes and knowledge of their nucleic acid sequences pave the way for many scientific and commercial advancements, both in research applications and in diagnostic and therapeutic applications. For example, advances in gene identification and sequencing allow the production of the products encoded by these genes, such as by recombinant and synthetic means. Furthermore, identification of
15 genes and the products they encode provide important information about the mechanism of disease and can provide new diagnostic tests and therapeutic treatments for the diagnosis and treatment of disease. Thus, identification and sequencing of genes provide valuable information and compositions for use in the biotechnology and pharmaceutical industries.

20 In multicellular organisms, homeostasis is maintained by balancing the rate of cell proliferation against the rate of cell death. Cell proliferation is influenced by numerous growth factors and the expression of proto-oncogenes, which typically encourage progression through the cell cycle. In contrast, numerous events, including the expression of tumor suppressor genes, can lead to an arrest of cellular
25 proliferation.

In differentiated cells, a particular type of cell death called apoptosis occurs when an internal suicide program is activated. This program can be initiated by a variety of external signals as well as signals that are generated within the cell in response to, for example, genetic damage. Dying cells are eliminated by phagocytes,
30 without an inflammatory response.

Programmed cell death (PCD) is a highly regulated process (Wilson (1998) *Biochem. Cell. Biol.* 76:573-582). The death signal is then transduced through various signaling pathways that converge on caspase-mediated degradative cascades resulting in the activation of late effectors of morphological and physiological aspects of apoptosis, including DNA fragmentation and cytoplasmic condensation. In addition, regulation of programmed cell death may be integrated with regulation of energy, redox- and ion homeostasis in the mitochondria (reviewed by Kroemer (1998) *Cell Death and Differentiation* 5:547), and/or cell-cycle control in the nucleus and cytoplasm (reviewed by Choisy-Rossi and Yonish-Rouach (1998) *Cell Death and Differentiation* 5:129-131; Dang (1999) *Molecular and Cellular Biology* 19:1-11; and Kasten and Giordano (1998) *Cell Death and Differentiation* 5:132-140). Many mammalian genes regulating apoptosis have been identified as homologs of genes originally identified genetically in *Caenorhabditis elegans* or *Drosophila melanogaster*, or as human oncogenes. Other programmed cell death genes have been found by domain homology to known motifs, such as death domains, that mediate protein-protein interactions within the programmed cell death pathway.

The mechanisms that mediate apoptosis include, but are not limited to, the activation of endogenous proteases, loss of mitochondrial function, and structural changes such as disruption of the cytoskeleton, cell shrinkage, membrane blebbing, and nuclear condensation due to degradation of DNA. The various signals that trigger apoptosis may bring about these events by converging on a common cell death pathway that is regulated by the expression of genes that are highly conserved.

Caspases (cysteine proteases having specificity for aspartate at the substrate cleavage site) are central to the apoptotic program, are. These proteases are responsible for degradation of cellular proteins that lead to the morphological changes seen in cells undergoing apoptosis. One of the human caspases was previously known as the interleukin-1 β (IL-1 β) converting enzyme (ICE), a cysteine protease responsible for the processing of pro-IL-1 β to the active cytokine. Overexpression of ICE in Rat-1 fibroblasts induces apoptosis (Miura *et al.* (1993) *Cell* 75:653).

Many caspases and proteins that interact with caspases possess domains of about 60 amino acids called a caspase recruitment domain (CARD). Apoptotic proteins may bind to each other via their CARDS. Different subtypes of CARDS may

confer binding specificity, regulating the activity of various caspases. (Hofmann *et al.* (1997) *TIBS* 22:155).

The functional significance of CARDs have been demonstrated in two recent publications. Duan *et al.* (1997) *Nature* 385:86 showed that deleting the CARD at the N-terminus of RAIDD, a newly identified protein involved in apoptosis, abolished the ability of RAIDD to bind to caspases. In addition, Li *et al.* (1997) *Cell* 91:479 showed that the N-terminal 97 amino acids of apoptotic protease activating factor-1 (Apaf-1) was sufficient to confer caspase-9-binding ability.

Thus, programmed cell death (apoptosis) is a normal physiological activity necessary to proper and differentiation in all vertebrates. Defects in apoptosis programs result in disorders including, but not limited to, neurodegenerative disorders, cancer, immunodeficiency, heart disease and autoimmune diseases (Thompson *et al.* (1995) *Science* 267:1456).

In vertebrate species, neuronal programmed cell death mechanisms have been associated with a variety of developmental roles, including the removal of neuronal precursors which fail to establish appropriate synaptic connections (Oppenheim *et al.* (1991) *Annual Rev. Neuroscience* 14:453-501), the quantitative matching of pre- and post-synaptic population sizes (Herrup *et al.* (1987) *J. Neurosci.* 7:829-836), and sculpting of neuronal circuits, both during development and in the adult (Bottjer *et al.* (1992) *J. Neurobiol.* 23:1172-1191).

Inappropriate apoptosis has been suggested to be involved in neuronal loss in various neurodegenerative diseases such as Alzheimer's disease (Loo *et al.* (1993) *Proc. Natl. Acad. Sci.* 90:7951-7955), Huntington's disease (Portera-Cailliau *et al.* (1995) *J. Neurosci.* 15:3775-3787), amyotrophic lateral sclerosis (Rabizadeh *et al.* (1995) *Proc. Natl. Acad. Sci.* 92:3024-3028), and spinal muscular atrophy (Roy *et al.* (1995) *Cell* 80:167-178).

In addition, improper expression of genes involved in apoptosis has been implicated in carcinogenesis. Thus, it has been shown that several "oncogenes" are in fact involved in apoptosis, such as in the Bcl family.

Accordingly, genes involved in apoptosis are important targets for therapeutic intervention. It is important, therefore, to identify novel genes involved in apoptosis or to discover whether known genes function in this process.

Nucleic acid probes have long been used to detect complementary nucleic acid sequences in a nucleic acid of interest (the "target" nucleic acid). In some assay formats, the nucleic acid is tethered, i.e., by covalent attachment, to a solid support. Arrays of nucleic acid sequences immobilized on solid supports have been used to
5 detect specific nucleic acid sequences in a target nucleic acid. See, e.g., PCT patent publication Nos. WO 89/10977 and 89/11548. Others have proposed the use of large numbers of nucleic acid sequences to provide the complete nucleic acid sequence of a target nucleic with methods for using arrays of immobilized nucleic acid sequences for this purpose. See U.S. Pat. Nos. 5,202,231 and 5,002,867 and PCT patent
10 publication No. WO 93/17126.

The development of specific microarray technology has provided methods for making very large arrays of nucleic acid sequences in very small physical arrays. See U.S. Pat. No. 5,143,854 and PCT patent publication Nos. WO 90/15070 and
92/10092, each of which is incorporated herein by reference. U.S. patent application
15 No. 082,937, filed Jun. 25, 1993, describes methods for making arrays of sequences that can be used to provide the complete sequence of a target nucleic acid and to detect the presence of a nucleic acid containing a specific nucleotide sequence. Thus, microfabricated arrays of large numbers of nucleic acid sequences, called "DNA chips" offer great promise for a wide variety of applications.

20

SUMMARY OF THE INVENTION

The present invention is based on the identification of novel nucleic acid molecules derived from rat brain and programmed cell death cDNA libraries.

Thus, in one aspect, the invention provides an isolated nucleic acid molecule
25 that comprises a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

The invention also provides an isolated fragment or portion of any of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complement of the
30 sequences shown in SEQ ID NOS: 1-6, 8, and 10. In some embodiments, the fragment is useful as a probe or primer, and/or is at least 15, at least 18, or at least 20, 22, 25, 30, 35, 50, 100, 200 or more nucleotides in length.

In another embodiment, the invention provides an isolated nucleic acid molecule that comprises a nucleotide sequence that is at least about 60% identical, about 65% identical, about 70% identical, about 80% identical, about 90% identical, about 95% identical, about 96% identical, about 97% identical, about 98% identical, 5 or about 99% or more identical to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

In another embodiment, the invention provides an isolated nucleic acid molecule that hybridizes under highly stringent conditions to a nucleotide sequence 10 selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10.

The invention further provides nucleic acid vectors comprising the nucleic acid molecules described above. In one embodiment, the nucleic acid molecules of the invention are operatively linked to at least one expression control element.

15 The invention further includes host cells, such as bacterial cells, fungal cells, plant cells, insect cells and mammalian cells, comprising the nucleic acid vectors described above.

In another aspect, the invention provides isolated gene products, proteins and polypeptides encoded by nucleic acid molecules of the invention.

20 The invention further provides antibodies, including monoclonal antibodies, or antigen-binding fragments thereof, which selectively bind to the isolated proteins and polypeptides of the invention.

The invention also provides methods for preparing proteins and polypeptides encoded by isolated nucleic acid molecules described herein by culturing a host cell 25 containing a vector molecule of the invention.

Additionally, the invention provides a method for assaying for the presence of a nucleic acid sequence, protein or polypeptide of the present invention, in a biological sample, e.g., in a tissue sample, by contacting said sample with an agent (e.g., an antibody or a nucleic acid molecule) suitable for specific detection of the 30 nucleic acid sequence, protein or polypeptide.

The invention also provides a kit comprising a nucleic acid probe which hybridizes to a nucleotide sequence of claim 1 and instructions for use, and a kit comprising an agent which binds to a polypeptide of claim 10 and instructions for use.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 illustrates the construction of the "Smart Chip™ I". cDNAs were cloned from rat frontal cortex and from differentiated PC12 cells deprived of nerve growth factor, a model of programmed cell death as described in detail in the experimental section. PC12 cells are an adrenal gland cell line from rat that provides a pre-neuron set that can be differentiated *in vitro*. The application of nerve growth factor induces the formation of axons and dendritic structures. This serves as a model for neuronal differentiation. When the nerve growth factor is withdrawn, the cells undergo programmed cell death (apoptosis). Approximately 300 control nucleic acid sequences (of known function) were added as an internal control and for transcriptional profiling of the cloned cDNA sequences. These sequences were then subjected to BLASTX analysis to determine the correspondence between the cDNA and a known cDNA and to determine to which protein family, if any, the proteins encoded by each cDNA belong. Computer analysis was used to assemble the cDNA sequences into unique clusters. The majority of the clusters as well as control genes were gridded on Smart Chip™ I.

Figure 2 shows the coefficient of variation (standard deviation/mean for triplicate hybridizations) after normalization for each array element plotted against the mean intensity for the gene (gene expression intensity). The figure shows the moving average (with a window of 200) for three different mRNA probes, 3 hour KCl-withdrawn, 3 hour control, and 6 hour control (See the examples and figures 3 and 4). As is typical for all probes, past a threshold of 30 to 40, the coefficient of variation averages below 0.2. The inset compares one triplicate hybridization (Filter Y) to another (Filter Z). Each point represents a different gene graphed on log-log axis comparing the intensity measured on one filter versus the other.

Figure 3 shows temporal expression clusters observed following KCl and serum withdrawal. A hierarchical clustering algorithm was used to cluster genes based on expression patterns across 10 time points (from left to right), 1, 3, 6, 12, and 24 hours post-KCl/serum-replacement (sham), and 1, 3, 6, 12, and 24 hours post-KCl/serum-withdrawal (treatment) (See Examples). Expression values for each gene were scaled based on the number of standard deviations from the mean intensity of each gene across all 10 time points. Scaled expression values are color-coded such

that red, yellow, and blue indicated above, at, and below mean intensity, respectively. The correlation between expression patterns of neighboring genes is depicted by the dendrogram on the right. Genes regulated by programmed cell death (KCl/serum-withdrawal alone) are enlarged in B. Representative non-scaled gene expression bar graphs with standard deviation error bars are aligned next to the four major clusters for Late Effector, Middle, Early, and Immediate Early gene expression classes. Regulated genes within each temporal expression class are listed in order of hierarchical clustering in SEQ ID NOS: 1-6, 8, and 106.

Figure 4 shows expression clusters for all of the CGN programmed cell death models (KCl and serum withdrawal, KCl withdrawal alone, and kainate treatment). Figure 4A shows a self-organizing map (SOM) algorithm (See, e.g. Kohonen, Self Organizing Maps: Springer, Berlin (1997)) that was used to cluster genes based on expression in 26 experiments (in order: serum added back, 1, 3, 6, 12, 24 hours; KCl/serum withdrawal, 1, 3, 6, 12, 24 hours; controls for KCl withdrawal, 1, 3, 6, 12 hours; KCl withdrawal alone, 1, 3, 6, 12 hours; controls for kainate treatment, 2, 4, 8, 12 hours; kainate treatment, 2, 4, 8, 12 hours; see examples for experimental details). As shown, a 5 x 4 geometry was used to organize the genes into 20 groups. A cluster (3,3) of 17 programmed cell death-induced genes is highlighted. The inset shows a tiled depiction of all the genes in the (3,3) cluster; red = above mean expression, white = mean expression, blue = below mean expression; the tiles are ordered in columns as indicated above for experimental order; each row represents a different array element gene in the order indicated by distance from the cluster centroid. Caspase 3, a gene involved in apoptosis, is part of the array and depicted in the raw values graph (i.e. relative expression in the 26 experiments); each experiment is represented in order on the x-axis; the y-axis indicates gene expression intensity.

Figure 4 B, C, D, and E show the raw gene expression intensity plotted for a representative gene from programmed cell death-regulated, regulated by KCl withdrawal only, immediate early genes, and serum-repressed constitutive expression classes, respectively. Each panel shows the data for a representative member of the cluster (indicated in the gene list by *), along with a list of genes included in the expression cluster.

Figure 4B shows the raw gene expression intensity for a gene representative from the list on the right. The graph shows increased expression with KCl and serum withdrawal, and kainate treatment. Accordingly, genes with these characteristics are designated "programmed cell death regulated." The list of genes with this pattern (on the chip) is shown on the right. Known genes include genes regulated in apoptosis.

Figure 4C lists genes which show increased expression after withdrawal of KCL or KCL and serum, but following kainate treatment. The list includes genes known to be involved in apoptosis.

Figure 4D shows genes that demonstrate constitutive immediate early expression.

Figure 4E shows genes that demonstrate constitutive expression in the absence of serum. The list on the right shows that this class contains mediators of programmed cell death.

Figure 5 shows information relating to various NARC genes. Accordingly the first column gives the NARC (neuronal apoptosis regulated candidate) designation. The second column provides specific information, such as the number of nucleotides sequenced, the region sequenced, for example, the 3' untranslated region, information regarding open reading frames, information regarding human orthologs (whose sequences may also be found in SEQ ID NOS: 1-6, 8, and 10), information regarding homology to known amino acid or nucleotide sequences, information regarding function, and other information related to specific physical or functional characteristics. The third column shows the gene expression class as described and designated in Figure 4. The fourth column shows the results of Northern blot hybridization, for example whether expression is restricted to specific organs or ubiquitous, and transcript size.

Figure 6 shows a tabulation of expression data of genes known to be related to programmed cell death, the data being obtained from experiments disclosed herein wherein nucleic acid sequences on the microarray were hybridized to mRNA derived from the two programmed cell death models (see examples). The first column indicates the clone designation. Where the clone is a previously known gene (for example, c-fos and c-jun), the gene name is given rather than the cDNA clone designation. The second column indicates the gene designation for each clone based on a BLASTX search. The third column indicates the expression pattern for each of

the clones. This tabulation can serve as an internal control to assess the fidelity of the experimental conditions and thus can serve as a background to compare the expression pattern of uncharacterized clones in the array. Accordingly, this figure shows a subarray that can serve as an internal control for discovering genes related to apoptosis and cell proliferation.

Figure 7 shows all genes (i.e., that are represented by nucleic acid sequences on the chip) that are regulated in specific experimental conditions described in the examples and shown in Figure 4. Specific genes are clustered (in an underlined category). Each cluster represents clones having a specific expression characteristic. For example, the first cluster is transiently down-regulated by serum and down-regulated by KCl withdrawal. The second column identifies cDNA clones whose function is previously known. The third column indicates the cluster number. See Figure 4A. In addition, an analysis of the functions of the genes in each cluster showed that within a cluster, certain functional classes of genes may be over-represented. Thus, the material in parentheses indicates the biological functions that are associated with a disproportionate number of genes in the cluster. This includes secretion and synaptic vesicle release (cluster 0,0), cell proliferation (cluster 0,3), secretion/synaptic vesicle release/cytoskeletal reorganization (cluster 1,0), stress response/hormone response (cluster 1,3), stress response/hormone response (cluster 1,4), calcium signal transduction (cluster 2,0), and cytoskeleton/synapse cytoskeleton (cluster 2,4).

Figure 8 summarizes tissue expression data for the Smart Chip I™ microarray elements. The data were obtained by membrane blotting of the microarray against mRNA from testes, brain, heart, smooth muscle, spleen, kidney, skeletal muscle, lung, liver, and pancreatic tissue. Following hybridization with labeled cDNA synthesized from RNA from the indicated tissue type, the signal from each sequence on the array filters was quantitated by phosphorimaging.

Figure 9 provides a list of genes that were shown to be regulated by KCl and serum withdrawal in the microarray experiments described herein.

DETAILED DESCRIPTION OF THE INVENTION

I. Isolated Nucleic Acid Molecules

5 The invention encompasses the discovery and isolation of nucleic acid molecules that are expressed in rat brain and in programmed cell death *in vitro* models (neuronal apoptosis regulated candidates or NARCs) and their human homologs. The sequences of these human homologs are specifically disclosed in SEQ ID NOS:1 (human NARC 9B), 2 (human NARC 8B), 3 (human NARC 2A), 4 (human NARC 10B), 5 (human NARC 10C), 6 (human NARC 1C), 8 (human NARC 1A), and 10 (human NARC 25).

As appropriate, the isolated nucleic acid molecules of the present invention can be RNA, for example, mRNA, or DNA, such as cDNA and genomic DNA. DNA molecules can be double-stranded or single-stranded; single stranded RNA or DNA 15 can be either the coding, or sense, strand or the non-coding, or antisense, strand. The nucleic acid molecule can include all or a portion of the coding sequence of the genes of the invention. Additionally, the nucleic acid molecule can be fused to a marker sequence, for example, a sequence that encodes a polypeptide to assist in isolation or purification of the polypeptide. Such sequences include, but are not limited to, those 20 which encode a glutathione-S-transferase (GST) fusion protein and those which encode a hemagglutinin A (HA) polypeptide marker from influenza.

An "isolated" nucleic acid molecule, as used herein, is one that is separated from nucleic acid which normally flanks the nucleic acid molecule in nature. With regard to genomic DNA, the term "isolated" refers to nucleic acid molecules which 25 are separated from the chromosome with which the genomic DNA is naturally associated. For example, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotides which flank the nucleic acid molecule in the genomic DNA of the cell from which the nucleic acid is derived.

30 Moreover, an isolated nucleic acid of the invention, such as a cDNA or RNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or chemical precursors or other chemicals when chemically synthesized. However, the nucleic acid molecule can be fused to other

coding or regulatory sequences and still be considered isolated. In some instances, the isolated material will form part of a composition (for example, a crude extract containing other substances), buffer system or reagent mix. In other circumstances, the material may be purified to essential homogeneity, for example as determined by
5 PAGE or column chromatography such as HPLC. Preferably, an isolated nucleic acid comprises at least about 50, 80 or 90% (on a molar basis) of all macromolecular species present.

Further, recombinant DNA contained in a vector is included in the definition of "isolated" as used herein. Also, isolated nucleic acid molecules include
10 recombinant DNA molecules in heterologous host cells, as well as partially or substantially purified DNA molecules in solution. "Isolated" nucleic acid molecules also encompass *in vivo* and *in vitro* RNA transcripts of the DNA molecules of the present invention.

The invention further provides variants of the isolated nucleic acid molecules
15 of the invention. Such variants can be naturally occurring, such as allelic variants (same locus), homologs (different locus), and orthologs (different organism), or may be constructed by recombinant DNA methods or by chemical synthesis. Such non-naturally occurring variants can be made using well-known mutagenesis techniques, including those applied to polynucleotides, cells, or organisms.
20 Accordingly, variants can contain nucleotide substitutions, deletions, inversions and/or insertions in either or both the coding and non-coding region of the nucleic acid molecule. Further, the variations can produce both conservative and non-conservative amino acid substitutions.

Typically, variants have a substantial identity with a nucleic acid molecule
25 selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements thereof. Particularly preferred are nucleic acid molecules and fragments which have at least about 60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99% or more identity
30 with nucleic acid molecules described herein.

Such nucleic acid molecules can be readily identified as being able to hybridize under stringent conditions to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the

complements thereof. In one embodiment, the variants hybridize under high stringency hybridization conditions (e.g., for selective hybridization) to a nucleotide sequence selected from the sequences shown in SEQ ID NOS:1-6, 8, and 10.

As used herein, the term "hybridizes under stringent conditions" describes conditions for hybridization and washing. Stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. Aqueous and nonaqueous methods are described in that reference and either can be used. A preferred, example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 50°C. Another example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 55°C. A further example of stringent hybridization conditions is hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 60°C. Preferably, stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 65°C. Particularly preferred stringency conditions (and the conditions that should be used if the practitioner is uncertain about what conditions should be applied to determine if a molecule is within a hybridization limitation of the invention) are 0.5M Sodium Phosphate, 7% SDS at 65°C, followed by one or more washes at 0.2X SSC, 1% SDS at 65°C. The hybridization step may be performed for 4, 8, 12, or 16 hours, and the wash steps are generally 15 minutes or 30 minutes in length.

The percent identity of two nucleotide or amino acid sequences can be determined by aligning the sequences for optimal comparison purposes (e.g., gaps can be introduced in the sequence of a first sequence). The nucleotides or amino acids at corresponding positions are then compared, and the percent identity between the two sequences is a function of the number of identical positions shared by the sequences. In certain embodiments, the length of a sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 60%, and even more preferably at least 70%, 80% or 90% of the length of the reference sequence. The actual comparison of the two sequences can be accomplished by well-known methods, for example, using a mathematical algorithm. A preferred, non-limiting

example of such a mathematical algorithm is described in Karlin *et al.* (1993) *Proc. Natl. Acad. Sci. USA*, 90:5873-5877. Such an algorithm is incorporated into the NBLAST and XBLAST programs (version 2.0) as described in Altschul *et al.* (1997) *Nucleic Acids Res.*, 25:389-3402. When utilizing BLAST and Gapped BLAST
5 programs, the default parameters of the respective programs (e.g., NBLAST) can be used. See <http://www.ncbi.nlm.nih.gov>. In one embodiment, parameters for sequence comparison can be set at score=100, wordlength=12, or can be varied (e.g., W=5 or W=20).

Another preferred, non-limiting example of a mathematical algorithm utilized
10 for the comparison of sequences is the algorithm of Myers and Miller, CABIOS (1989). Such an algorithm is incorporated into the ALIGN program (version 2.0) which is part of the CGC sequence alignment software package. When utilizing the ALIGN program for comparing amino acid sequences, a PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used. Additional
15 algorithms for sequence analysis are known in the art and include ADVANCE and ADAM as described in Torellis and Robotti (1994) *Comput. Appl. Biosci.* 10:3-5; and FASTA described in Pearson and Lipman (1988) *PNAS*, 85:2444-8.

In another embodiment, the percent identity between two amino acid sequences can be accomplished using the GAP program in the CGC software package
20 (available at <http://www.cgc.com>) using either a BLOSUM 63 matrix or a PAM250 matrix, and a gap weight of 12, 10, 8, 6, or 4 and a length weight of 2, 3, or 4. In yet another embodiment, the percent identity between two nucleic acid sequences can be accomplished using the GAP program in the CGC software package (available at <http://www.cgc.com>), using a gap weight of 50 and a length weight of 3.

25 The present invention also provides isolated nucleic acids that contain a fragment or portion that hybridizes under highly stringent conditions to a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS:1-6, 8, and 10. In one embodiment, the nucleic acid consists of a fragment of
30 nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements of the sequences shown in SEQ ID NOS:1-6, 8, and 10. The nucleic acid fragments of the invention are at least about 15, preferably at least about 18, 20, 23 or 25 nucleotides, and can be 30, 40, 50, 100, 200

or more nucleotides in length. Longer fragments, for example, 30 or more nucleotides in length, which encode antigenic proteins or polypeptides described herein are useful. Additionally, nucleotide sequences described herein can also be contigged (e.g., overlapped or joined) to produce longer sequences (see, for example, 5 <http://bozeman.mbt.washington.edu/phrap.docs/phrap.html>).

In a related aspect, the nucleic acid fragments of the invention are used as probes or primers in assays such as those described herein. "Probes" are oligonucleotides that hybridize in a base-specific manner to a complementary strand of nucleic acid. Such probes include polypeptide nucleic acids, as described in 10 Nielsen *et al.* (1991) *Science*, 254, 1497-1500. Typically, a probe comprises a region of nucleotide sequence that hybridizes under highly stringent conditions to at least about 15, typically about 20-25, and more typically about 40, 50 or 75 consecutive nucleotides of a nucleic acid selected from the group consisting of the sequences shown in SEQ ID NOS:1-6, 8, and 10 and the complements thereof. More typically, 15 the probe further comprises a label, e.g., radioisotope, fluorescent compound, enzyme, or enzyme co-factor.

As used herein, the term "primer" refers to a single-stranded oligonucleotide which acts as a point of initiation of template-directed DNA synthesis using well-known methods (e.g., PCR, LCR) including, but not limited to those described 20 herein. The appropriate length of the primer depends on the particular use, but typically ranges from about 15 to 30 nucleotides. The term "primer site" refers to the area of the target DNA to which a primer hybridizes. The term "primer pair" refers to a set of primers including a 5' (upstream) primer that hybridizes with the 5' end of the nucleic acid sequence to be amplified and a 3' (downstream) primer that hybridizes 25 with the complement of the sequence to be amplified.

The nucleic acid molecules of the invention such as those described above can be identified and isolated using standard molecular biology techniques and the sequence information provided in the sequences shown in SEQ ID NOS:1-6, 8, and 10. For example, nucleic acid molecules can be amplified and isolated by the 30 polymerase chain reaction using synthetic oligonucleotide primers designed based on one or more of the sequences provided in the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. See generally *PCR Technology: Principles and Applications for DNA Amplification* (ed. H.A. Erlich, Freeman Press, NY, NY,

1992); *PCR Protocols: A Guide to Methods and Applications* (Eds. Innis, *et al.* Academic Press, San Diego, CA, 1990); Mattila *et al.* (1991) *Nucleic Acids Res.* 19:4967; Eckert *et al.* (1991) *PCR Methods and Applications*, 1:17; PCR (eds. McPherson *et al.* IRL Press, Oxford); and U.S. Patent 4,683,202. The nucleic acid
5 molecules can be amplified using cDNA, mRNA or genomic DNA as a template, cloned into an appropriate vector and characterized by DNA sequence analysis.

Other suitable amplification methods include the ligase chain reaction (LCR) (see Wu and Wallace (1989) *Genomics*, 4:560, Landegren *et al.* (1988) *Science*, 241:1077, transcription amplification (Kwoh *et al.* (1989) *Proc. Natl. Acad. Sci. USA*, 86:1173), and self-sustained sequence replication (Guatelli *et al.* (1990) *Proc. Nat. Acad. Sci. USA*, 87:1874) and nucleic acid based sequence amplification (NASBA). The latter two amplification methods involve isothermal reactions based on isothermal transcription, which produce both single stranded RNA (ssRNA) and double stranded DNA (dsDNA) as the amplification products in a ratio of about 30 or
15 100 to 1, respectively.

The amplified DNA can be radiolabelled and used as a probe for screening a cDNA library, mRNA in zap express, ZIPLOX or other suitable vector. Corresponding clones can be isolated, DNA can obtained following *in vivo* excision, and the cloned insert can be sequenced in either or both orientations by art recognized
20 methods to identify the correct reading frame encoding a protein of the appropriate molecular weight. For example, the direct analysis of the nucleotide sequence of nucleic acid molecules of the present invention can be accomplished using well-known methods that are commercially available. See, for example, Sambrook *et al.* *Molecular Cloning, A Laboratory Manual* (2nd Ed., CSHP, New York 1989);
25 Zyskind *et al.* *Recombinant DNA Laboratory Manual*, (Acad. Press, 1988)). Using these or similar methods, the protein(s) and the DNA encoding the protein can be isolated, sequenced and further characterized.

Antisense nucleic acids of the invention can be designed using the nucleotide sequences of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and constructed
30 using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase

the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

Additionally, the nucleic acid molecules of the invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, *e.g.*, the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup *et al.* (1996) *Bioorganic & Medicinal Chemistry*, 4:5). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.*, DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup *et al.* (1996), *supra*; Perry-O'Keefe *et al.* (1996) *Proc. Natl. Acad. Sci. USA*, 93:14670. PNAs can be further modified, *e.g.*, to enhance their stability, specificity or cellular uptake, by attaching lipophilic or other helper groups to PNA, by the formation of

PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996), *supra*, Finn *et al.* (1996) *Nucleic Acids Res.* 24(17):3357-63, Mag *et al.* (1989) *Nucleic Acids Res.* 17:5973, and Peterser *et al.* (1975) *Bioorganic Med. Chem. Lett.* 5:1119.

The nucleic acid molecules and fragments of the invention can also include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, *e.g.*, Letsinger *et al.* (1989) *Proc. Natl. Acad. Sci. USA*, 86:6553-6556; Lemaitre *et al.* (1987) *Proc. Natl. Acad. Sci. USA*, 84:648-652; PCT Publication No. WO88/0918) or the blood brain barrier (see, *e.g.*, PCT Publication No. WO89/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (see, *e.g.*, Krol *et al.* (1988) *Bio-Techniques*, 6:958-976) or intercalating agents (see, *e.g.*, Zon (1988) *Pharm Res.* 5:539-549).

Uses of the nucleic acids of the invention are described in detail in below. In general, the isolated nucleic acid sequences can be used as molecular weight markers on Southern gels, and as chromosome markers which are labeled to map related gene positions. The nucleic acid sequences can also be used to compare with endogenous DNA sequences in patients to identify genetic disorders, and as probes, such as to hybridize and discover related DNA sequences or to subtract out known sequences from a sample. The nucleic acid sequences can further be used to derive primers for genetic fingerprinting, to raise anti-protein antibodies using DNA immunization techniques, and as an antigen to raise anti-DNA antibodies or elicit immune responses. Additionally, the nucleotide sequences of the invention can be used to identify and express recombinant proteins for analysis, characterization or therapeutic use, or as markers for tissues in which the corresponding protein is expressed, either constitutively, during tissue differentiation, or in disease states.

II. Vectors and Host Cells

Another aspect of the invention pertains to nucleic acid vectors containing a nucleic acid selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10. These vectors comprise a sequence of the invention that has been inserted in a sense or antisense orientation. As used herein, the term "vector"

refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors, expression vectors, are capable of directing the expression of genes to which they are operably linked. In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids (vectors). However, the invention is intended to include such other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses) that serve equivalent functions.

Preferred recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell. This means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operably linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (*e.g.*, polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and those which direct expression of the nucleotide sequence only in certain host cells (*e.g.*, tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be

transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein .

The recombinant expression vectors of the invention can be designed for
5 expression of a polypeptide of the invention in prokaryotic or eukaryotic cells, *e.g.*, bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors), yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *supra*. Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7
10 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein.
15 Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable
20 separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson (1988) *Gene*, 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia,
25 Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann *et al.* (1988) *Gene*, 69:301-315) and pET 11d (Studier *et al.* *Gene Expression Technology: Methods in Enzymology*, 185, Academic Press, San Diego,
30 California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid *trp-lac* fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 *gn10-lac* fusion promoter mediated by a coexpressed viral RNA polymerase (T7 *gn1*). This viral

polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 *gn1* gene under the transcriptional control of the *lacUV 5* promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to
5 express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, *Gene Expression Technology: Methods in Enzymology*, 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those
10 preferentially utilized in *E. coli* (Wada *et al.* (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari
15 *et al.* (1987) *EMBO J.* 6:229-234), pMFa (Kurjan and Herskowitz (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.* (1987) *Gene*, 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and pPicZ (InVitrogen Corp, San Diego, CA).

Alternatively, a nucleic acid of the invention can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of
20 proteins in cultured insect cells (*e.g.*, Sf 9 cells) include the pAc series (Smith *et al.* (1983) *Mol. Cell Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology*, 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian
25 expression vectors include pCDM8 (Seed (1987) *Nature*, 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both
30 prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook *et al. supra*.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (*e.g.*, tissue-specific regulatory elements are used to express the nucleic acid).

Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.* (1987) *Genes Dev.* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv. Immunol.* 43:235-275), in particular promoters of T cell
5 receptors (Winoto and Baltimore (1989) *EMBO J.* 8:729-733) and immunoglobulins (Banerji *et al.* (1983) *Cell*, 33:729-740; Queen and Baltimore (1983) *Cell*, 33:741-748), neuron-specific promoters (*e.g.*, the neurofilament promoter; Byrne and Ruddle (1989) *Proc. Natl. Acad. Sci. USA*, 86:5473-5477), pancreas-specific promoters (Edlund *et al.* (1985) *Science*, 230:912-916), and mammary gland-specific
10 promoters (*e.g.*, milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) *Science*, 249:374-379) and the alpha-fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev.* 3:537-546).

15 The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operably linked to at least one expression control element in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to an mRNA of the invention.

20 Regulatory sequences operably linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression
25 vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub *et al.* (*Reviews - Trends in Genetics, Vol. 1(1)* 1986).

30 Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but also to the progeny or potential

progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

5 A host cell can be any prokaryotic or eukaryotic cell. For example, a nucleic acid of the invention can be expressed in bacterial cells (*e.g.*, *E. coli*), insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

10 Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or
15 transfecting host cells can be found in Sambrook, *et al.* (*supra*), and other laboratory manuals.

20 For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (*e.g.*, for resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that nucleic acid of the invention
25 or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

30 A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) a polypeptide of the invention. Accordingly, the invention further provides methods for producing a polypeptide using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding a polypeptide of the invention has been introduced) in a suitable medium

such that the polypeptide is produced. In another embodiment, the method further comprises isolating the polypeptide from the medium or the host cell.

The host cells of the invention can also be used to produce nonhuman transgenic animals. For example, in one embodiment, a host cell of the invention is a
5 fertilized oocyte or an embryonic stem cell into which a nucleic acid of the invention have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous nucleotide sequences have been introduced into their genome or homologous recombinant animals in which endogenous nucleotide sequences have been altered. Such animals are useful for studying the
10 function and/or activity of the nucleotide sequence and polypeptide encoded by the sequence and for identifying and/or evaluating modulators of their activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include
15 non-human primates, sheep, dogs, cows, goats, chickens, amphibians, etc. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, an "homologous recombinant
20 animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, *e.g.*, an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing a nucleic
25 acid of the invention into the male pronuclei of a fertilized oocyte, *e.g.*, by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The sequence can be introduced as a transgene into the genome of a non-human animal. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression
30 of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to the transgene to direct expression of a polypeptide in particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are

described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, U.S. Patent No. 4,873,191 and in Hogan, *Manipulating the Mouse Embryo* (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified
5 based upon the presence of the transgene in its genome and/or expression of mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding the transgene can further be bred to other transgenic animals carrying other transgenes.

10 Homologously recombinant host cells can also be produced that allow the *in situ* alteration of endogenous polynucleotide sequences of the invention in a host cell genome. The host cell includes, but is not limited to, a stable cell line, cell *in vivo*, or cloned microorganism. This technology is more fully described in WO 93/09222, WO 91/12650, WO 91/06667, U.S. 5,272,071, and U.S. 5,641,670. Briefly, specific
15 polynucleotide sequences corresponding to the polynucleotides or sequences proximal or distal to a gene are allowed to integrate into a host cell genome by homologous recombination where expression of the gene can be affected. In one embodiment, regulatory sequences are introduced that either increase or decrease expression of an endogenous sequence. Accordingly, a protein can be produced in a cell not normally
20 producing it. Alternatively, increased expression of a protein can be effected in a cell normally producing the protein at a specific level. Further, expression can be decreased or eliminated by introducing a specific regulatory sequence. The regulatory sequence can be heterologous to the protein sequence or can be a homologous sequence with a desired mutation that affects expression. Alternatively, the entire gene can be deleted.
25 The regulatory sequence can be specific to the host cell or capable of functioning in more than one cell type. Still further, specific mutations can be introduced into any desired region of the gene to produce mutant proteins of the invention. Such mutations could be introduced, for example, into the specific functional regions.

To create an homologous recombinant animal, a vector is prepared which
30 contains at least a portion of a nucleic acid of the invention into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, functionally disrupt, the endogenous gene. In one embodiment, the vector is designed such that, upon homologous recombination, the endogenous gene is functionally disrupted (*i.e.*, no

longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous gene is mutated or otherwise altered but still encodes functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous protein). In the homologous recombination vector, the altered portion of the gene is flanked at its 5' and 3' ends by additional nucleic acid of the gene to allow for homologous recombination to occur between the exogenous gene carried by the vector and an endogenous gene in an embryonic stem cell. The additional flanking nucleic acid is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the vector (see, e.g., Thomas and Capecchi (1987) *Cell* 51:503 for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced nucleic acid has homologously recombined with the endogenous gene are selected (see, e.g., Li *et al.* (1992) *Cell* 69:915). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see, e.g., Bradley in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination vectors and homologous recombinant animals are described further in Bradley (1991) *Current Opinion in Bio/Technology* 2:823-829 and in PCT Publication Nos. WO 90/11354, WO 91/01140, WO 92/0968, and WO 93/04169.

In another embodiment, transgenic non-human animals can be produced which contain selected systems that allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.* (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate

expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut *et al.* (1997) *Nature* 385:810-813 and PCT Publication Nos. WO 97/07668 and WO 97/07669.

10 III. Polypeptides

The present invention also provides isolated polypeptides and variants and fragments thereof that are encoded by the nucleic acid molecules of the invention, especially as shown in SEQ ID NOS: 1-6, 8, and 10. For example, as described above, the nucleotide sequences can be used to design primers to clone and express cDNAs encoding the polypeptides of the invention. Further, the nucleotide sequences of the invention, *e.g.*, the sequences shown in SEQ ID NOS: 1-6, 8, and 10, can be analyzed using routine search algorithms (*e.g.*, BLAST, Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410; BLAZE, Brutlag *et al.* (1993) *Comp. Chem.* 17:203-207) to identify open reading frames (ORFs).

As used herein, a polypeptide is said to be "isolated" or "purified" when it is substantially free of cellular material when it is isolated from recombinant and non-recombinant cells, or free of chemical precursors or other chemicals when it is chemically synthesized. A polypeptide, however, can be joined to another polypeptide with which it is not normally associated in a cell and still be "isolated" or "purified."

The polypeptides of the invention can be purified to homogeneity. It is understood, however, that preparations in which the polypeptide is not purified to homogeneity are useful and considered to contain an isolated form of the polypeptide. The critical feature is that the preparation allows for the desired function of the polypeptide, even in the presence of considerable amounts of other components. Thus, the invention encompasses various degrees of purity. In one embodiment, the language "substantially free of cellular material" includes preparations of the polypeptide having less than about 30% (by dry weight) other proteins (*i.e.*,

contaminating protein), less than about 20% other proteins, less than about 10% other proteins, or less than about 5% other proteins.

When a polypeptide is recombinantly produced, it can also be substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, less than about 10%, or less than about 5% of the volume of the protein preparation. The language "substantially free of chemical precursors or other chemicals" includes preparations of the polypeptide in which it is separated from chemical precursors or other chemicals that are involved in its synthesis. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of the polypeptide having less than about 30% (by dry weight) chemical precursors or other chemicals, less than about 20% chemical precursors or other chemicals, less than about 10% chemical precursors or other chemicals, or less than about 5% chemical precursors or other chemicals.

In one embodiment, a polypeptide comprises an amino acid sequence encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. However, the invention also encompasses sequence variants. Variants include a substantially homologous protein encoded by the same genetic locus in an organism, *i.e.*, an allelic variant. Variants also encompass proteins derived from other genetic loci in an organism, but having substantial homology to a polypeptide encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10 and the complements thereof. Variants also include proteins substantially homologous to these polypeptides but derived from another organism, *i.e.*, an ortholog. Variants also include proteins that are substantially homologous to these polypeptides that are produced by chemical synthesis. Variants also include proteins that are substantially homologous or identical to these polypeptides that are produced by recombinant methods.

As used herein, two proteins (or a region of the proteins) are substantially homologous or identical when the amino acid sequences are at least about 45-55%, typically at least about 70-75%, more typically at least about 80-85%, and most typically at least about 90, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical. A substantially homologous amino acid sequence, according to the present invention, will be encoded by a nucleic acid hybridizing to a nucleic acid

sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, or fragment thereof under stringent conditions as more described above.

To determine the percent similarity or identity of two amino acid sequences, or of two nucleic acids, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the sequence of one protein or nucleic acid for optimal alignment with the other protein or nucleic acid). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in one sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the other sequence, then the molecules are homologous at that position. As used herein, amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity". The percent homology between the two sequences is a function of the number of identical positions shared by the sequences (*i.e.*, per cent homology equals the number of identical positions/total number of positions times 100).

The invention also encompasses polypeptides having a lower degree of identity but having sufficient similarity so as to perform one or more of the same functions performed by a polypeptide encoded by a nucleic acid of the invention. Similarity is determined by conserved amino acid substitution. Such substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Conservative substitutions are likely to be phenotypically silent. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu, and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr. Guidance concerning which amino acid changes are likely to be phenotypically silent are found in Bowie *et al.* (1990) *Science* 247:1306-1310.

TABLE 1. Conservative Amino Acid Substitutions.

Aromatic	Phenylalanine
	Tryptophan
	Tyrosine
Hydrophobic	Leucine
	Isoleucine
	Valine
Polar	Glutamine
	Asparagine
Basic	Arginine
	Lysine
	Histidine
Acidic	Aspartic Acid
	Glutamic Acid
Small	Alanine
	Serine
	Threonine
	Methionine
	Glycine

- Both identity and similarity can be readily calculated (*Computational*
- 5 *Molecular Biology*, Lesk, A.M., ed., Oxford University Press, New York, 1988;
- Biocomputing: Informatics and Genome Projects*, Smith, D.W., ed., Academic Press,
- New York, 1993; *Computer Analysis of Sequence Data, Part 1*, Griffin, A.M., and
- Griffin, H.G., eds., Humana Press, New Jersey, 1994; *Sequence Analysis in Molecular*
- Biology*, von Heinje, G., Academic Press, 1987; and *Sequence Analysis Primer*,
- 10 Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991).

Preferred computer program methods to determine identify and similarity between two sequences include, but are not limited to, GCG program package (Devereux, J., *et al.* (1984) *Nucleic Acids Res.* 12(1):387), BLASTP, BLASTN, FASTA (Atschul, S.F. *et al.* (1990) *J. Molec. Biol.* 215:403).

A variant polypeptide can differ in amino acid sequence by one or more substitutions, deletions, insertions, inversions, fusions, and truncations or a combination of any of these. Further, variant polypeptides can be fully functional or can lack function in one or more activities. Fully functional variants typically contain only conservative variation or variation in non-critical residues or in non-critical regions. Functional variants can also contain substitution of similar amino acids that result in no change or an insignificant change in function. Alternatively, such substitutions may positively or negatively affect function to some degree.

Non-functional variants typically contain one or more non-conservative amino acid substitutions, deletions, insertions, inversions, or truncation or a substitution, insertion, inversion, or deletion in a critical residue or critical region.

As indicated, variants can be naturally-occurring or can be made by recombinant means or chemical synthesis to provide useful and novel characteristics for the polypeptide. This includes preventing immunogenicity from pharmaceutical formulations by preventing protein aggregation.

Amino acids that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham *et al.* (1989) *Science* 244:1081-1085). The latter procedure introduces single alanine mutations at every residue in the molecule. The resulting mutant molecules are then tested for biological activity *in vitro*, or *in vitro* proliferative activity. Sites that are critical for polypeptide activity can also be determined by structural analysis such as crystallization, nuclear magnetic resonance or photoaffinity labeling (Smith *et al.* (1992) *J. Mol. Biol.* 224:899-904; de Vos *et al.* (1992) *Science* 255:306-312).

The invention also includes polypeptide fragments of the polypeptides of the invention. Fragments can be derived from a polypeptide encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, and the complements thereof. However, the invention also encompasses fragments of the variants of the polypeptides described herein.

As used herein, a fragment comprises at least 6 contiguous amino acids. Useful fragments include those that retain one or more of the biological activities of

the polypeptide as well as fragments that can be used as an immunogen to generate polypeptide specific antibodies.

Biologically active fragments (peptides which are, for example, 6, 9, 12, 15, 20, 30, 35, 36, 37, 38, 39, 40, 50, 100 or more amino acids in length) can comprise a domain, segment, or motif that has been identified by analysis of the polypeptide sequence using well-known methods, *e.g.*, signal peptides, extracellular domains, one or more transmembrane segments or loops, ligand binding regions, zinc finger domains, DNA binding domains, acylation sites, glycosylation sites, or phosphorylation sites.

The invention also provides fragments with immunogenic properties. These contain an epitope-bearing portion of the polypeptides and variants of the invention. These epitope-bearing peptides are useful to raise antibodies that bind specifically to a polypeptide or region or fragment. These peptides can contain at least 6, 7, 8, 9, 12, at least 14, or between at least about 15 to about 30 amino acids. The epitope-bearing peptide and polypeptides may be produced by any conventional means (Houghten (1985) *Proc. Natl. Acad. Sci. USA* 82:5131-5135). Simultaneous multiple peptide synthesis is described in U.S. Patent No. 4,631,211.

Fragments can be discrete (not fused to other amino acids or polypeptides) or can be within a larger polypeptide. Further, several fragments can be comprised within a single larger polypeptide. In one embodiment a fragment designed for expression in a host can have heterologous pre- and pro-polypeptide regions fused to the amino terminus of the polypeptide fragment and an additional region fused to the carboxyl terminus of the fragment.

The invention thus provides chimeric or fusion proteins. These comprise a polypeptide of the invention operatively linked to a heterologous protein having an amino acid sequence not substantially homologous to the polypeptide. "Operatively linked" indicates that the polypeptide protein and the heterologous protein are fused in-frame. The heterologous protein can be fused to the N-terminus or C-terminus of the polypeptide. In one embodiment the fusion protein does not affect function of the polypeptide *per se*. For example, the fusion protein can be a GST-fusion protein in which the polypeptide sequences are fused to the C-terminus of the GST sequences. Other types of fusion proteins include, but are not limited to, enzymatic fusion proteins, for example beta-galactosidase fusions, yeast two-hybrid GAL fusions,

poly-His fusions and Ig fusions. Such fusion proteins, particularly poly-His fusions, can facilitate the purification of recombinant polypeptide. In certain host cells (*e.g.*, mammalian host cells), expression and/or secretion of a protein can be increased by using a heterologous signal sequence. Therefore, in another embodiment, the fusion protein contains a heterologous signal sequence at its N-terminus.

EP-A-O 464 533 discloses fusion proteins comprising various portions of immunoglobulin constant regions. The Fc is useful in therapy and diagnosis and thus results, for example, in improved pharmacokinetic properties (EP-A 0232 262). In drug discovery, for example, human proteins have been fused with Fc portions for the purpose of high-throughput screening assays to identify antagonists. Bennett *et al.* (1995) *Journal of Molecular Recognition* 8:52-58 and Johanson *et al.* (1995) *The Journal of Biological Chemistry* 270,16:9459-9471. Thus, this invention also encompasses soluble fusion proteins containing a polypeptide of the invention and various portions of the constant regions of heavy or light chains of immunoglobulins of various subclass (IgG, IgM, IgA, IgE). Preferred as immunoglobulin is the constant part of the heavy chain of human IgG, particularly IgG1, where fusion takes place at the hinge region. For some uses it is desirable to remove the Fc after the fusion protein has been used for its intended purpose, for example when the fusion protein is to be used as antigen for immunizations. In a particular embodiment, the Fc part can be removed in a simple way by a cleavage sequence that is also incorporated and can be cleaved with factor Xa.

A chimeric or fusion protein can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different protein sequences are ligated together in-frame in accordance with conventional techniques. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of nucleic acid fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive nucleic acid fragments which can subsequently be annealed and re-amplified to generate a chimeric nucleic acid sequence (see Ausubel *et al.*, *Current Protocols in Molecular Biology*, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST protein). A nucleic acid encoding a polypeptide of the invention can be

cloned into such an expression vector such that the fusion moiety is linked in-frame to the polypeptide protein.

The isolated polypeptide can be purified from cells that naturally express it, purified from cells that have been altered to express it (recombinant), or synthesized
5 using known protein synthesis methods.

In one embodiment, the protein is produced by recombinant DNA techniques. For example, a nucleic acid molecule encoding the polypeptide is cloned into an expression vector, the expression vector introduced into a host cell and the protein expressed in the host cell. The protein can then be isolated from the cells by an
10 appropriate purification scheme using standard protein purification techniques.

Polypeptides often contain amino acids other than the 20 amino acids commonly referred to as the 20 naturally-occurring amino acids. Further, many amino acids, including the terminal amino acids, may be modified by natural processes, such as processing and other post-translational modifications, or by
15 chemical modification techniques well known in the art. Common modifications that occur naturally in polypeptides are described in basic texts, detailed monographs, and the research literature, and they are well known to those of skill in the art.

Accordingly, the polypeptides also encompass derivatives or analogs in which a substituted amino acid residue is not one encoded by the genetic code, in which a
20 substituent group is included, in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence for purification of the mature polypeptide or a pro-protein sequence.

25 Known modifications include, but are not limited to, acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of
30 covalent crosslinks, formation of cystine, formation of pyroglutamate, formylation, gamma carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing,

phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

Such modifications are well-known to those of skill in the art and have been described in great detail in the scientific literature. Several particularly common
5 modifications, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic texts, such as *Proteins - Structure and Molecular Properties*, 2nd Ed., T.E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as by Wold, F., *Posttranslational*
10 *Covalent Modification of Proteins*, B.C. Johnson, Ed., Academic Press, New York 1-12 (1983); Seifter *et al.*, *Meth. Enzymol.* 182: 626-646 (1990) and Rattan *et al.* (1992) *Ann. N.Y. Acad. Sci.* 663:48-62.

As is also well known, polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination, and they may be
15 circular, with or without branching, generally as a result of post-translation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and branched circular polypeptides may be synthesized by non-translational natural processes and by synthetic methods.

Modifications can occur anywhere in a polypeptide, including the peptide
20 backbone, the amino acid side-chains and the amino or carboxyl termini. Blockage of the amino or carboxyl group in a polypeptide, or both, by a covalent modification, is common in naturally-occurring and synthetic polypeptides. For instance, the amino terminal residue of polypeptides made in *E. coli*, prior to proteolytic processing, almost invariably will be N-formylmethionine.

25 The modifications can be a function of how the protein is made. For recombinant polypeptides, for example, the modifications will be determined by the host cell posttranslational modification capacity and the modification signals in the polypeptide amino acid sequence. Accordingly, when glycosylation is desired, a polypeptide should be expressed in a glycosylating host, generally a eukaryotic cell.
30 Insect cells often carry out the same posttranslational glycosylations as mammalian cells and, for this reason, insect cell expression systems have been developed to efficiently express mammalian proteins having native patterns of glycosylation. Similar considerations apply to other modifications.

The same type of modification may be present in the same or varying degree at several sites in a given polypeptide. Also, a given polypeptide may contain more than one type of modification.

Uses of the polypeptides of the invention are described in detail below. In general, polypeptides or proteins of the present invention can be used as a molecular weight marker on SDS-PAGE gels or on molecular sieve gel filtration columns using art-recognized methods. The polypeptides of the present invention can be used to raise antibodies or to elicit an immune response. The polypeptides can also be used as a reagent, *e.g.*, a labeled reagent, in assays to quantitatively determine levels of the protein or a molecule to which it binds (*e.g.*, a receptor or a ligand) in biological fluids. The polypeptides can also be used as markers for tissues in which the corresponding protein is preferentially expressed, either constitutively, during tissue differentiation, or in a diseased state. The polypeptides can be used to isolate a corresponding binding partner, *e.g.*, receptor or ligand, such as, for example, in an interaction trap assay, and to screen for peptide or small molecule antagonists or agonists of the binding interaction.

IV. Antibodies

In another aspect, the invention provides antibodies to the polypeptides and polypeptide fragments of the invention, *e.g.*, having an amino acid encoded by a nucleic acid comprising all or a portion of a nucleotide sequence selected from the group consisting of the sequences shown in SEQ ID NOS: 1-6, 8, and 10. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site that specifically binds an antigen. A molecule that specifically binds to a polypeptide of the invention is a molecule that binds to that polypeptide or a fragment thereof, but does not substantially bind other molecules in a sample, *e.g.*, a biological sample, which naturally contains the polypeptide. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind to a polypeptide of the invention. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that

contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of a polypeptide of the invention. A monoclonal antibody composition thus typically displays a single binding affinity for a particular polypeptide of the invention with which it immunoreacts.

5 Polyclonal antibodies can be prepared as described above by immunizing a suitable subject with a desired immunogen, *e.g.*, polypeptide of the invention or fragment thereof. The antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized polypeptide. If desired, the antibody molecules directed
10 against the polypeptide can be isolated from the mammal (*e.g.*, from the blood) and further purified by well-known techniques, such as protein A chromatography to obtain the IgG fraction. At an appropriate time after immunization, *e.g.*, when the antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the
15 hybridoma technique originally described by Kohler and Milstein (1975) *Nature* 256:495-497, the human B cell hybridoma technique (Kozbor *et al.* (1983) *Immunol. Today* 4:72), the EBV-hybridoma technique (Cole *et al.* (1985), *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing hybridomas is well known (see generally *Current*
20 *Protocols in Immunology* (1994) Coligan *et al.* (eds.) John Wiley & Sons, Inc., New York, NY). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with an immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds a
25 polypeptide of the invention.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating a monoclonal antibody to a polypeptide of the invention (see, *e.g.*, *Current Protocols in Immunology, supra*; Galfre *et al.* (1977) *Nature* 266:55052; R.H. Kenneth, in
30 *Monoclonal Antibodies: A New Dimension In Biological Analyses*, Plenum Publishing Corp., New York, New York (1980); and Lerner (1981) *Yale J. Biol. Med.* 54:387-402. Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods that also would be useful. Typically, the immortal

cell line (*e.g.*, a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line, *e.g.*, a myeloma cell line that is sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a number of myeloma cell lines can be used as a fusion partner according to standard techniques, *e.g.*, the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind a polypeptide of the invention, *e.g.*, using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal antibody to a polypeptide of the invention can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (*e.g.*, an antibody phage display library) with the polypeptide to thereby isolate immunoglobulin library members that bind the polypeptide. Kits for generating and screening phage display libraries are commercially available (*e.g.*, the Pharmacia *Recombinant Phage Antibody System*, Catalog No. 27-9400-01; and the Stratagene *SurfZAP™* Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. Patent No. 5,223,409; PCT Publication No. WO 92/18619; PCT Publication No. WO 91/17271; PCT Publication No. WO 92/20791; PCT Publication No. WO 92/15679; PCT Publication No. WO 93/01288; PCT Publication No. WO 92/01047; PCT Publication No. WO 92/09690; PCT Publication No. WO 90/02809; Fuchs *et al.* (1991) *Bio/Technology* 9:1370-1372; Hay *et al.* (1992) *Hum. Antibod. Hybridomas* 3:81-85; Huse *et al.* (1989) *Science* 246:1275-1281; Griffiths *et al.* (1993) *EMBO J.* 12:725-734.

Additionally, recombinant antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can

be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in PCT Publication No. WO 87/02671; European Patent Application 184,187;

- 5 European Patent Application 171,496; European Patent Application 173,494; PCT Publication No. WO 86/01533; U.S. Patent No. 4,816,567; European Patent Application 125,023; Better *et al.* (1988) *Science* 240:1041-1043; Liu *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:3439-3443; Liu *et al.* (1987) *J. Immunol.* 139:3521-3526; Sun *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:214-218; Nishimura *et al.* (1987) *Canc. Res.* 47:999-1005; Wood *et al.* (1985) *Nature* 314:446-449; and
- 10 Shaw *et al.* (1988) *J. Natl. Cancer Inst.* 80:1553-1559; Morrison (1985) *Science* 229:1202-1207; Oi *et al.* (1986) *Bio/Techniques* 4:214; U.S. Patent 5,225,539; Jones *et al.* (1986) *Nature* 321:552-525; Verhoeyan *et al.* (1988) *Science* 239:1534; and Beidler *et al.* (1988) *J. Immunol.* 141:4053-4060.

- 15 Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Such antibodies can be produced using transgenic mice that are incapable of expressing endogenous immunoglobulin heavy and light chains genes, but which can express human heavy and light chain genes. The transgenic mice are immunized in the normal fashion with a selected antigen, *e.g.*, all or a
- 20 portion of a polypeptide of the invention. Monoclonal antibodies directed against the antigen can be obtained using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA
- 25 and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg and Huszar (1995) *Int. Rev. Immunol.* 13:65-93. For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, *e.g.*, U.S. Patent 5,625,126; U.S. Patent 5,633,425; U.S. Patent 5,569,825; U.S. Patent
- 30 5,661,016; and U.S. Patent 5,545,806. In addition, companies such as Abgenix, Inc. (Freemont, CA), can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

Completely human antibodies that recognize a selected epitope can be generated using a technique referred to as "guided selection." This technology is described, for example, in Jespers *et al.* (1994) *Bio/technology* 12:899-903).

Uses of the antibodies of the invention are described in detail below. In general, antibodies of the invention (*e.g.*, a monoclonal antibody) can be used to isolate a polypeptide of the invention by standard techniques, such as affinity chromatography or immunoprecipitation. A polypeptide specific antibody can facilitate the purification of natural polypeptide from cells and of recombinantly produced polypeptide expressed in host cells. Moreover, an antibody specific for a polypeptide of the invention can be used to detect the polypeptide (*e.g.*, in a cellular lysate, cell supernatant, or tissue sample) in order to evaluate the abundance and pattern of expression of the polypeptide. Antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, (-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H .

V. Computer Readable Means

The nucleotide or amino acid sequences of the invention are also provided in a variety of mediums to facilitate use thereof. As used herein, "provided" refers to a manufacture, other than an isolated nucleic acid or amino acid molecule, which contains a nucleotide or amino acid sequence of the present invention. Such a manufacture provides the nucleotide or amino acid sequences, or a subset thereof (*e.g.*, a subset of open reading frames (ORFs)) in a form which allows a skilled artisan

to examine the manufacture using means not directly applicable to examining the nucleotide or amino acid sequences, or a subset thereof, as they exists in nature or in purified form.

In one application of this embodiment, a nucleotide or amino acid sequence of the present invention can be recorded on computer readable media. As used herein, "computer readable media" refers to any medium that can be read and accessed directly by a computer. Such media include, but are not limited to: magnetic storage media, such as floppy discs, hard disc storage medium, and magnetic tape; optical storage media such as CD-ROM; electrical storage media such as RAM and ROM; and hybrids of these categories such as magnetic/optical storage media. The skilled artisan will readily appreciate how any of the presently known computer readable mediums can be used to create a manufacture comprising computer readable medium having recorded thereon a nucleotide or amino acid sequence of the present invention.

As used herein, "recorded" refers to a process for storing information on computer readable medium. The skilled artisan can readily adopt any of the presently known methods for recording information on computer readable medium to generate manufactures comprising the nucleotide or amino acid sequence information of the present invention.

A variety of data storage structures are available to a skilled artisan for creating a computer readable medium having recorded thereon a nucleotide or amino acid sequence of the present invention. The choice of the data storage structure will generally be based on the means chosen to access the stored information. In addition, a variety of data processor programs and formats can be used to store the nucleotide sequence information of the present invention on computer readable medium. The sequence information can be represented in a word processing text file, formatted in commercially-available software such as WordPerfect and MicroSoft Word, or represented in the form of an ASCII file, stored in a database application, such as DB2, Sybase, Oracle, or the like. The skilled artisan can readily adapt any number of dataprocessor structuring formats (*e.g.*, text file or database) in order to obtain computer readable medium having recorded thereon the nucleotide sequence information of the present invention.

By providing the nucleotide or amino acid sequences of the invention in computer readable form, the skilled artisan can routinely access the sequence

information for a variety of purposes. For example, one skilled in the art can use the nucleotide or amino acid sequences of the invention in computer readable form to compare a target sequence or target structural motif with the sequence information stored within the data storage means. Search means are used to identify fragments or regions of the sequences of the invention which match a particular target sequence or target motif.

As used herein, a "target sequence" can be any DNA or amino acid sequence of six or more nucleotides or two or more amino acids. A skilled artisan can readily recognize that the longer a target sequence is, the less likely a target sequence will be present as a random occurrence in the database. The most preferred sequence length of a target sequence is from about 10 to 100 amino acids or from about 30 to 300 nucleotide residues. However, it is well recognized that commercially important fragments, such as sequence fragments involved in gene expression and protein processing, may be of shorter length.

As used herein, "a target structural motif," or "target motif," refers to any rationally selected sequence or combination of sequences in which the sequence(s) are chosen based on a three-dimensional configuration which is formed upon the folding of the target motif. There are a variety of target motifs known in the art. Protein target motifs include, but are not limited to, enzyme active sites and signal sequences. Nucleic acid target motifs include, but are not limited to, promoter sequences, hairpin structures and inducible expression elements (protein binding sequences).

Computer software is publicly available which allows a skilled artisan to access sequence information provided in a computer readable medium for analysis and comparison to other sequences. A variety of known algorithms are disclosed publicly and a variety of commercially available software for conducting search means are and can be used in the computer-based systems of the present invention. Examples of such software includes, but is not limited to, MacPattern (EMBL), BLASTN and BLASTX (NCBIA).

For example, software which implements the BLAST (Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410) and BLAZE (Brutlag *et al.* (1993) *Comp. Chem.* 17:203-207) search algorithms on a Sybase system can be used to identify open reading frames (ORFs) of the sequences of the invention which contain homology to ORFs or proteins from other libraries. Such ORFs are protein encoding fragments and are

useful in producing commercially important proteins such as enzymes used in various reactions and in the production of commercially useful metabolites.

VI. Detection Assays

5 Portions or fragments of the nucleotide sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue
10 typing); and (iii) aid in forensic identification of a biological sample. These applications are described in the subsections below.

1. Chromosome Mapping

 Once the nucleic acid (or a portion of the sequence) has been isolated, it can
15 be used to map the location of the gene on a chromosome. The mapping of the sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease. Briefly, genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the nucleic acid molecules described herein. Computer analysis of the sequences can be used to
20 predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the appropriate nucleotide sequences will yield an amplified fragment.

25 Somatic cell hybrids are prepared by fusing somatic cells from different mammals (*e.g.*, human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that
30 contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of

individual genes to specific human chromosomes. (D'Eustachio *et al.* (1983) *Science* 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

5 PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycle. Using the nucleic acid molecules of the invention to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can
10 similarly be used to map a specified sequence to its chromosome include *in situ* hybridization (described in Fan *et al.* (1990) *PNAS* 97:6223-27), pre-screening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

 Fluorescence *in situ* hybridization (FISH) of a nucleotide sequence to a
15 metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each
20 chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a nucleotide sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good
25 results at a reasonable amount of time. for a review of this technique, see Verma *et al.*, *Human Chromosomes: A Manual of Basic Techniques* (Pergamon Press, New York 1988).

 Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used
30 for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are preferred for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, *Medelian Inheritance in Man*, available on-line through Johns Hopkins University Welch Medical Library).

- 5 The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland *et al.* (1987) *Nature* 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with a specified gene, can be determined. If a
10 mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR
15 based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

2. Tissue Typing

20 The nucleotide sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield
25 unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an
30 alternative technique that determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the nucleic acid molecules described herein can be used to prepare two PCR primers from the 5' and 3' ends of

the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

5 Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The nucleic acid molecules of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals. The noncoding sequences of these sequences can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers which each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

20 If a panel of reagents from nucleic acid molecules described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

25

3. Use of Partial Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means of positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, *e.g.*, hair or skin, or body fluids, *e.g.*, blood, saliva, or semen found at a crime scene. The

amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

The sequences of the present invention can be used to provide polynucleotide reagents, *e.g.*, PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (*i.e.* another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Sequences targeted to noncoding regions of sequences described herein are particularly appropriate for this use, as greater numbers of polymorphisms occur in the noncoding regions, making it easier to differentiate individuals using this technique. Examples of polynucleotide reagents include the nucleic acid molecules or the invention, or portions thereof, *e.g.*, fragments having a length of at least 20 bases, preferably at least 30 bases.

The nucleic acid molecules described herein can further be used to provide polynucleotide reagents, *e.g.*, labeled or labelable probes which can be used in, or example, an *in situ* hybridization technique, to identify a specific tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, primers or probes can be used to screen tissue culture for contamination (*i.e.*, screen for the presence of a mixture of different types of cells in a culture).

VII. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining protein and/or nucleic acid expression as well as activity of proteins of the invention, in the context of a biological sample (*e.g.*, blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant expression or activity. The

invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with activity or expression of proteins or nucleic acids of the invention.

Disorders relating to programmed cell death are particularly relevant as
5 discussed in detail herein below.

For example, mutations in a specified gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby prophylactically treat an individual prior to the onset of a disorder characterized by or associated with expression or activity of nucleic acid molecules or proteins of the
10 invention.

Another aspect of the invention pertains to monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression or activity of proteins of the invention in clinical trials.

These and other agents are described in further detail in the following sections.
15

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of proteins or nucleic acids of the invention in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or
20 an agent capable of detecting the protein, or nucleic acid (*e.g.*, mRNA, genomic DNA) that encodes the protein, such that the presence of the protein or nucleic acid is detected in the biological sample. A preferred agent for detecting mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to mRNA or genomic DNA sequences described herein. The nucleic acid probe can be, for example, a
25 full-length nucleic acid, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to appropriate mRNA or genomic DNA. For example, the nucleic acid probe can be all or a portion of the sequences shown in SEQ ID NOS: 1-6, 8, and 10, or the complement of the sequences shown in SEQ ID NOS: 1-6, 8, and
30 10, or a portion thereof. Other suitable probes for use in the diagnostic assays of the invention are described herein.

In one embodiment, the agent for detecting proteins of the invention is an antibody capable of binding to the protein, preferably an antibody with a detectable

label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (*e.g.*, Fab or F(ab')₂) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (*i.e.*, physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, calls and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect mRNA, protein, or genomic DNA of the invention in a biological sample *in vitro* as well as *in vivo*. For example, *in vitro* techniques for detection of mRNA include Northern hybridizations and *in situ* hybridizations. *In vitro* techniques for detection of protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of genomic DNA include Southern hybridizations. Furthermore, *in vivo* techniques for detection of protein include introducing into a subject a labeled anti-protein antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a serum sample or biopsy isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting protein, mRNA, or genomic DNA of the invention, such that the presence of protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of protein, mRNA or genomic DNA in the control sample with the presence of protein, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting the presence of proteins or nucleic acid molecules of the invention in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting protein or mRNA in a biological sample; means for determining the amount of in the sample; and means for
5 comparing the amount of in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect protein or nucleic acid.

2. Prognostic Assays

10 The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant expression or activity of proteins and nucleic acid molecules of the invention. Accordingly, the term "diagnostic" refers not only to ascertaining whether a subject has an active disease but also relates to ascertaining whether a subject is predisposed
15 to developing active disease as well as ascertaining the probability that treatment of active disease will be effective. For example, the assays described herein, such as the preceding diagnostic assays or the following assays can be utilized to identify a subject having or at risk of developing a disorder associated with protein or nucleic acid expression or activity such as a proliferative disorder, a differentiative or
20 developmental disorder, or a hematopoietic disorder. Alternatively, the prognostic assays can be utilized to identify a subject having or at risk for developing a differentiative or proliferative disease (*e.g.*, cancer). Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant expression or activity of proteins or nucleic acid molecules of the invention, in which
25 a test sample is obtained from a subject and protein or nucleic acid (*e.g.*, mRNA, genomic DNA) is detected, wherein the presence of protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant expression or activity of the protein or nucleic acid sequence of the invention. As used herein, a "test sample" refers to a biological sample
30 obtained from a subject of interest. For example, a test sample can be a biological fluid (*e.g.*, serum), cell or tissue sample.

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, polypeptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant expression or activity of a protein or nucleic acid molecule of the invention. For example, such methods can be used to determine whether a subject can be effectively treated with an agent for a disorder, such as a proliferative disorder, a differentiative or a developmental disorder. Alternatively, such methods can be used to determine whether a subject can be effectively treated with an agent for a differentiative or proliferative disease (*e.g.*, cancer). Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant expression or activity of a protein or nucleic acid of the present invention, in which a test sample is obtained and protein or nucleic acid expression or activity is detected (*e.g.*, wherein the abundance of particular protein or nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant expression or activity.)

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

The methods of the invention can also be used to detect genetic alterations in genes or nucleic acid molecules of the present invention, thereby determining if a subject with the altered gene is at risk for a disorder characterized by aberrant development, aberrant cellular differentiation, aberrant cellular proliferation or an aberrant hematopoietic response. In certain embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding a particular protein, or the mis-expression of the gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of (1) a deletion of one or more nucleotides; (2) an addition of one or more nucleotides; (3) a substitution of one or more nucleotides, (4) a chromosomal rearrangement; (5) an alteration in the level of a messenger RNA transcript; (6) aberrant modification, such as of the methylation pattern of the genomic DNA; (7) the presence of a non-wild type splicing pattern of a messenger RNA transcript; (8) a non-wild type level; (9) allelic loss; and (10) inappropriate post-translational modification. As described

herein, there are a large number of assay techniques known in the art that can be used for detecting alterations in a particular gene. A preferred biological sample is a tissue or serum sample isolated by conventional means from a subject.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, *e.g.*, U.S. Patent Nos. 4,683,195 and 4,683,202), such an anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, *e.g.*, Landegran *et al.* (1988) *Science* 241:1077-1080; and Nakazawa *et al.* (1994) *PNAS* 91:360-364), the latter of which can be particularly useful for detecting point mutations (see Abravaya *et al.* (1995) *Nucleic Acids Res.* 23:675-682). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (*e.g.*, genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to the gene under conditions such that hybridization and amplification of the gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:1874-1878), transcriptional amplification system (Kwoh *et al.*, (1989) *Proc. Natl. Acad. Sci. USA* 86:1173-1177), Q-Beta Replicase (Lizardi *et al.* (1988) *Bio/Technology* 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in a given gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicate mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for sample, U.S. Patent No. 5,498,531) can be used

to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations can be identified by hybridizing a sample and control nucleic acids, *e.g.*, DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotide probes (Cronin *et al.* (1996) *Human Mutation* 7:244-255; Kozal *et al.* (1996) *Nature Medicine* 2:753-759). For example, genetic mutations can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. *et al. supra*. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the gene and detect mutations by comparing the sequence of the gene from the sample with the corresponding wild-type (control) gene sequence. Examples of sequencing reactions include those based on techniques developed by Maxim and Gilbert ((1997) *PNAS* 74:560) or Sanger ((1977) *PNAS* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, *e.g.*, PCT International Publication No. WO 94/16101; Cohen *et al.* (1996) *Adv. Chromatogr.* 36:127-162; and Griffin *et al.* (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

Other methods for detecting mutations include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers *et al.* (1985) *Science* 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wild-type sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-standard duplexes are treated with an agent that cleaves single-stranded regions of the duplex such as which

will exist due to base pair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with Rnase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digest the mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size
5 on denaturing polyacrylamide gels to determine the site of mutation. See, for example Cotton *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:4397; Saleeba *et al.* (1992) *Methods Enzymol.* 217:286-295. In certain embodiments, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or
10 more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994)
15 *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on an nucleotide sequence of the invention is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

20 In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:2766, see also Cotton (1993) *Mutat Res* 285:125-144; and Hayashi (1992) *Genet Anal. Tech.*
25 *Appl.* 9:73-79). Single-stranded DNA fragments of sample and control nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be
30 enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In one embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen *et al.* (1991) *Trends Genet.* 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers *et al.* (1985) *Nature* 313:495). When DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys. Chem.* 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki *et al.* (1986) *Nature* 324:163); Saiki *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:6320). Such allele-specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology that depends on selective PCR amplification may be used in conjunction with the instant invention. Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs *et al.* (1989) *Nucleic Acids Res.* 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner (1993) *Tibtech* 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini *et al.* (1992) *Mol. Cell Probes* 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) *Proc. Natl. Acad. Sci. USA* 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness

- 5 involving a gene of the present invention. Any cell type or tissue in which the gene is expressed may be utilized in the prognostic assays described herein.

3. Monitoring of Effects During Clinical Trials

- Monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression
10 or activity of nucleic acid molecules or proteins of the present invention (*e.g.*, modulation of cellular signal transduction, regulation of gene transcription in a cell involved in development or differentiation, regulation of cellular proliferation) can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to
15 increase gene expression, protein levels, or upregulate protein activity, can be monitored in clinical trials of subjects exhibiting decreased gene expression, protein levels, or downregulated protein activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease gene expression, protein levels, or downregulate protein activity, can be monitored in clinical trials of subjects exhibiting
20 increased gene expression, protein levels, or upregulated protein activity. In such clinical trials, the expression or activity of the specified gene and, preferably, other genes that have been implicated in, for example, a proliferative disorder can be used as a "read out" or markers of the phenotype of a particular cell.

- For example, and not by way of limitation, genes that are modulated in cells
25 by treatment with an agent (*e.g.*, compound, drug or small molecule) which modulates protein activity (*e.g.*, identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on proliferative disorders, developmental or differentiative disorder, or hematopoietic disorder, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of
30 expression of the specified gene and other genes implicated in the proliferative disorder, developmental or differentiative disorder, or hematopoietic disorder, respectively. The levels of gene expression (*i.e.*, a gene expression pattern) can be quantified by Northern blot analysis or RT-PCR, as described herein, or alternatively

by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of the specified gene or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may
5 be determined before, and at various points during, treatment of the individual with the agent.

Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

In one embodiment, the present invention provides a method for monitoring
10 the effectiveness of treatment of a subject with an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, polypeptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) comprising the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of a specified protein, mRNA, or genomic
15 DNA of the invention in the pre-administration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the protein, mRNA, or genomic DNA in the pre-administration sample with the protein, mRNA, or genomic DNA in
20 the post-administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of the protein or nucleic acid molecule to higher levels than detected, *i.e.*, to increase effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease
25 effectiveness of the agent. According to such an embodiment, protein or nucleic acid expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

VIII. Screening Assays

30 The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, antisense, polypeptides, peptidomimetics, small molecules or other drugs) which bind to nucleic acid molecules, polypeptides or proteins described herein or have a

stimulatory or inhibitory effect on, for example, expression or activity of the nucleic acid molecules, polypeptides or proteins of the invention.

As an example, apoptosis-specific assays may be used to identify modulators of any of the target nucleic acids or proteins of the present invention, which proteins
5 and/or nucleic acids are related to apoptosis. Accordingly, an agent that modulates the level or activity of any of these nucleic acids or proteins can be identified by means of apoptosis-specific assays. For example, high throughput screens exist to identify apoptotic cells by the use of chromatin or cytoplasmic-specific dyes. Thus, hallmarks of apoptosis, cytoplasmic condensation and chromosome fragmentation;
10 can be used as a marker to identify modulators of any of the genes related to programmed-cell death described herein. Other assays include, but are not limited to, the activation of specific endogenous proteases, loss of mitochondrial function, cytoskeletal disruption, cell shrinkage, membrane blebbing, and nuclear condensation due to degradation of DNA.

15 In one embodiment, the invention provides assays for screening candidate or test compounds that bind to or modulate the activity of protein or polypeptide described herein or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries;
20 spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to polypeptide libraries, while the other four approaches are applicable to polypeptide, non-peptide oligomer or small molecule libraries of
25 compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in DeWitt *et al.* (1993) *Proc. Natl. Acad. Sci. U.S.A.* 90:6909; Erb
et al. (1994) *Proc. Natl. Acad. Sci. U.S.A.* 91:11422; Zuckermann *et al.* (1994). *J. Med. Chem.* 37:2678; Cho *et al.* (1993) *Science* 261:1303; Carell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.* 33:2059; Carell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.*
30 33:2061; and in Gallop *et al.* (1994) *J. Med. Chem.* 37:1233.

Libraries of compounds may be presented in solution (*e.g.*, Houghten (1992) *Biotechniques* 13:412-421), or on beads (Lam (1991) *Nature* 354:82-84), chips (Fodor

(1993) *Nature* 364:555-556), bacteria (Ladner U.S. Patent No. 5,223,409), spores (Ladner USP '409), plasmids (Cull *et al.* (1992) *Proc. Natl. Acad. Sci. U.S.A.* 89:1865-1869) or on phage (Scott and Smith (1990) *Science* 249:386-390); (Devlin (1990) *Science* 249:404-406); (Cwirla *et al.* (1990) *Proc. Natl. Acad. Sci.* 87:6378-6382); (Felici (1991) *J. Mol. Biol.* 222:301-310); (Ladner *supra*).

In one embodiment, an assay is a cell-based assay in which a cell that expresses an encoded polypeptide (*e.g.*, cell surface protein such as a receptor) is contacted with a test compound and the ability of the test compound to bind to the polypeptide is determined. The cell, for example, can be of mammalian origin, such as a keratinocyte. Determining the ability of the test compound to bind to the polypeptide can be accomplished, for example, by coupling the test compound with a radioisotope or enzymatic label such that binding of the test compound to the polypeptide can be determined by detecting the labeled with ^{125}I , ^{35}S , ^{14}C , or ^3H , either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, test compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a test compound to interact with the polypeptide without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of a test compound with the polypeptide without the labeling of either the test compound or the polypeptide. McConnell *et al.* (1992) *Science* 257:1906-1912. As used herein, a "microphysiometer" (*e.g.*, CytosensorTM) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between ligand and polypeptide.

In one embodiment, the assay comprises contacting a cell which expresses an encoded protein described herein on the cell surface (*e.g.*, a receptor) with a polypeptide ligand or biologically-active portion thereof, to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the polypeptide, wherein determining the ability of the test compound to interact with the polypeptide comprises determining the ability of

the test compound to preferentially bind to the polypeptide as compared to the ability of the ligand, or a biologically active portion thereof, to bind to the polypeptide.

In another embodiment, an assay is a cell-based assay comprising contacting a cell expressing a particular target molecule described herein with a test compound and determining the ability of the test compound to modulate or alter (*e.g.* stimulate or inhibit) the activity of the target molecule. Determining the ability of the test compound to modulate the activity of the target molecule can be accomplished, for example, by determining the ability of a known ligand to bind to or interact with the target molecule. Determining the ability of the known ligand to bind to or interact with the target molecule can be accomplished by one of the methods described above for determining direct binding. In one embodiment, determining the ability of the known ligand to bind to or interact with the target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (*e.g.*, intracellular Ca^{2+} , diacylglycerol, IP_3 , etc.), detecting catalytic/enzymatic activity of the target on an appropriate substrate, detecting the induction of a reporter gene (comprising a target-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase), or detecting a cellular response, for example, development, differentiation or rate of proliferation.

In yet another embodiment, an assay of the present invention is a cell-free assay in which protein of the invention or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the protein or biologically active portion thereof is determined. Binding of the test compound to the protein can be determined either directly or indirectly as described above. In one embodiment, the assay includes contacting the protein or biologically active portion thereof with a known compound which binds the protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the protein. Determining the ability of the test compound to interact with the protein comprises determining the ability of the test compound to preferentially bind to the protein or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which a protein of the invention or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate or alter (*e.g.*, stimulate or inhibit) the activity of the protein or biologically active portion thereof is determined.

- 5 Determining the ability of the test compound to modulate the activity of the protein can be accomplished, for example, by determining the ability of the protein to bind to a known target molecule by one of the methods described above for determining direct binding. Determining the ability of the protein to bind to a target molecule can also be accomplished using a technology such as real-time Bimolecular Interaction
- 10 Analysis (BIA). Sjolander and Urbaniczky (1991) *Anal. Chem.* 63:2338-2345 and Szabo *et al.* (1995) *Curr. Opin. Struct. Biol.* 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (*e.g.*, BIAcore™). Changes in the optical phenomenon surface plasmon resonance (SPR) can be used as an indication of real-time reactions between
- 15 biological molecules.

- In an alternative embodiment, determining the ability of the test compound to modulate the activity of a protein of the invention can be accomplished by determining the ability of the protein to further modulate the activity of a target molecule. For example, the catalytic/enzymatic activity of the target molecule on an
- 20 appropriate substrate can be determined as previously described.

- In yet another embodiment, the cell-free assay involves contacting a protein of the invention or biologically active portion thereof with a known compound which binds the protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the
- 25 protein, wherein determining the ability of the test compound to interact with the protein comprises determining the ability of the protein to preferentially bind to or modulate the activity of a target molecule.

- The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of isolated proteins. In the case of cell-free
- 30 assays in which a membrane-bound form an isolated protein is used it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the isolated protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside,

n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton®X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPS), 3-[(3-cholamidopropyl)dimethylamminio]-2-hydroxy-1-propane sulfonate (CHAPSO), or N-dodecyl-N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either the protein or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to the protein, or interaction of the protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or protein of the invention, and the mixture incubated under conditions conducive to complex formation (*e.g.*, at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either a protein of the invention or a target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated protein of the invention or target molecules can be prepared from biotin-NHS(N-hydroxy-succinimide) using techniques well known in the art (*e.g.*, biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with a protein of the invention or target molecules, but which do not interfere

with binding of the protein to its target molecule, can be derivatized to the wells of the plate, and unbound target or protein trapped in the wells by antibody conjugation.

Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using

5 antibodies reactive with the protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the protein or target molecule.

In another embodiment, modulators of expression of nucleic acid molecules of the invention are identified in a method wherein a cell is contacted with a candidate
10 compound and the expression of appropriate mRNA or protein in the cell is determined. The level of expression of appropriate mRNA or protein in the presence of the candidate compound is compared to the level of expression of mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of expression based on this comparison. For example,
15 when expression of mRNA or protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator or enhancer of the mRNA or protein expression. Alternatively, when expression of the mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the
20 candidate compound is identified as an inhibitor of the mRNA or protein expression. The level of mRNA or protein expression in the cells can be determined by methods described herein for detecting mRNA or protein.

In yet another aspect of the invention, the proteins of the invention can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, *e.g.*, U.S. Patent
25 No. 5,283,317; Zervos *et al.* (1993) *Cell* 72:223-232; Madura *et al.* (1993) *J. Biol. Chem.* 268:12046-12054; Bartel *et al.* (1993) *Biotechniques* 14:920-924; Iwabuchi *et al.* (1993) *Oncogene* 8:1693-1696; and Brent WO94/10300), to identify other proteins (captured proteins) which bind to or interact with the proteins of the invention and modulate their activity. Such captured proteins are also likely to be involved in the
30 propagation of signals by the proteins of the invention as, for example, downstream elements of a protein-mediated signaling pathway. Alternatively, such captured proteins are likely to be cell-surface molecules associated with non-protein-expressing cells, wherein such captured proteins are involved in signal transduction.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (*e.g.*, a
5 modulating agent, an antisense nucleic acid molecule, a specific antibody, or a protein-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of
10 novel agents identified by the above-described screening assays for treatments as described herein.

IX. Methods of Treatment

The present invention provides for both prophylactic and therapeutic methods
15 of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant expression or activity of or related to proteins or nucleic acids of the invention. Methods of treatment involve modulating nucleic acid or polypeptide level or activity in a subject having a disorder that can be treated by such modulation. Accordingly, modulation can cause up regulation or down regulation of
20 the levels of expression or up regulation or down regulation of the activity of the nucleic acid or protein. Disorders relating to programmed cell death are particularly relevant as discussed in detail herein below.

Expression of the nucleic acids of the invention has been shown for the following tissues: testes, brain, heart, kidney, skeletal muscle, spleen, lung, smooth
25 muscle, pancreas, and liver as shown in Figure 8. Accordingly, disorders to which the methods disclosed herein are particularly relevant include those involving these tissues.

Disorders involving the spleen include, but are not limited to, splenomegaly, including nonspecific acute splenitis, congestive splenomegaly, and splenic infarcts;
30 neoplasms, congenital anomalies, and rupture. Disorders associated with splenomegaly include infections, such as nonspecific splenitis, infectious mononucleosis, tuberculosis, typhoid fever, brucellosis, cytomegalovirus, syphilis, malaria, histoplasmosis, toxoplasmosis, kala-azar, trypanosomiasis, schistosomiasis,

leishmaniasis, and echinococcosis; congestive states related to partial hypertension, such as cirrhosis of the liver, portal or splenic vein thrombosis, and cardiac failure; lymphohematogenous disorders, such as Hodgkin disease, non-Hodgkin lymphomas/leukemia, multiple myeloma, myeloproliferative disorders, hemolytic
5 anemias, and thrombocytopenic purpura; immunologic-inflammatory conditions, such as rheumatoid arthritis and systemic lupus erythematosus; storage diseases such as Gaucher disease, Niemann-Pick disease, and mucopolysaccharidoses; and other conditions, such as amyloidosis, primary neoplasms and cysts, and secondary neoplasms.

- 10 Disorders involving the lung include, but are not limited to, congenital anomalies; atelectasis; diseases of vascular origin, such as pulmonary congestion and edema, including hemodynamic pulmonary edema and edema caused by microvascular injury, adult respiratory distress syndrome (diffuse alveolar damage), pulmonary embolism, hemorrhage, and infarction, and pulmonary hypertension and
15 vascular sclerosis; chronic obstructive pulmonary disease, such as emphysema, chronic bronchitis, bronchial asthma, and bronchiectasis; diffuse interstitial (infiltrative, restrictive) diseases, such as pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia (pulmonary infiltration with eosinophilia),
20 *Bronchiolitis obliterans*-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, including Goodpasture syndrome, idiopathic pulmonary hemosiderosis and other hemorrhagic syndromes, pulmonary involvement in collagen vascular disorders, and pulmonary alveolar proteinosis; complications of therapies, such as drug-induced lung disease, radiation-induced lung disease, and lung transplantation;
25 tumors, such as bronchogenic carcinoma, including paraneoplastic syndromes, bronchioloalveolar carcinoma, neuroendocrine tumors, such as bronchial carcinoid, miscellaneous tumors, and metastatic tumors; pathologies of the pleura, including inflammatory pleural effusions, noninflammatory pleural effusions, pneumothorax, and pleural tumors, including solitary fibrous tumors (pleural fibroma) and malignant
30 mesothelioma.

Disorders involving the liver include, but are not limited to, hepatic injury; jaundice and cholestasis, such as bilirubin and bile formation; hepatic failure and cirrhosis, such as cirrhosis, portal hypertension, including ascites, portosystemic

shunts, and splenomegaly; infectious disorders, such as viral hepatitis, including hepatitis A-E infection and infection by other hepatitis viruses, clinicopathologic syndromes, such as the carrier state, asymptomatic infection, acute viral hepatitis, chronic viral hepatitis, and fulminant hepatitis; autoimmune hepatitis; drug- and toxin-induced liver disease, such as alcoholic liver disease; inborn errors of metabolism and pediatric liver disease, such as hemochromatosis, Wilson disease, α_1 -antitrypsin deficiency, and neonatal hepatitis; intrahepatic biliary tract disease, such as secondary biliary cirrhosis, primary biliary cirrhosis, primary sclerosing cholangitis, and anomalies of the biliary tree; circulatory disorders, such as impaired blood flow into the liver, including hepatic artery compromise and portal vein obstruction and thrombosis, impaired blood flow through the liver, including passive congestion and centrilobular necrosis and peliosis hepatis, hepatic vein outflow obstruction, including hepatic vein thrombosis (Budd-Chiari syndrome) and veno-occlusive disease; hepatic disease associated with pregnancy, such as preeclampsia and eclampsia, acute fatty liver of pregnancy, and intrahepatic cholestasis of pregnancy; hepatic complications of organ or bone marrow transplantation, such as drug toxicity after bone marrow transplantation, graft-versus-host disease and liver rejection, and nonimmunologic damage to liver allografts; tumors and tumorous conditions, such as nodular hyperplasias, adenomas, and malignant tumors, including primary carcinoma of the liver metastatic tumors, and liver fibrosis.

Disorders involving the brain include, but are not limited to, disorders involving neurons, and disorders involving glia, such as astrocytes, oligodendrocytes, ependymal cells, and microglia; cerebral edema, raised intracranial pressure and herniation, and hydrocephalus; malformations and developmental diseases, such as neural tube defects, forebrain anomalies, posterior fossa anomalies, and syringomyelia and hydromyelia; perinatal brain injury; cerebrovascular diseases, such as those related to hypoxia, ischemia, and infarction, including hypotension, hypoperfusion, and low-flow states--global cerebral ischemia and focal cerebral ischemia--infarction from obstruction of local blood supply, intracranial hemorrhage, including intracerebral (intraparenchymal) hemorrhage, subarachnoid hemorrhage and ruptured berry aneurysms, and vascular malformations, hypertensive cerebrovascular disease, including lacunar infarcts, slit hemorrhages, and hypertensive encephalopathy; infections, such as acute meningitis, including acute pyogenic (bacterial) meningitis

and acute aseptic (viral) meningitis, acute focal suppurative infections, including brain abscess, subdural empyema, and extradural abscess, chronic bacterial meningoencephalitis, including tuberculosis and mycobacterioses, neurosyphilis, and neuroborreliosis (Lyme disease), viral meningoencephalitis, including arthropod-

5 borne (Arbo) viral encephalitis, *Herpes simplex* virus Type 1, *Herpes simplex* virus Type 2, *Varicella-zoster* virus (*Herpes zoster*), cytomegalovirus, poliomyelitis, rabies, and human immunodeficiency virus 1, including HIV-1 meningoencephalitis (subacute encephalitis), vacuolar myelopathy, AIDS-associated myopathy, peripheral neuropathy, and AIDS in children, progressive multifocal leukoencephalopathy,

10 subacute sclerosing panencephalitis, fungal meningoencephalitis, other infectious diseases of the nervous system; transmissible spongiform encephalopathies (prion diseases); demyelinating diseases, including multiple sclerosis, multiple sclerosis variants, acute disseminated encephalomyelitis and acute necrotizing hemorrhagic encephalomyelitis, and other diseases with demyelination; degenerative diseases, such

15 as degenerative diseases affecting the cerebral cortex, including Alzheimer disease and Pick disease, degenerative diseases of basal ganglia and brain stem, including Parkinsonism, idiopathic Parkinson disease (paralysis agitans), progressive supranuclear palsy, corticobasal degeneration, multiple system atrophy, including striatonigral degeneration, Shy-Drager syndrome, and olivopontocerebellar atrophy,

20 and Huntington disease; spinocerebellar degenerations, including spinocerebellar ataxias, including Friedreich ataxia, and ataxia-telanglectasia, degenerative diseases affecting motor neurons, including amyotrophic lateral sclerosis (motor neuron disease), bulbospinal atrophy (Kennedy syndrome), and spinal muscular atrophy; inborn errors of metabolism, such as leukodystrophies, including Krabbe disease,

25 metachromatic leukodystrophy, adrenoleukodystrophy, Pelizaeus-Merzbacher disease, and Canavan disease, mitochondrial encephalomyopathies, including Leigh disease and other mitochondrial encephalomyopathies; toxic and acquired metabolic diseases, including vitamin deficiencies such as thiamine (vitamin B₁) deficiency and vitamin B₁₂ deficiency, neurologic sequelae of metabolic disturbances, including

30 hypoglycemia, hyperglycemia, and hepatic encephatopathy, toxic disorders, including carbon monoxide, methanol, ethanol, and radiation, including combined methotrexate and radiation-induced injury; tumors, such as gliomas, including astrocytoma, including fibrillary (diffuse) astrocytoma and glioblastoma multiforme, pilocytic

astrocytoma, pleomorphic xanthoastrocytoma, and brain stem glioma, oligodendroglioma, and ependymoma and related paraventricular mass lesions, neuronal tumors, poorly differentiated neoplasms, including medulloblastoma, other parenchymal tumors, including primary brain lymphoma, germ cell tumors, and pineal parenchymal tumors, meningiomas, metastatic tumors, paraneoplastic syndromes, peripheral nerve sheath tumors, including schwannoma, neurofibroma, and malignant peripheral nerve sheath tumor (malignant schwannoma), and neurocutaneous syndromes (phakomatoses), including neurofibromatosis, including Type 1 neurofibromatosis (NF1) and TYPE 2 neurofibromatosis (NF2), tuberous sclerosis, and Von Hippel-Lindau disease.

Disorders involving the heart, include but are not limited to, heart failure, including but not limited to, cardiac hypertrophy, left-sided heart failure, and right-sided heart failure; ischemic heart disease, including but not limited to angina pectoris, myocardial infarction, chronic ischemic heart disease, and sudden cardiac death; hypertensive heart disease, including but not limited to, systemic (left-sided) hypertensive heart disease and pulmonary (right-sided) hypertensive heart disease; valvular heart disease, including but not limited to, valvular degeneration caused by calcification, such as calcific aortic stenosis, calcification of a congenitally bicuspid aortic valve, and mitral annular calcification, and myxomatous degeneration of the mitral valve (mitral valve prolapse), rheumatic fever and rheumatic heart disease, infective endocarditis, and noninfected vegetations, such as nonbacterial thrombotic endocarditis and endocarditis of systemic lupus erythematosus (Libman-Sacks disease), carcinoid heart disease, and complications of artificial valves; myocardial disease, including but not limited to dilated cardiomyopathy, hypertrophic cardiomyopathy, restrictive cardiomyopathy, and myocarditis; pericardial disease, including but not limited to, pericardial effusion and hemopericardium and pericarditis, including acute pericarditis and healed pericarditis, and rheumatoid heart disease; neoplastic heart disease, including but not limited to, primary cardiac tumors, such as myxoma, lipoma, papillary fibroelastoma, rhabdomyoma, and sarcoma, and cardiac effects of noncardiac neoplasms; congenital heart disease, including but not limited to, left-to-right shunts--late cyanosis, such as atrial septal defect, ventricular septal defect, patent ductus arteriosus, and atrioventricular septal defect, right-to-left shunts--early cyanosis, such as tetralogy of fallot, transposition of great arteries,

truncus arteriosus, tricuspid atresia, and total anomalous pulmonary venous connection, obstructive congenital anomalies, such as coarctation of aorta, pulmonary stenosis and atresia, and aortic stenosis and atresia, and disorders involving cardiac transplantation.

- 5 Disorders involving the kidney include, but are not limited to, congenital anomalies including, but not limited to, cystic diseases of the kidney, that include but are not limited to, cystic renal dysplasia, autosomal dominant (adult) polycystic kidney disease, autosomal recessive (childhood) polycystic kidney disease, and cystic diseases of renal medulla, which include, but are not limited to, medullary sponge kidney, and
- 10 nephronophthisis-uremic medullary cystic disease complex, acquired (dialysis-associated) cystic disease, such as simple cysts; glomerular diseases including pathologies of glomerular injury that include, but are not limited to, in situ immune complex deposition, that includes, but is not limited to, anti-GBM nephritis, Heymann nephritis, and antibodies against planted antigens, circulating immune complex nephritis,
- 15 antibodies to glomerular cells, cell-mediated immunity in glomerulonephritis, activation of alternative complement pathway, epithelial cell injury, and pathologies involving mediators of glomerular injury including cellular and soluble mediators, acute glomerulonephritis, such as acute proliferative (poststreptococcal, postinfectious) glomerulonephritis, including but not limited to, poststreptococcal glomerulonephritis
- 20 and nonstreptococcal acute glomerulonephritis, rapidly progressive (crescentic) glomerulonephritis, nephrotic syndrome, membranous glomerulonephritis (membranous nephropathy), minimal change disease (lipoid nephrosis), focal segmental glomerulosclerosis, membranoproliferative glomerulonephritis, IgA nephropathy (Berger disease), focal proliferative and necrotizing glomerulonephritis (focal
- 25 glomerulonephritis), hereditary nephritis, including but not limited to, Alport syndrome and thin membrane disease (benign familial hematuria), chronic glomerulonephritis, glomerular lesions associated with systemic disease, including but not limited to, systemic lupus erythematosus, Henoch-Schönlein purpura, bacterial endocarditis, diabetic glomerulosclerosis, amyloidosis, fibrillary and immunotactoid
- 30 glomerulonephritis, and other systemic disorders; diseases affecting tubules and interstitium, including acute tubular necrosis and tubulointerstitial nephritis, including but not limited to, pyelonephritis and urinary tract infection, acute pyelonephritis, chronic pyelonephritis and reflux nephropathy, and tubulointerstitial nephritis induced

by drugs and toxins, including but not limited to, acute drug-induced interstitial nephritis, analgesic abuse nephropathy, nephropathy associated with nonsteroidal anti-inflammatory drugs, and other tubulointerstitial diseases including, but not limited to, urate nephropathy, hypercalcemia and nephrocalcinosis, and multiple myeloma; diseases
5 of blood vessels including benign nephrosclerosis, malignant hypertension and accelerated nephrosclerosis, renal artery stenosis, and thrombotic microangiopathies including, but not limited to, classic (childhood) hemolytic-uremic syndrome, adult hemolytic-uremic syndrome/thrombotic thrombocytopenic purpura, idiopathic HUS/TTP, and other vascular disorders including, but not limited to, atherosclerotic
10 ischemic renal disease, atheroembolic renal disease, sickle cell disease nephropathy, diffuse cortical necrosis, and renal infarcts; urinary tract obstruction (obstructive uropathy); urolithiasis (renal calculi, stones); and tumors of the kidney including, but not limited to, benign tumors, such as renal papillary adenoma, renal fibroma or hamartoma (renomedullary interstitial cell tumor), angiomyolipoma, and oncocytoma, and malignant
15 tumors, including renal cell carcinoma (hypernephroma, adenocarcinoma of kidney), which includes urothelial carcinomas of renal pelvis.

Disorders involving the testis and epididymis include, but are not limited to, congenital anomalies such as cryptorchidism, regressive changes such as atrophy, inflammations such as nonspecific epididymitis and orchitis, granulomatous
20 (autoimmune) orchitis, and specific inflammations including, but not limited to, gonorrhea, mumps, tuberculosis, and syphilis, vascular disturbances including torsion, testicular tumors including germ cell tumors that include, but are not limited to, seminoma, spermatocytic seminoma, embryonal carcinoma, yolk sac tumor choriocarcinoma, teratoma, and mixed tumors, tumors of sex cord-gonadal stroma
25 including, but not limited to, leydig (interstitial) cell tumors and sertoli cell tumors (androblastoma), and testicular lymphoma, and miscellaneous lesions of tunica vaginalis.

Disorders involving the skeletal muscle include tumors such as rhabdomyosarcoma.

Disorders involving the pancreas include those of the exocrine pancreas such as
30 congenital anomalies, including but not limited to, ectopic pancreas; pancreatitis, including but not limited to, acute pancreatitis; cysts, including but not limited to, pseudocysts; tumors, including but not limited to, cystic tumors and carcinoma of the pancreas; and disorders of the endocrine pancreas such as, diabetes mellitus; islet cell

tumors, including but not limited to, insulinomas, gastrinomas, and other rare islet cell tumors.

Preferred disorders include those involving the central nervous system and particularly the brain.

- 5 With regard to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics. "Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market.
- 10 More specifically, the term refers the study of how a patient's genes determine his or her response to a drug (*e.g.*, a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with the molecules of the present invention or modulators according to that individual's drug response genotype.
- 15 Pharmacogenomics allows a clinician or physician to target prophylactic or therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug related side effects.

1. Prophylactic Methods

- 20 In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with aberrant expression or activity of genes or proteins of the present invention, by administering to the subject an agent which modulates expression or at least one activity of a gene or protein of the invention. Subjects at risk for a disease that is caused or contributed to by aberrant gene
- 25 expression or protein activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of aberrancy, for example, an agonist or
- 30 antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating expression or activity of genes or proteins of the invention for therapeutic purposes. The modulatory method of the invention involves contacting a cell with an agent that modulates one or more of the activities of the specified protein associated with the cell. An agent that modulates protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of a protein described herein, a polypeptide, a peptidomimetic, or other small molecule. In one embodiment, the agent stimulates one or more protein activities. Examples of such stimulatory agents include active protein as well as a nucleic acid molecule encoding the protein that has been introduced into the cell. In another embodiment, the agent inhibits one or more protein activities. Examples of such inhibitory agents include antisense nucleic acid molecules and anti-protein antibodies. These modulatory methods can be performed *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of a protein or nucleic acid molecule of the invention. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) expression or activity of a gene or protein of the invention. In another embodiment, the method involves administering a protein or nucleic acid molecule of the invention as therapy to compensate for reduced or aberrant expression or activity of the protein or nucleic acid molecule.

Stimulation of protein activity is desirable in situations in which the protein is abnormally downregulated and/or in which increased protein activity is likely to have a beneficial effect. Likewise, inhibition of protein activity is desirable in situations in which the protein is abnormally upregulated and/or in which decreased protein activity is likely to have a beneficial effect. One example of such a situation is where a subject has a disorder characterized by aberrant development or cellular differentiation. Another example of such a situation is where the subject has a proliferative disease (e.g., cancer) or a disorder characterized by an aberrant

hematopoietic response. Yet another example of such a situation is where it is desirable to achieve tissue regeneration in a subject (*e.g.*, where a subject has undergone brain or spinal cord injury and it is desirable to regenerate neuronal tissue in a regulated manner).

5

Pharmaceutical Compositions

The nucleic acid molecules, protein modulators of the protein, and antibodies (also referred to herein as "active compounds") can be incorporated into pharmaceutical compositions suitable for administration to a subject, *e.g.*, a human. Such compositions typically comprise the nucleic acid molecule, protein, modulator, or antibody and a pharmaceutically acceptable carrier.

The term "administer" is used in its broadest sense and includes any method of introducing the compositions of the present invention into a subject. This includes producing polypeptides or polynucleotides *in vivo* as by transcription or translation, *in vivo*, of polynucleotides that have been exogenously introduced into a subject. Thus, polypeptides or nucleic acids produced in the subject from the exogenous compositions are encompassed in the term "administer."

As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, such media can be used in the compositions of the invention. Supplementary active compounds can also be incorporated into the compositions. A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as

ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampules, disposable syringes or multiple dose
5 vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water,
10 Cremophor EL™ (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for
15 example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and
20 antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example,
25 aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a ubiquitin protease protein or anti- ubiquitin protease antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions
30 are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields

a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For oral administration, the agent can be contained in enteric forms to survive the stomach or further coated or mixed to be released in a particular region of the GI tract by known methods. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser, which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. "Dosage unit form" as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (U.S. 5,328,470) or by stereotactic injection (see e.g., Chen *et al.* (1994) *PNAS* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, e.g. retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

As defined herein, a therapeutically effective amount of protein or polypeptide (i.e., an effective dosage) ranges from about 0.001 to 30 mg/kg body weight,

preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight.

The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments. In a preferred example, a subject is treated with antibody, protein, or polypeptide in the range of between about 0.1 to 20 mg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of antibody, protein, or polypeptide used for treatment may increase or decrease over the course of a particular treatment. Changes in dosage may result and become apparent from the results of diagnostic assays as described herein.

The present invention encompasses agents which modulate expression or activity. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (i.e., including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds.

It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon the identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if

applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention. Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein. When one or more of these small molecules is to be administered to an animal (e.g., a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, gender, and diet of the subject, the time of administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

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3. Pharmacogenomics

The molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on the protein activity (e.g., gene expression) as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) disorders (e.g., proliferative or developmental disorders) associated with aberrant protein activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer a molecule of the invention or modulator thereof,

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as well as tailoring the dosage and/or therapeutic regimen of treatment with such a molecule or modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See *e.g.*, Eichelbaum (1996) *Clin Exp. Pharmacol. Physiol.* 23(10-11):983-985 and Linder (1997) *Clin. Chem.* 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (*e.g.*, a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants). Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a "SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1,000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict drug response. According to this method, if a gene that encodes a drug's target is known (*e.g.*, a protein or a polypeptide of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (*e.g.*, N-acetyltransferase 2(NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme is the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (*e.g.*, a molecule or modulator of the present invention) can given an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with a

molecule or modulator of the invention, such as a modulator identified by one of the exemplary screening assays described herein.

Disorders which may be treated or diagnosed by methods described herein include, but are not limited to disorders involving apoptosis. Certain disorders are associated with an increased number of surviving cells, which are produced and continue to survive or proliferate when apoptosis is inhibited.

As used herein, "programmed cell death" refers to a genetically regulated process involved in the normal development of multicellular organisms. This process occurs in cells destined for removal in a variety of normal situations, including larval development of the nematode *C. elegans*, insect metamorphosis, development in mammalian embryos, including the nephrogenic zone in the developing kidney, and regression or atrophy (e.g., in the prostate after castration). Programmed cell death can occur following the withdrawal of growth and trophic factors in many cells, nutritional deprivation, hormone treatment, ultraviolet irradiation, and exposure to toxic and infectious agents including reactive oxygen species and phosphatase inhibitors, e.g., okadaic acid, calcium ionophores, and a number of cancer chemotherapeutic agents. See Wilson (1998) *Biochem. Cell Biol.* 76:573-582 and Hetts (1998) *JAMA* 279:300-307, the contents of which are incorporated herein by reference. Thus, the proteins of the invention, by being differentially expressed during programmed cell death, e.g., neuronal programmed cell death, can modulate a programmed cell death pathway activity and provide novel diagnostic targets and therapeutic agents for disorders characterized by deregulated programmed cell death, particularly in cells that express the protein.

As used herein, a "disorder characterized by deregulated programmed cell death" refers to a disorder, disease or condition which is characterized by a deregulation, e.g., an upregulation or a downregulation, of programmed cell death. Programmed cell death deregulation can lead to deregulation of cellular proliferation and/or cell cycle progression. Examples of disorders characterized by deregulated programmed cell death include, but are not limited to, neurodegenerative disorders, e.g., Alzheimer's disease, dementias related to Alzheimer's disease (such as Pick's disease), Parkinson's and other Lewy diffuse body diseases, multiple sclerosis, amyotrophic lateral sclerosis, progressive supranuclear palsy, epilepsy, Jakob-Creutzfeldt disease, or AIDS related dementias; myelodysplastic syndromes, e.g.,

aplastic anemia; ischemic injury, e.g., myocardial infarction, stroke, or reperfusion injury; autoimmune disorders, e.g., systemic lupus erythematosus, or immune-mediated glomerulonephritis; or proliferative disorders, e.g., cancer, such as follicular lymphomas, carcinomas with p53 mutations, or hormone-dependent tumors, e.g., breast cancer, prostate cancer, or ovarian cancer). Clinical manifestations of faulty apoptosis are also seen in stroke and in rheumatoid arthritis. Wilson (1998) *Biochem. Cell. Biol.* 76:573-582.

Failure to remove autoimmune cells that arise during development or that develop as a result of somatic mutation during an immune response can result in autoimmune disease. One of the molecules that plays a critical role in regulating cell death in lymphocytes is the cell surface receptor for Fas.

Viral infections, such as those caused by herpesviruses, poxviruses, and adenoviruses, may result in aberrant apoptosis. Populations of cells are often depleted in the event of viral infection, with perhaps the most dramatic example being the cell depletion caused by the human immunodeficiency virus (HIV). Most T cells that die during HIV infections do not appear to be infected with HIV. Stimulation of the CD4 receptor may result in the enhanced susceptibility of uninfected T cells to undergo apoptosis.

Many disorders can be classified based on whether they are associated with abnormally high or abnormally low apoptosis. Thompson (1995) *Science* 267:1456-1462. Apoptosis may be involved in acute trauma, myocardial infarction, stroke, and infectious diseases, such as viral hepatitis and acquired immunodeficiency syndrome.

Primary apoptosis deficiencies include graft rejection. Accordingly, the invention is relevant to the identification of genes useful in inhibiting graft rejection.

Primary apoptosis deficiencies also include autoimmune diabetes. Accordingly, the invention is relevant to the identification of genes involved in autoimmune diabetes and accordingly, to the identification of agents that act on these targets to modulate the expression of these genes and hence, to treat or diagnose this disorder. Further, it has been suggested that all autoimmune disorders can be viewed as primary deficiencies of apoptosis (Hetts, above). Accordingly, the invention is relevant for screening for gene expression and transcriptional profiling in any autoimmune disorder and for screening for agents that affect the expression or transcriptional profile of these genes.

Primary apoptosis deficiencies also include local self reactive disorder. This includes Hashimoto thyroiditis.

Primary apoptosis deficiencies also include lymphoproliferation and autoimmunity. This includes, but is not limited to, Canale-Smith syndrome.

- 5 Primary apoptosis deficiencies also include cancer. For example, p53 induces apoptosis by acting as a transcription factor that activates expression of various apoptosis-mediating genes or by upregulating apoptosis-mediating genes such as Bax.

- 10 Primary apoptosis excesses are associated with neurodegenerative disorders including Alzheimer's disease, Parkinson's disease, spinal muscular atrophy, and amyotrophic lateral sclerosis.

Primary apoptosis excesses are also associated with heart disease including idiopathic dilated cardiomyopathy, ischemic cardiomyopathy, and valvular heart disease. Evidence has also been shown of apoptosis in heart failure resulting from arrhythmogenic right ventricular dysplasia. For all these disorders, see Hetts, above.

- 15 Death receptors also include the TNF receptor-I and hence, TNF acts as a death ligand.

- 20 A wide variety of neurological diseases are characterized by the gradual loss of specific sets of neurons. Such disorders include Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis (ALS) retinitis pigmentosa, spinal muscular atrophy, and various forms of cerebellar degeneration. The cell loss in these diseases does not induce an inflammatory response, and apoptosis appears to be the mechanism of cell death.

- 25 In addition, a number of hematologic diseases are associated with a decreased production of blood cells. These disorders include anemia associated with chronic disease, aplastic anemia, chronic neutropenia, and the myelodysplastic syndromes. Disorders of blood cell production, such as myelodysplastic syndrome and some forms of aplastic anemia, are associated with increased apoptotic cell death within the bone marrow.

- 30 These disorders could result from the activation of genes that promote apoptosis, acquired deficiencies in stromal cells or hematopoietic survival factors, or the direct effects of toxins and mediators of immune responses.

Two common disorders associated with cell death are myocardial infarctions and stroke. In both disorders, cells within the central area of ischemia, which is

produced in the event of acute loss of blood flow, appear to die rapidly as a result of necrosis. However, outside the central ischemic zone, cells die over a more protracted time period and morphologically appear to die by apoptosis.

The invention also pertains to disorders of the central nervous system (CNS).

5 These disorders include, but are not limited to cognitive and neurodegenerative disorders such as Alzheimer's disease, senile dementia, Huntington's disease, amyotrophic lateral sclerosis, and Parkinson's disease, as well as Gilles de la Tourette's syndrome, autonomic function disorders such as hypertension and sleep disorders, and neuropsychiatric disorders that include, but are not limited to
10 schizophrenia, schizoaffective disorder, attention deficit disorder, dysthymic disorder, major depressive disorder, mania, obsessive-compulsive disorder, psychoactive substance use disorders, anxiety, panic disorder, as well as bipolar affective disorder, e.g., severe bipolar affective (mood) disorder (BP-I), bipolar affective (mood) disorder with hypomania and major depression (BP-II). Further CNS-related
15 disorders include, for example, those listed in the American Psychiatric Association's Diagnostic and Statistical manual of Mental Disorders (DSM), the most current version of which is incorporated herein by reference in its entirety.

As used herein, "differential expression" or differentially expressed" includes both quantative and qualitative differences in the temporal and/or cellular expression
20 pattern of a gene, e.g., the programmed cell death genes disclosed herein, among, for example, normal cells and cells undergoing programmed cell death. Genes which are differentially expressed can be used as part of a prognostic or diagnostic marker for the evaluation of subjects at risk for developing a disorder characterized by deregulated programmed cell death. Depending on the expression level of the gene,
25 the progression state of the disorder can also be evaluated.

X. Arrays and Microarrays

The term "array" refers to a set of nucleic acid sequences that comprise at least one of SEQ ID NOS: 1-6, 8, and 10. Preferred arrays contain numerous genes. The
30 term can refer to all of the sequences in SEQ ID NOS: 1-6, 8, and 10 but could also include additional sequences, for example, sequences included as controls for specific biological processes. A "subarray" is also an array but is obtained by creating an array of less than all of the sequences in a starting array.

In one embodiment of the invention, the functional subarray comprises nucleic acid sequences expressed in programmed cell death as disclosed herein.

The array comprises not only the specific designated sequences but also variants of these sequences, as described herein. As described, variants include,
5 allelic variants, homologs from other loci in the same animal, orthologs, and sequences sufficiently similar such that they fulfill the requisites for sequence similarity/homology as described herein.

Further, the array not only comprises the specific designated sequences, but also comprises fragments thereof. As described herein, the range of fragments will
10 vary depending upon the specific sequence involved. Accordingly, the range of fragments is considerable, for example, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 125, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 etc. In no way, however, is a fragment to be construed as having a sequence identical to that which may be found in the prior art.

15 The array can be used to assay expression of one or more genes in the array.

In one embodiment, the array can be used to assay gene expression in a tissue to ascertain tissue specificity of genes in the array.

In addition to such qualitative determination, the invention allows the quantitation of gene expression. Thus, not only tissue specificity, but also the level of
20 expression of a battery of genes in the tissue is ascertainable. Thus, genes can be grouped on the basis of their tissue expression *per se* and level of expression in that tissue. This is useful, for example, in ascertaining the relationship of gene expression between or among tissues. Thus, one tissue can be perturbed and the effect on gene expression in a second tissue can be determined. In this context, the effect of one cell
25 type on another cell type in response to a biological stimulus can be determined. Such a determination is useful, for example, to know the effect of cell-cell interaction at the level of gene expression. If an agent is administered therapeutically to treat one cell type but has an undesirable effect on another cell type, the invention provides an assay to determine the molecular basis of the undesirable effect and thus provides the
30 opportunity to co-administer a counteracting agent or otherwise treat the undesired effect. Similarly, even within a single cell type, undesirable biological effects can be determined at the molecular level. Thus, the effects of an agent on expression of other than the target gene can be ascertained and counteracted.

In another embodiment, the array can be used to monitor the time course of expression of one or more genes in the array. This can occur in various biological contexts, as disclosed herein, for example development and differentiation, tumor progression, progression of other diseases, *in vitro* processes, such as cellular transformation and senescence, autonomic neural and neurological processes, such as, for example, pain and appetite, and cognitive functions, such as learning or memory.

The array is also useful for ascertaining the effect of the expression of a gene on the expression of other genes in the same cell or in different cells. This provides, for example, for a selection of alternate molecular targets for therapeutic intervention if the ultimate or downstream target cannot be regulated.

The array is also useful for ascertaining differential expression patterns of one or more genes in normal and abnormal cells. This provides a battery of genes that could serve as a molecular target for diagnosis or therapeutic intervention.

In one embodiment, the array, and particularly subarrays containing one or more of the nucleic acid sequences related to programmed cell death, are useful for diagnosing disease or predisposition to disease involving apoptosis. These disorders include, but are not limited to, those discussed in detail herein. In addition, the array or subarrays created therefrom are useful for diagnosing active disorders of the central nervous system or for predicting the tenancy to develop such disorders. Disorders of the central nervous system include, but are not limited to, those disclosed in detail herein. Furthermore, the array and subarrays thereof are useful for diagnosing an active disorder or predicting the tendency to develop a disorder including, but not limited to, disorders involving secretion/synaptic vesicle release, cell proliferation, cytoskeletal reorganization, stress response/hormone response; and calcium signal transduction.

The array is also useful for ascertaining expression of one or more genes in model systems *in vitro* or *in vivo*. Various model systems have been developed to study normal and abnormal processes, including, but not limited to, apoptosis.

Apoptosis can be actively induced in animal cells by a diverse array of triggers that range from ionizing radiation to hypothermia to viral infections to immune reactions. Majno *et al.* (1995) *Amer. J. Pathol.* 146:3-15; Hockenberry *et al.* (1995) *Bio Essays* 17:631-638; Thompson *et al.* *Science* 267:1456-1462 (1995).

Transgenic mouse models have been developed for familial amyotrophic lateral sclerosis, familial Alzheimer's disease and Huntington's disease, reviewed in Price *et al.* (1998) *Science* 282:1079-1083. Amyotrophic lateral sclerosis is the most common adult onset motor neuron disease. Alzheimer's disease is the most common
5 cause of dementia in adult life. It is associated with the damage of regions and neurocircuits critical for cognition and memory, including neurons in the neocortex, hippocampus, amygdala, basal forebrain cholinergic system, and brain stem monoaminergic nuclei. Neurological diseases that are associated with autosomal dominant trinucleotide repeat mutations include Huntington's disease, several spinal
10 cerebellar ataxias and dentatorubral pallidoluysian atrophy. SCA-1 and SCA-3 or Machado-Joseph disease are characterized by ataxia and lack of coordination. In Huntington's disease, symptoms are related to degeneration of subsets of striatal and cortical neurons. Apoptosis is thought to play a role in the degeneration of these cells. In SCA-1, SCA-3, and in dentatorubral pallidoluysian atrophy, a variety of cell
15 populations, and particularly cells in the cerebellum, have been shown to degenerate. See Price *et al.* above, which is incorporated by reference in its entirety for the teachings of model systems related to neurodegenerative diseases.

Mouse models have been developed for non-obese diabetic mice, to study disease progression for the treatment of autoimmune diabetes mellitus. Bellgrau *et al.*
20 (1995) *Nature* 377:630-632. Models have also been developed in mice wherein the mice lack one or two copies of the p53 gene. Study of these mice has shown that apoptosis is involved in suppressing tumor development in vivo. Lozano *et al.* (1998) *Semin. Canc. Biol.* 8:337-344. Another animal model relevant to the study of apoptosis involves the targeted gene disruption of caspase genes creating caspase
25 gene knockout mice. Colussi *et al.* (1999) *J. Immun. Cell. Biol.* 77:58-63. A further mouse model pertains to cold injury in mice, such injury inducing neuronal apoptosis. Murakami *et al.* (1999) *Prog. Neurobiol.* 57:289-299.

Knockout mice have been created for Apaf1. In these mice, defects are found in essentially all tissues whose development depends on cell death, including loss of
30 interdigital webs, formation of the palate, control of neuron cell number, and development of the lens and retina. Cecconi *et al.* (1998) *Cell* 94:727-737.

Caspase knockout mice have also been achieved for caspase 1, 2, 3, and 9. Green (1998) *Cell* 94:695-698.

The array allows the simultaneous determination of a battery of genes involved in these processes and thus provides multiple candidates for *in vivo* verification and clinical testing. Because the array allows the determination of expression of multiple genes, it provides a powerful tool to ascertain coordinate gene expression, that is co-expression of two or more genes in a time and/or tissue-specific manner, both qualitatively and quantitatively. Thus, genes can be grouped on the basis of their expression *per se* and/or level of expression. This allows the classification of genes into functional categories even when the gene is completely uncharacterized with respect to function. Accordingly, if a first gene is expressed coordinately with a second gene whose function is known, a putative function can be assigned to that first gene. This first gene thus provides a new target for affecting that function in a diagnostic or therapeutic context. The larger the number of genes in an array, the greater is the probability that numerous known genes having the same or similar function will be expressed. In this case, the coordinate expression of one or more novel genes (with respect to function and/or structure) strongly allows discovery of genes in the same functional category as the known genes.

Accordingly, the array of the invention provides for "internal control" groups of genes whose functions are known and can thus be used to identify genes as being in the same functional category of the control group if they are coordinated expressed.

As an alternative to relying on such internal control groups, external control groups can be added to the array. The genes in such a group would have a known function. Genes coordinately expressed with these genes would thus be *prima facie* involved in the same function.

Therefore, the array provides a method not only for discovering novel genes having a specific function but also for assigning function to genes whose function is unknown or assigning to a known gene an additional function, previously unknown for that gene.

Accordingly, as disclosed and exemplified herein, previously characterized genes were grouped into new functional categories (i.e., previously the function was not known to be possessed by that gene). Furthermore, several uncharacterized genes could be functionally classified on the basis of coordinate expression with the "internal control group of genes". In a specific embodiment, disclosed and exemplified herein, genes related to programmed cell death in brain were selected.

The array could, accordingly be used to select for genes related to other important biological processes, such as those disclosed herein. Nucleic acid from any tissue in any biological process is hybridized to nucleic acid sequences in an array. The expression pattern of genes in the array allows for their classification into functional groups based on specific expression patterns. Internal or external control genes (i.e. genes known to be expressed in the specific tissue/biological process) provide verification to classify other genes in the specific category.

Just as the array was useful for identifying programmed cell death genes, other relevant normal biological models include differentiation programs and disorders such as those disclosed herein.

The array is also useful for drug discovery. Candidate compounds can be used to screen cells and tissues in any of the biological contexts disclosed herein, such as pathology, development, differentiation, etc. Thus the expression of one or more genes in the array can be monitored by using the array to screen for RNA expression in a cell or tissue exposed to a candidate compound. Compounds can be selected on the basis of the overall effect on gene expression, not necessarily on the basis of its effect on a single gene. Thus, for example, where a compound is desired that affects a particular first gene or genes but has no effect on a second gene or genes, the array provides a way to globally monitor the effect on gene expression of a compound.

Alternatively, it may be desirable to target more than one gene, i.e. to modulate the expression of more than one gene. The array provides a way to discover compounds that will modulate a set of genes. All genes of the set can be upregulated or downregulated. Alternatively, some of the genes may be upregulated and others downregulated by the same compound. Moreover, compounds are discoverable that modulate desired genes to desired degrees.

In the context of drug discovery, functional subarrays of genes are especially useful. Thus, using the methods disclosed herein and those routinely available, groups of genes can be assembled based on their relationships to a specific biological function. The expression of this group of genes can be used for diagnostic purposes and to discover compounds relevant to the biological function. Thus, the subarray can provide the basis for discovering drugs relevant to treatment and diagnosis of disease, for example those disclosed herein.

In the present case, the group of genes whose expression is correlated with programmed cell death can be used to discover compounds that affect programmed cell death, and especially disorders in which programmed cell death is involved. These include but are not limited to those disclosed herein.

5 Apoptosis can be triggered by the addition of apoptosis-promoting ligands to a cell in culture or in vivo. In one embodiment of the invention, therefore, the arrays and subarrays described herein are useful to identify genes that respond to apoptosis-promoting ligands and conversely to identify ligands that act on genes involved in apoptosis. Apoptosis can also be triggered by decreasing or removing an apoptosis-
10 inhibiting or survival-promoting ligand. Accordingly, apoptosis is triggered in view of the fact that the cell lacks a signal from a cell surface survival factor receptor. Ligands include, but are not limited to, FasL. Death-inhibiting ligands include, but are not limited to, IL-2. See Hetts *et al.* (1998) *JAMA* 279:300-307 (incorporated by reference in its entirety for teaching of ligands involved in active and passive
15 apoptosis pathways.) Central in the pathway, and also serving as potential molecules for inducing (or releasing from inhibition) apoptosis pathways include FADD, caspases, human CED4 homolog (also called apoptotic protease activating factor 1), the Bcl-2 family of genes including, but not limited to, apoptosis promoting (for example, Bax and Bad) and apoptosis inhibiting (for example, Bcl-2 and Bcl-x_L)
20 molecules. See Hetts *et al.*, above.

Multiple caspases upstream of caspase-3 can be inhibited by viral proteins such as cowpox, CrmA, and baculovirus, p35, synthetic tripeptides and tetrapeptides inhibit caspase-3 specifically (Hetts, above). Accordingly, the arrays and subarrays are useful for determining the modulation of gene expression in response to these
25 agents.

The array is also useful for obtaining a set of human (or other animal) orthologs that can be used for drug discovery, treatment, diagnosis, and the other uses disclosed herein. The subarrays can be used to specifically create a corresponding human (or other animal) subarray that is relevant to a specific biological function.
30 Accordingly, a method is provided for obtaining sets of genes from other organisms, which sets are correlated with, for example, disease or developmental disorders.

In a preferred embodiment of the invention, the arrays and subarrays disclosed herein are in a "microarray". The term "microarray" is intended to designate an array

of nucleic acid sequences on a chip. This includes *in situ* synthesis of desired nucleic acid sequences directly on the chip material, or affixing previously chemically synthesized nucleic acid sequences or nucleic acid sequences produced by recombinant DNA methodology onto the chip material. In the case of recombinant DNA methodology, nucleic acids can include whole vectors containing desired inserts, such as phages and plasmids, the desired inserts removed from the vector as by, PCR cloning, cDNA synthesized from mRNA, mRNA modified to avoid degradation, and the like.

A series of state-of-the-art reviews of the technology for production of nucleic acid microarrays in various formats and examples of their utilization to address biological problems is provided in *Nature Genetics*, 21 Supplement, January 1999. These topics include molecular interactions on microarrays, expression profiling using cDNA microarrays, making and reading microarrays, high density synthetic oligonucleotide arrays, sequencing and mutation analysis using oligonucleotide microarrays, the use of microarrays in drug discovery and development, gene expression informatics, and use of arrays in population genetics. Various microarray substrates, methods for processing the substrates to affix the nucleic acids onto the substrates, processes for hybridization of the nucleic acid on the substrate to an external nucleic acid sample, methods for detection, and methods for analyzing expression data using specific algorithms have been widely disclosed in the art. References disclosing various microarray technologies are listed below.

Lashkari *et al.* (1997) "Yeast Microarrays for Genome Wide Parallel Genetic and Gene Expression Analysis", *Proc. Natl. Acad. Sci.* 94:13057-13062; Ramsay (1998) "DNA Chips: State-of-the-Art", *Nature Biotechnology* 16:40-44; Marshall *et al.* (1998) "DNA Chips: An Array of Possibilities", *Nature Biotechnology* 16:27-31; Wodicka *et al.* (1997) "Genome-Wide Expression Monitoring In *Saccharomyces Cerevisiae*", *Nature Biotechnology* 15:1359-1367; Southern *et al.* (1999) "Molecular Interactions On Microarrays", *Nature Genetics* 21(1):5-9; Duggan, *et al.* (1999) *Nature Genetics* 21(1):10-14; Cheung *et al.* (1999) "Making and Reading Microarrays", *Nature Genetics* 21(1):15-19; Lipshutz *et al.* (1999) "High Density Synthetic Oligonucleotide Arrays", *Nature Genetics* 21(1):20-24; Bowtell (1999) *Nature Genetics* 21:25-32; Brown *et al.* (1999) "Exploring the New World of the Genome with DNA Microarrays" *Nature Genetics* 21(1):33-37; Cole *et al.* (1999)

- “The Genetics of Cancer--A 3D Model” *Nature Genetics* 21(1):38-41; Hacia (1999) “Resequencing and Mutational Analysis Using Oligonucleotide Microarrays”, *Nature Genetics* 21(1):42-47; Debouck *et al.* (1999) “DNA Microarrays in Drug Discovery and Development”, *Nature Genetics* 21(1):48-50; Bassett, Jr. *et al.* (1999) “Gene Expression Informatics--It’s All In Your Mine”, *Nature Genetics* 21(1):51-55;
- 5 Chakravarti (1999) “Population Genetic--Making Sense Out of Sequence”, *Nature Genetics* 21(1):56-60; Chee *et al.* (1996) “Accessing Genetic Information with High-Density DNA Arrays”, *Science* 274:610-614; Lockhart *et al.* (1996) “Expression Monitoring by Hybridization to High-Density Oligonucleotide Arrays”, *Nature*
- 10 *Biotechnology* 14:1675-1680; Tamayo *et al.* (1999) “Interpreting Patterns of Gene Expression with Self-Organizing Maps: Methods and Application to Hematopoietic Differentiation”, *Proc. Natl. Acad. Sci.* 96:2907-2912; Eisen *et al.* (1998) “Cluster Analysis and Display of Genome-Wide Expression Patterns”, *Proc. Natl. Acad. Sci.* 95:14863-14868; Wen *et al.* (1998) “Large-Scale Temporal Gene Expression
- 15 Mapping of Central Nervous System Development”, *Proc. Natl. Acad. Sci.* 95:334-339; Ermolaeva *et al.* (1998) “Data Management and Analysis for Gene Expression Arrays”, *Nature Genetics* 20:19-23; Wang *et al.* (1998) “A Strategy for Genome-Wide Gene Analysis: Integrated Procedure for Gene Identification”, *Proc. Natl. Acad. Sci.* 95:11909-11914; U.S. Patent No. 5,837,832; U.S. Patent No. 5,861,242;
- 20 WO 97/10363.

In the instant case, the microarray contains nucleic acid sequences on a Biotyne B filter. However, any medium, including those that are well-known and available to the person of ordinary skill in the art, to which nucleic acids can be affixed in a manner suitable to allow hybridization, are encompassed by the invention.

25 This includes, but is not limited to, any of the membranes disclosed in the references above, which are incorporated herein for reference to those membranes, and other membranes that are commercially available, including but not limited to, nitrocellulose-1, supported nitrocellulose-1, and Biotyne A, which is a neutrally-charged nylon membrane suitable for Southern transfer and dot blotting procedures.

30 (All are available from Life Technologies.)

EXAMPLE

Summary

Programmed cell death (PCD) in rat cerebellar granule neurons (CGNs)
5 induced by potassium (K^+) withdrawal has been shown to depend on *de novo* RNA
synthesis. The inventors characterized this transcriptional component of CGN
programmed cell death using a custom-built brain-biased cDNA array representing
over 7000 different rat genes. Consistent with carefully orchestrated mRNA
regulation, the profiles of 234 differentially expressed genes segregated into distinct
10 temporal groups (immediate early, early, middle, and late) encompassing genes
involved in distinct physiological responses including cell-cell signaling, nuclear
reorganization, apoptosis, and differentiation. A set of 64 genes, including 22 novel
genes, were regulated by both K^+ withdrawal and kainate treatment. Human
homologs were isolated for 8 of these novel regulated genes: The sequences of these
15 human homologs are shown in SEQ ID NOS:1 (human NARC 9B), 2 (human NARC
8B), 3 (human NARC 2A), 4 (human NARC 16B), 5 (human NARC 10C), 6 (human
NARC 1C), 8 (human NARC 1A), and 10 (human NARC 25).
Thus, array technology was used to broadly characterize physiological responses at
the transcriptional level and identify novel genes induced by multiple models of
20 programmed cell death.

Background

In neurons, programmed cell death is an essential component of neuronal
development (Jacobson *et al.* 1997; Pettmann and Henderson (1998); Pettmann and
25 Henderson (1998) *Neuron* 20:633-747) and has been associated with many forms of
neurodegeneration (Hetts (1998) *Journal of the American Medical Association*
279:300-307). In the cerebellum, granule cell development occurs postnatally. The
final number of neurons represents the combined effects of additive processes such as
cell division and subtractive processes such as target-related programmed cell death.
30 Depolarization due to high concentrations (25 mM) of extracellular potassium (K^+)
promotes the survival of cerebellar granule neurons (CGNs) *in vitro*. CGNs
maintained in serum containing medium with high K^+ will undergo programmed cell
death when switched to serum-free medium with low K^+ (5 mM) (D'Mello *et al.*

(1993) *Proc. Natl. Acad. Sci. USA* 90:10989-10993; Miller and Johnson (1996) *Journal of Neuroscience* 16:7487-7495). The resulting programmed cell death has a transcriptional component that can be blocked by inhibitors of new RNA synthesis (Galli *et al.* (1995) *Journal of Neuroscience* 15:1172-1179; and Schulz and Klockgether (1996) *Journal of Neuroscience* 16:4696-4706). Traditionally, the regulation of limited numbers of specific genes were characterized during CGN programmed cell death using Northern nucleic acid hybridization (*e.g.* PTZ-17, Roschier *et al.* (1998) *Biochemical and Biophysical Research Communications* 252:10-13), reverse transcription polymerase chain reaction (RT-PCR; *e.g.* *c-jun*, cyclophilin, cyclin D1, *c-fos* and caspase (Miller *et al.* (1997) *Journal of Cell Biology* 139:205-217), and *in situ* hybridization (*e.g.* RP-8; Owens *et al.* (1995) *Developmental Brain Research* 86:35-47).

High-density cDNA arrays have been successfully used to characterize genome-wide mRNA expression in yeast (Lashkari *et al.* (1997) *Proc. Natl. Acad. Sci. USA* 94:13057-13062; Wodicka *et al.* (1997) *Nature Biotechnology* 15:1997). In higher eukaryotes, the strategy has been to array as many sequences as possible from known genes, from expressed sequence tags (ESTs), or from uncharacterized cDNA clones from a library (Bowtell (1999) *Nature Genetics* 21:25-32; Duggan *et al.* (1999) *Nature Genetics* 21:10-14; Marshall and Hodgson (1998) *Nature Biotechnology* 16:27-31; and Ramsay (1998) *Nature Biotechnology* 16:40-44). Global RNA regulation during cellular processes including cell-cycle regulation (Cho *et al.* (1998) *Molecular Cell* 2:65-73, and Spellman *et al.* (1998) *Mol. Biol. Cell.* 95:14863-14868), fibroblast growth control (Iyer *et al.* (1999) *Science* 283:83-87), metabolic responses to growth medium (Derisi and Brown (1997) *Science* 278: 680-686), and germ cell development (Chu *et al.* (1998) *Science* 282:699-705) have been temporally monitored using arrays. The program of gene expression delineated in these studies demonstrated a correlation between common function and coordinate expression, and also provided a comprehensive, dynamic picture of the processes involved (Brown and Botstein (1999) *Nature Genetics* 21:33-37). For the cellular process of programmed cell death, a DNA chip has been used to identify twelve known genes as differentially expressed between two conditions, etoposide-treated and untreated cells (Wang *et al.* (1999) *FEBS Letters* 445:269-273).

A genome-wide approach for the comprehensive characterization of the transcriptional component of rat CGN programmed cell death and for identification of novel neuronal apoptosis genes requires an array consisting of both known and novel rat cDNAs. The inventors constructed a brain-biased and programmed cell death-enriched clone set by arraying ~7300 consolidated ESTs from two cDNA libraries cloned from rat frontal cortex and differentiated PC12 cells deprived of nerve growth factor (NGF), and >300 genes that are known markers for the central nervous system and/or programmed cell death. They reproducibly and simultaneously monitored the expression of the genes at 1, 3, 6, 12, and 24 hours after K⁺ withdrawal. They then categorized the regulated genes by time course expression pattern to identify cellular processes mobilized by CGN programmed cell death at the RNA level. In particular they focused on the expression profiles of many known pro- and anti-apoptotic regulatory proteins, including transcription factors, Bcl-2 family members, caspases, cyclins, heat shock proteins (HSPs), inhibitors of apoptosis (IAPs), growth factors and receptors, other signal transduction molecules, p53, superoxide dismutases (SODs); and other stress response genes. Finally, they compared the time courses of regulated genes induced by K⁺ withdrawal in the presence or absence of serum to those induced by glutamate toxicity. Thus, they identified a restricted set of relevant genes regulated by multiple models of programmed cell death in CGNs.

Results

Construction and validation of a brain-biased cDNA microarray

In order to characterize the transcriptional component of neuronal apoptosis in rat cerebellar granule neurons, the inventors constructed a cDNA array, called Smart Chip™ I, that contains primarily rat brain genes. Figure 1 shows a schematic representation of the construction of the microarray. Two cDNA libraries were cloned from rat frontal cortex and nerve growth factor-deprived rat PC12 cells to enrich for cDNAs expressed in the central nervous system and in one *in vitro* model of neuronal apoptosis. Expressed sequence tags (ESTs) from the 5'-end were identified for 8,304 clones in the cortical library and 5,680 in the PC12 library. These 13,984 ESTs were condensed into 7,399 unique sequence clusters by using the Basic Local Alignment Search Tool (BLAST) sequence comparison analysis (Altschul *et al.* 1990) to identify ESTs with overlapping sequence. One representative clone was

chosen from each of 7,296 of the unique sequence clusters and prepared for PCR amplification using a robotic sample processor. In addition to the ESTs, PCR templates were prepared for 289 known DNA sequences, including negative controls, genes with known function in the CNS and/or during programmed cell death, and genes previously identified as regulated by CGN programmed cell death using differential display (data not shown). To check the fidelity of the set of array elements, a robotic sample processor was used to randomly choose 212 clones for sequencing. Ten clones produced poor sequence. The remaining 202 matched their seed sequence (data not shown), implicating 100% fidelity in sample tracking.

10 A sample volume of 20 nl from each of the 7584 PCR products was arrayed onto nylon filters at a density of $\sim 64/\text{cm}^2$ using a pin robot. The arrayed DNA elements were denatured and covalently attached to the nylon filters for use in reverse Northern nucleic acid hybridization experiments. In a typical experiment, "radiolabeled RNA", 1 μg polyA RNA radiolabeled by ^{33}P -dCTP incorporation during cDNA synthesis, was hybridized to triplicate arrays following RNA hydrolysis. Subsequently, the filters were washed and exposed to phosphoimage screens. Gene expression was quantified for each array element by digitizing the phosphoimage-captured hybridization signal intensity. Figure 2 illustrates that the coefficient of variation between triplicate hybridizations averaged less than 0.2 for genes whose intensities were above a threshold of 30-40 units. From control experiments when *in vitro* transcribed RNAs were deliberately spiked into samples, this threshold amounted to a copy number of less than 1 in 100,000 (data not shown).

Tissue distribution of brain-biased Smart Chip ESTs

25 To characterize the brain-biased cDNA array and possibly identify brain-specific genes, radiolabeled RNA from ten different normal rat tissues was hybridized to Smart Chip. Compared to heart, kidney, liver, lung, pancreas, skeletal muscle, smooth muscle, spleen, and testes, radiolabeled rat brain RNA produced more hybridization signal intensity against most of the brain-biased array elements. After data normalization and averaging between replicates, the threshold of detection was determined for each experiment and the number of genes detected for each tissue was tabulated (Figure 6). Most (6127 out of 7296) but not all of the ESTs were detected in at least one of the tissues profiled. The number of genes detected in brain was the

highest. 582 genes appeared to be brain-specific, as defined by detection above threshold for brain but below threshold for any of the other nine tissues.

The physiology of CGN KCl/serum-withdrawal as characterized by transcription profiling on Smart Chip

Using the brain-biased, programmed cell death nucleic acid-enriched Smart Chip, global mRNA expression was profiled throughout a time course of KCl/serum-withdrawal-induced cell death in primary cultures of CGNs. The transcription-dependent CGN programmed cell death was coordinated, resulting in less than 30% survival at 24 hours post-withdrawal as quantified by cell counting (data not shown). RNA samples, designated "treated", were isolated at 1, 3, 6, 12, and 24 hours after switching post-natal day eight CGNs from medium containing 5% serum and 25 mM KCl to serum-free medium with 5 mM KCl. For controls, the 5% serum/25 mM KCl medium was replaced, and "sham" RNA at 1, 3, 6, 12, and 24 hours was isolated.

Since the average coefficient of variation for gene expression intensities between triplicate hybridizations was less than 0.2, genes regulated at least three-fold during the time course (790 out of 6818 detected; data not shown) were further addressed. Using hierarchical clustering algorithms (see Experimental Procedures), the regulated genes were ordered based on their gene expression pattern across the ten experimental points (five time points, sham and treated (Figure 3)). The dendrogram in Figure 3 depicts the hierarchy of relatedness between gene expression profiles. The first major branch point segregated those genes regulated by sham treatment (first five columns), and those regulated by KCl/serum-withdrawal treatment only (last five columns). A majority of genes (556) were regulated by sham treatment. These genes included *trk A*, PSD-95, SV 2A, and VAMP 1, and were most likely induced by serum-add-back in the sham since the medium was exchanged at t=0 with unconditioned medium.

Figure 3 shows the expression pattern of 234 programmed cell death-induced genes that were regulated by KCl/serum-withdrawal only, and were not regulated by serum-add-back in the sham experiments. Their coefficient of variation in expression level throughout the five serum-add-back experiments was less than 20%. Since the serum-add-back experiments were non-discriminating for these genes, the serum-add-back data were averaged to generate a single control data set for clustering with the

KCl/serum withdrawal time course. Four apparent temporal regulation classes were designated immediate early (peaking at 1 hour followed by rapid decay), early (peaking at 3-6 hours), middle (peaking at 6-12 hours), and late (up-regulated at 24 hours). Almost all of the immediate early genes encoded proteins with known roles in regulating secretion and synaptic vesicle release including synaptotagmin, synaphin, NSG-1, calcium calmodulin-dependent kinase II, synapsin, complexin, LDL receptor, and fodrin (Figure 7). Histones 1, 2A, and 3 fell in the early class. Middle genes comprised several known genes induced by programmed cell death or stress, including caspase 3, the mammalian oxy R homolog, cytochrome c oxidase and protein phosphatase Wip-1. Functions encoded for by late genes could be effectors of survival mechanisms including inhibitory neurotransmission (GAD, GABA-A receptor, GABA transporter), cell adhesion (nexin, basement membrane protein 40, phosphacan, rat GRASP), down-regulation of excitatory neurotransmission (glutamate transporter, sodium-dependent glutamate/aspartate transporter), leukotriene metabolism (dithiolethione-induced NADP-dependent leukotriene B4 12-hydroxydehydrogenase, leukotriene A-4 hydrolase), protein stabilization (cysteine proteinase inhibitor cystatin C, N-alpha-acetyl transferase, CaBP2, elongation factor 1-gamma, APG-1), and ionic balance and cell volume (SLC12A integral membrane protein transporter). Based on four distinct waves of gene expression, the major transcriptional responses observed for KCl/serum-withdrawal included initial up-regulation of synaptic vesicle release/recycling, then, of histone biosynthesis, followed by various constituents of programmed cell death regulation and stress-response signaling, and finally, of multiple survival mechanisms. The apparent changes in transcription most likely also reflect changes in the relative cell populations, since late mRNAs may be markers of neurons and non-neuronal cells which have survived KCl/serum-withdrawal at 24 hours. Another contributing factor may be the presence of two populations of dying neurons that respond with different kinetics to serum versus KCl withdrawal, as has been described by other groups.

30 **Neuronal apoptosis regulated candidates (NARCs) regulated by multiple models of programmed cell death**

112 novel ESTs were significantly regulated by KCl/serum-withdrawal in rat CGNs (data not shown). Some exhibited similar expression profiles throughout

KCl/serum-withdrawal and serum-add-back to genes with known function during programmed cell death, such as caspase 3. The temporally-coupled expression of these novel genes may reflect related functionality with caspase 3, since they probably share common RNA regulatory elements, including those regulating initiation, elongation, processing, and/or stability. Apparent coordinate transcriptional up-regulation of synaptic vesicle release/recycling possibly reflects a physiological response to near cessation of synaptic transmission that may or may not contribute to the programmed cell death pathway. To help further distinguish genes that are specifically regulated in response to programmed cell death, CGN programmed cell death induced by glutamate (excitatory neurotransmitter) toxicity was studied. In addition, the effect of KCl-withdrawal alone on gene expression was examined. This was done under defined medium conditions to minimize the effect of serum on the sham and treated samples.

Rat CGNs from post-natal day seven pups were isolated as before and plated into basal medium Eagle containing "high", 10% dialysed fetal bovine serum, and "high", 25 mM KCl. After two days in culture, the medium was replaced with neurobasal medium supplemented with "low", 0.5% serum, and high KCl. To initiate KCl-withdrawal on day eight, the KCl concentration was switched to 5 mM for the treated samples. The same low serum, high KCl, neurobasal medium was replaced in the controls to minimize gene induction by high serum. For the glutamate toxicity experiment, the cells were treated for 30 min in sodium-free Locke's medium with or without 100 μ M kainate for treated samples and controls, respectively.

After isolation from treated and control samples at 1, 3, 6, and 12 hours after KCl-withdrawal and 2, 4, 6, 12 hours after kainate treatment, mRNA was subjected to expression profiling analysis on Smart Chip I. Figure 4 illustrates the changes in gene expression that occur over time when CGNs are induced to undergo programmed cell death by KCl/serum-withdrawal, KCl-withdrawal alone, or kainate treatment. In the scatter plots, due to differential expression, large numbers of regulated genes migrated away from a line of slope one when withdrawn (W) or treated (T) samples were compared to control (C). The sham treated cells for the KCl/serum-withdrawal clearly responded to basal medium serum-add-back, whereas shams for KCl-withdrawal alone and kainate treatment did not respond to conditioned neurobasal

medium add-back. Profiling across the mRNA levels of thousands of genes provided a clear index of changes in overall cell physiology.

In general, apparent changes in gene expression were less robust in the cells cultured on neurobasal medium. The number of genes detected above threshold was similar for all three paradigms, 6634, 7017, and 6818, respectively, for KCl-withdrawal, kainate treatment, and KCl/serum withdrawal (data not shown). Yet the number of genes regulated by at least three-fold during KCl-withdrawal and kainate treatment was only 156 and 167, respectively (data not shown), compared to the 790 discussed above for KCl/serum withdrawal.

A hierarchical clustering algorithm was used to order the regulated genes based on their gene expression pattern across all CGN programmed cell death paradigms investigated. Twenty-six individual profiling experiments in duplicate or triplicate were performed across the 7584 rat genes on Smart Chip I using mRNA isolated from 5 serum-add-back time points, 5 KCl/serum-withdrawal time points, 4 time points each for sham and KCl-withdrawal, and 4 time points each for sham and kainate treatment.

Figure 4 shows expression clusters generated by one hierarchical clustering algorithm. The inset shows a specific group of genes having similar expression patterns. This group includes genes known to be regulated in programmed cell death, for example caspase 3 and Wip 1, as well as other nucleic acid sequences on the array not previously known to be regulated. Those sequences meeting specific criteria were designated "neuronal apoptosis regulated candidate" (NARC). Criteria for designating such genes were based on specific expression criteria as shown in Figure 4. Nucleic acid sequences having an expression pattern similar to genes known to be involved in apoptosis were designated as NARC sequences.

Gene expression validation by RT-PCR

Although the reproducibility in transcription profiling experiments was quite high (average CV<0.2), the gene expression regulation of known and novel genes was validated by semi-quantitative RT-PCR. The rat CGN model system was used to independently validate the expression of several NARC genes that had shown expression (when hybridized with sequences on the chip) related to programmed cell death. Reverse transcriptase-assisted PCR was performed to assess expression of

NARC 1-7, 9, 12, 13, 15, and 16. Experimental samples received KCl withdrawal treatment. Control samples show cells receiving no treatment. The PCR reactions contained 10, 5, 2.5, 1.3, and 0.7 ng of total RNA each. The RT-PCR protocol is disclosed in the exemplary material herein. NARC 1, 2, 4, 5, 7, 9, 12, 13, 15 and 16 all showed significantly increased expression 3-6 hours after KCl withdrawal. The designation "N" above is an abbreviation of the acronym "NARC" which is an abbreviation of "neuronal apoptosis regulated candidate" as described in the Examples section.

10

NARC1 and NARC2 regulation *in vivo* during cerebellar development

Two novel neuronal apoptosis regulated candidates, NARC1 and NARC2, were validated by *in situ* hybridization and shown to be coordinately up-regulated with caspase 3 during postnatal development when increased apoptosis is associated with synapse consolidation in the cerebellum (not shown).

15

Experimental Procedures

BLAST sequence comparison analysis

20

ESTs determined for the 5'-end of cDNA clones picked from two cDNA libraries, rat frontal cortex (8,304 clones) and NGF-deprived differentiated PC12 cells (5,680 clones), ranged from 100-1000 nt in sequence length and averaged 500 nt (data not shown). Sequence comparisons were done using BLAST (Altschul et al. 1990). Contiguous matches defined a sequence cluster. Large clusters were checked by hand to eliminate apparent chimeras. From 13,984 sequences inputted, the analysis identified 5,779 singletons and 1,620 larger clusters (data not shown). The 5'-most clone was selected from the larger clusters. Because two 96-well microtiter plates of clones were missing, a total of 7,296 out of the 7,399 identified were selected for Smart Chip™ I.

25

30

cDNA microarray construction

Using a Genesis RSP 150 robotic sample processor (Tecan AG, Switzerland), bacterial cultures of individual EST clones from the two libraries were consolidated

from 13,792 clones spanning 144 96-well microtiter plates to 7296 Smart Chip 1 clones spanning 76 plates. To prepare templates for array elements, oligonucleotide primers specific for vector sequences up- and downstream of the cloning site were used to amplify the cDNA insert by PCR. Following ethanol precipitation and concentration (to 1-10 mg/ml), the array element templates were resuspended in 3X SSC (1X SSC: 150 mM sodium chloride, 15 mM sodium citrate, pH 7.0). A sample volume of 20 nl from each template was arrayed onto nylon filters (Biodyne B, Gibco BRL Life Technologies, Gaithersburg, MD) at a density of $\sim 64/\text{cm}^2$ using a 96-well format pin robot (THOR). After the filters were dry, the arrayed DNA was denatured in 0.4 M sodium hydroxide, neutralized in 0.1 M Tris-HCl, pH 7.5, rinsed in 2X SSC, and dried to completion.

Array hybridization

Rat poly A⁺ RNA was purchased from Clontech (Palo Alto, CA) for the organ recital (Figure 8) or was isolated as total RNA from cultured CGNs using RNA STAT-60™ (Tel-Test, Inc., Friendswood, TX) and then prepared using Oligotex™ (Qiagen, Inc., Chatsworth, CA). Re-annealed 1 µg mRNA and 1 µg oligo(dT)₃₀ was incubated at 50°C for 30 min with SuperScript™ II as recommended by Gibco in the presence of 0.5 mM each deoxynucleotide dATP, dGTP, and dTTP, and 100 µCi $\alpha^{33}\text{P}$ -dCTP (2000-4000 Ci/mmol; NEN™ Life Science Products, Boston, MA). After purification over Chroma Spin™ +TE-30 columns (Clontech), the labeled cDNA was annealed with 10 µg poly(dA)_{>200} and 10 µg rat Cot-1 DNA (prepared as described in Britten *et al.* (1974) *Methods in Enzymology* 29:263-418). At 2×10^6 cpm/ml, the annealed cDNA mixture was added to array filters in pre-annealing solution containing 100 mg/ml sheared salmon sperm DNA in 7% SDS (sodium dodecyl sulfate), 0.25 M sodium phosphate, 1 mM ethylenediaminetetraacetic acid, and 10% formamide. Following over night hybridization at 65°C in a rotisserie-style incubator (Robbins Scientific, Sunnyvale, CA), the array filters were washed twice for 15 min at 22°C in 2X SSC, 1% SDS, twice for 30 min at 65°C in 0.2X SSC, 0.5% SDS, and twice for 15 min at 22°C in 2X SSC. The array filters were then dried and exposed to phosphoimage screens for 48 h. The radioactive hybridization signals were captured with a Fuji BAS 2500 phosphoimager and quantified using Array Vision™ software (Imaging Research Inc., Canada). Array hybridizations for the organ recital, the CGN

KCl only-withdrawal, and the CGN kainate treatment experiments were performed in triplicate; for the CGN KCl/serum-withdrawal, they were performed in duplicate.

Transcription profiling data analysis

5 For replicate array hybridizations, the distribution of signal intensities across all rat genes was normalized to a median of 100. Replicate measurements were averaged and a coefficient of variation (CV; standard deviation/mean for triplicates or the absolute value of the difference/mean for duplicates) was determined for each gene. The detection threshold was chosen for each hybridization experiment by
10 graphing the moving average (with a window of 200) for CV versus mean gene expression intensity (Figure 2). The threshold was defined as the intensity at which lower intensities exhibited an average CV that was greater than 0.3. For most experiments, this threshold ranged from 10 to 40, and the number of genes detected above threshold ranged from 70% to 95%.

15

CGN cell culture

CGNs were prepared from seven day old rat pups as previously described (Johnson and Miller (1996) *Journal of Neuroscience* 16:74877-7495). Briefly, cerebella were isolated, and meningeal layers and blood vessels were removed under
20 a dissecting scope. Dissociated cells were plated at a density of 2.3×10^5 cells/cm² in basal medium Eagle (BME; Gibco) supplemented with 25 mM KCl, 10% dialyzed fetal bovine serum (Summit Biotechnology lot #04D35, Ft. Collins, CO), 100 U/ml penicillin, and 100 µg/ml streptomycin. Aphidicolin (Sigma, St. Louis, MO) was added to the cultures at 3.3 µg/ml, 24 hours after initial plating to reduce the number
25 of non-neuronal cells to less than 1-5%.

For KCl/serum-withdrawal experiments, after seven days in culture, the treated cells were switched to 5 mM KCl, BME, no serum, while the shams received a medium replacement. By 24 hours post-withdrawal, less than 30% of the cells were surviving as assayed by Hoechts cell counts (data not shown). This apparent cell
30 death could be rescued by actinomycin D at 2 µg/ml (data not shown).

For the KCl-withdrawal alone and kainate treatment experiments, on day two in culture, the medium was replaced with neurobasal medium (Gibco) supplemented with 25 mM KCl, 0.5% dialyzed fetal bovine serum, B27 supplement (Gibco), 0.5

mM L-glutamine (Gibco), 0.1 mg/ml AlbuMAX I (Gibco), 100 U/ml penicillin, 100 µg/ml streptomycin, and 3.3 µg/ml aphidicolin. On day seven, KCl-withdrawal was initiated by replacing the medium with 5 mM KCl while the shams received 25 mM. By 24 hours post-withdrawal, 40% of the cells were surviving as assayed by Hoechts cell counts (data not shown). As previously described, glutamate toxicity was induced by replacing the medium for 30 min with 5 mM KCl, 100 µM kainic acid (Sigma) in sodium free Locke's buffer, while the shams received no kainic acid (Coyle *et al.* (1996) *Neuroscience* 74:675-683). After 30 min, the supplemented neurobasal medium was replaced. By 12 hours post-withdrawal, 30% of the cells were surviving as assayed by Hoechts cell counts (data not shown). The KCl-withdrawal induced cell death was rescued by actinomycin D, whereas the kainate-induced was not.

Expression data clustering algorithms

After normalization and averaging of the KCl/serum-withdrawal data, 790 genes passed the following criteria over the 10 time points (5 treated, 5 sham) for input into heirarchical clustering analysis: 1. detection, maximum intensity greater than 30; 2. noise filter, the difference between maximum and minimum intensity greater than 30; and 3. regulation, fold induction between maximum and minimum intensity of at least 3 (data not shown). Hierarchical clusters were ordered based on Euclidian distances. 234 out of 790 genes that passed the significance filter described above were not regulated in the controls based on CV less than 0.2 for all five control time points (data not shown).

RT-PCR

Oligonucleotide primer sequences specific for each EST validated by RT-PCR were selected from quality sequence regions and designed to obtain a melting temperature of 55-60°C as predicted by PrimerSelect software (DNASTAR, Inc., Madison, WI) based on DNA stability measurements by (Breslauer *et al.* (1986) *Proc. Natl. Acad. Sci. USA* 83:3746-3750). The Stratagene Opti-Prime™ Kit (La Jolla, CA) was used to determine optimal RT-PCR amplification conditions for each primer pair. RT-PCR reactions on 2-fold serially diluted CGN programmed cell death cDNA were set up using the Genesis RSP 150 robotic sample processor and

incorporating the optimal buffer conditions for each primer pair. Every robot run included primers specific for housekeeping genes to control for day to day differences in cDNA template dilutions. The number of cycles was adjusted to obtain a linear range of amplification by comparing the amount of product made from the serially diluted templates as assessed by agarose gel electrophoresis.

Preparation of Array on Nylon

1. Procedure for Generating Labeled First Strand cDNA Using Superscript II Reverse Transcriptase

1. 10 mL (100 mCi) 33P α -dCTP was dried down by SpeedVac.
2. In a separate tube, the following components were mixed:
 - 1.0 ug Poly A+ RNA or 10 ug Total RNA
 - 1 uL 1 ug/uL oligo-dT(30)
 - x uL DEPC-H₂O, to 10 uL

The above sample was heated at 70°C for 4 minutes and then placed on ice.

3. 8uL from the oligo/RNA mixture (#2) was removed and used to resuspend the dried 3P3. The following components were added to the reaction:
 - 4 uL 5X First Strand Buffer (comes with Superscript II RT)
 - 2 uL 100 mM DTT
 - 1 uL 10 mM dAGT-TPs
 - 1 uL 0.1 mM cold dCTP
 - 1 uL Rnase Inhibitor
 - 1 uL Superscript II RT

The reaction was incubated for 30 minutes at 50°C.

4. After incubation, 2 uL 0.5 M NaOH, and 2 uL 10 mM EDTA were added. The reaction was heated at 65°C, for 10 minutes to degrade RNA template.
5. The volume was brought to 50 uL (i.e., add 26 uL H₂O).
6. One Choma-Spin +TE 30 column (Clontech, #K1321) was prepared for every probe made.
 - a. Air bubbles were removed from the column.

b. The break-away end of the column was removed and the column placed in an empty 2 mL tube and spun for 5 minutes at 700g (in Eppendorf 5415C "3.5").

5 c. The column was removed and the flow-through discarded. The column was placed in clean tube. The probe was added slowly to the center of the column bed without disturbing the matrix so that the liquid did not touch the side of the column and flow down the edge of the column wall.

d. The probe was eluted by spinning the column as above.

10 II. Hybridization

1. The hybridization chamber was preheated to 65°C.

2. 10 mL of 10% Formamide Church Buffer was added. This was placed in the hybridization chamber for around 15 minutes.

15 3. Sheared salmon sperm DNA was denatured at 95°C for 5 minutes, placed on ice, and then added to the hybridization mixture at a final concentration of 100 ug/mL. Prehybridization was for 1.5 hours.

4. The amount of probe was calculated necessary to achieve 2×10^6 cpm/mL for 10 mL.

5. The Cot Annealing Reactions (per bottle) were as follows:

20 Rat probe with Rat Filters:

10ug Poly dA (>200nt)

10ug Rat Cot 10 DNA

25uL 20 x SSC

probe + water to 100uL

25 Mouse probe with Rat Filters:

10ug Poly dA (>200nt)

10ug Mouse Cot 1 DNA

25uL 20 x SSC

probe + water to 100uL

30 Also added 5ug Rat Cot 10 DNA to the prehybridization.

Human probe with Human Filters:

10ug Poly dA (>200nt)

10ug Human Cot 1 DNA

25uL 20 x SSC

probe + water to 100uL

The probe was heated to 95°C, and then probe was allowed to preanneal at 65°C, for
5 1.5 hours.

6. The probe was added to prehybridizing filters (directly to the solution and not onto the filters) and hybridization was for approximately 20 hours.

III. Washing

10 1. Probe was removed.
2. Three quick washes were performed with preheated 2 x SSC/1% SDS, 65°C (washes could be done in roller bottles).

3. Two washes were performed for 15 minutes each with preheated high stringency wash buffer:

15 0.5 x SSC, 0.1% SDS for cross species washes

0.5 x SSC, 0.1% SDS for normal washes

0.1 x SSC, 0.1% SDS for very high stringency washes

4. After the high stringency washes, the filters were rinsed in a large square petri dish in 2 x SSC, no SDS. For experiments in which many filters are
20 used, the 2 x SSC is frequently changed so there is no residual SDS left on the filters.

5. The filters were removed from the 2 x SSC and placed on Whatman filter paper. Filters were baked at 85°C for 1 hour or longer. Screens were protected against any moisture. Filters were placed on a blank phosphorimager screen. No yellowed phosphorimager screens were used since they may not respond to
25 exposure linearly. Screens had been erased on a light box for no less than 20 minutes.

6. Blots were exposed to the screen at least 48 hours or as necessary.

IV. Scanning Filters on Fuji Phosphorimager

30 Gradation 16 bit, Resolution 50m, Dynamic Range S4000, select Read and Launch Image Gauge. Image was saved on the hard drive.

APPENDIX I:

10% Formamide-Church Buffer:

- 59.6mL water
- 70mL 20% SDS
- 50mL 2M NaPO₄ pH 7.2
- 5 20mL Ultrapure Formamide
- 0.4mL 0.5M EDTA pH 8.0

The above components were added to water, mixed, and filtered through a 0.2 um filter.

10 RT-PCR Protocol

I. For one PCR reaction mix, the following components were used:

- 28ul 5X First Strand Buffer
- 14ul 0.1M DTT
- 4ul dNTPs (20 mM)
- 15 7ul Rnase Inhibitor
- 7ul Superscript II

This buffer can be stored at -80°C for 3 months.

II. Total RNA was reversed transcribed as follows:

- 20 1.4ug Total RNA (DNAsed)
- 14ul Random Primers (50ng/ul--Gibco)

Water was added to 60ul. The mixture was incubated at 70°C for 10 minutes and then placed on ice for 2 minutes. 60ul of the RT Reaction Mix was added. Incubation was at room temperature for 10 minutes, then 50°C for 30 minutes, then 90°C for 10
25 minutes. The sample was diluted with 480ul water to result in 10ng per 5ul.

III. The PCR reaction was performed with the following ingredients:

- 5ul 4x PCR Buffer
- 5ul cDNA (at 10ng/5ul)
- 30 5ul 1uM Primer Pair
- 5ul Enzyme Cocktail (0.2ul Hot Start Taq, 1ul 2mM
dNTPs, 3.8ul water

IV. Cycling was as follows:

95°C 15 minutes
94°C 30 seconds
52°C 30 seconds
5 72°C 1 minute
Cycle 26-30 times
72°C 10 minutes
4°C Hold

10 Cerebellar granule cell isolation was performed according to the method disclosed in Johnson *et al.* (1996) *J. Neurosci.* 16:74877-7495.

The induction of apoptosis in neurites induced by kainate is described in *Neurosci.* 75:675-683 (1996). The procedure shown in this reference was followed.

The following parameters were checked:

- 15 (1) Cerebellum granule neuron viability following potassium and serum withdrawal at time points corresponding to PCR-based methods for differential gene expression (Hoechst stain).
- (2) Effects of 2 ug/ml actinomycin D on potassium and serum withdrawal at 24 hours on cerebellar granule neurons; viability by Hoeschst stained cell counts.
- 20 (3) Time course of kainate-induced cell death for parallel analysis of PCR-based method for differential gene expression of CGN Poly A mRNA.
- (4) Time course of kainate-induced (30 minute exposure) apoptosis in CGNs; analysis by Hoechst cell counts.
- (5) Time course of potassium withdrawal apoptosis in CGNs in defined
25 media for PCR-based method for differential gene expression of analysis by Hoechst counts.

30 While this invention has been particularly shown and described with reference to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the spirit and scope of the invention as defined by the appended claims.

THAT WHICH IS CLAIMED:

1. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:
 - (a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, and
 - (b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
5 6, 8, and 10.
2. An isolated nucleic acid molecule consisting of a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - 10 b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10.
3. An isolated nucleic acid molecule consisting of a fragment of a nucleotide sequence selected from the group consisting of:
 - 15 a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10;wherein said fragment is at least 15 nucleotides in length.
- 20 4. A nucleic acid molecule comprising a nucleotide sequence which is at least 60% identical to a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
6, 8, and 10.
- 25 5. A nucleic acid molecule which hybridizes under high stringency conditions to a nucleotide sequence selected from the group consisting of:
 - a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; and
 - b) the complements of the nucleotide sequences shown in SEQ ID NOS: 1-
30 6, 8, and 10.

6. A vector comprising a nucleotide sequence selected from the group consisting of:
- a) the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10,
 - b) a nucleotide sequence which is at least 60% identical to a nucleotide sequence shown in SEQ ID NOS: 1-6, 8, and 10
 - c) a nucleotide sequence which hybridizes under high stringency conditions to a nucleotide sequence shown in SEQ ID NOS: 1-6, 8, and 10.
 - d) a complement of a nucleotide sequence of a, b, or c.
7. The vector of claim 6, wherein the isolated nucleic acid molecule is operably linked to at least one expression control element.
8. A host cell comprising the vector of claim 7.
9. A method for preparing a polypeptide comprising culturing the host cell of claim 8 under conditions in which the nucleic acid molecule is expressed.
10. An isolated polypeptide encoded by the nucleic acid molecule of claim 1.
11. An isolated polypeptide encoded by the nucleic acid molecule of claim 4.
12. An isolated polypeptide encoded by the nucleic acid molecule of claim 5.
13. An antibody which selectively binds to the polypeptide of claim 10.
14. An antibody which selectively binds to the polypeptide of claim 11.
15. An antibody which selectively binds to the polypeptide of claim 12.
16. A method for assaying for the presence of a nucleic acid molecule in a sample, comprising the steps of
- (a) contacting said sample with a nucleic acid probe that selectively hybridizes to the nucleic acid molecule, wherein said nucleic acid probe is selected from

the group consisting of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10; the complements of the sequences shown in SEQ ID NOS: 1-6, 8, and 10; fragments of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, wherein said fragments are at least 15 nucleotides in length; and fragments of the complements of the nucleotide sequences shown in SEQ ID NOS: 1-6, 8, and 10, wherein said fragments of the complements are at least 15 nucleotides in length; and

(b) determining whether the nucleic acid probe binds to a nucleic acid molecule in the sample

10 17. A method for detecting a polypeptide of claim 10 in a sample, comprising the steps of:

(a) contacting the sample with an antibody that binds to a polypeptide of claim 10, and

15 (b) determining whether the compound binds to the polypeptide in the sample.

18. A method for modulating the activity of a polypeptide of claim 10, said method comprising contacting the polypeptide of claim 10 with an agent under conditions that allow the agent to modulate the activity of the polypeptide.

20

19. The method of claim 18, wherein said agent is an antibody that binds to said polypeptide

20. The method of claim 18, wherein said polypeptide is in a cell derived from the central nervous system.

25

21. The method of claim 18, wherein said cell derived from the central nervous system is undergoing aberrant apoptosis.

22. The method of claim 18, wherein said activity is modulated in a subject having or predisposed to having a disorder involving the central nervous system.

30

23. The method of claim 18, wherein said activity is modulated in a subject having or predisposed to having a disorder involving aberrant apoptosis.

24. A method for treating a disorder involving the central nervous system comprising administering any of the polypeptides of claim 10 to a subject having or at risk of developing said disorder.

25. A method for treating a disorder involving aberrant apoptosis comprising administering any of the polypeptides of claim 10 to a subject having or at risk of developing said disorder.

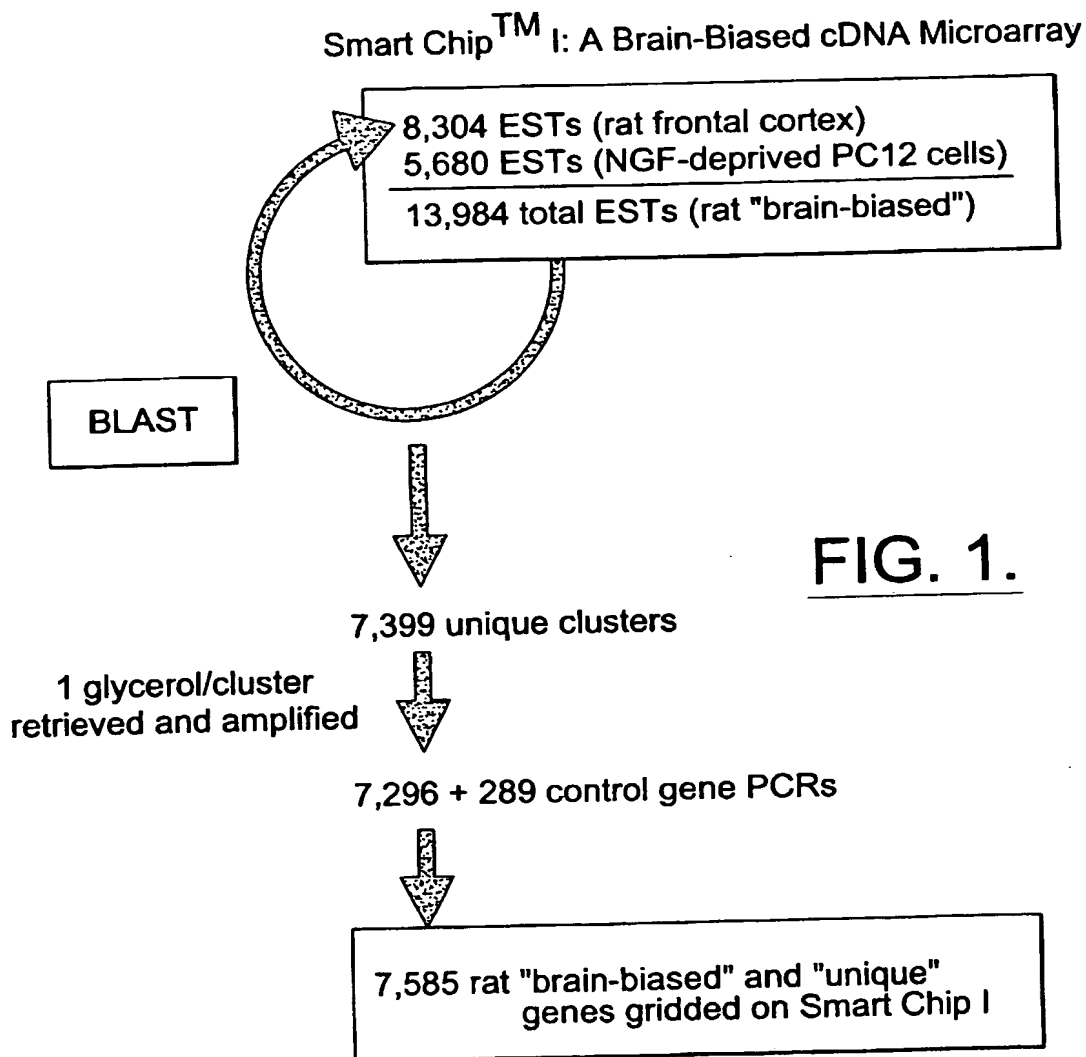
26. A kit comprising a nucleic acid probe which hybridizes to a nucleotide sequence of claim 1 and instructions for use.

27. A kit comprising an agent which binds to a polypeptide of claim 10 and instructions for use.

28. The kit of claim 35, wherein said agent is an antibody.

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1/31



2/31

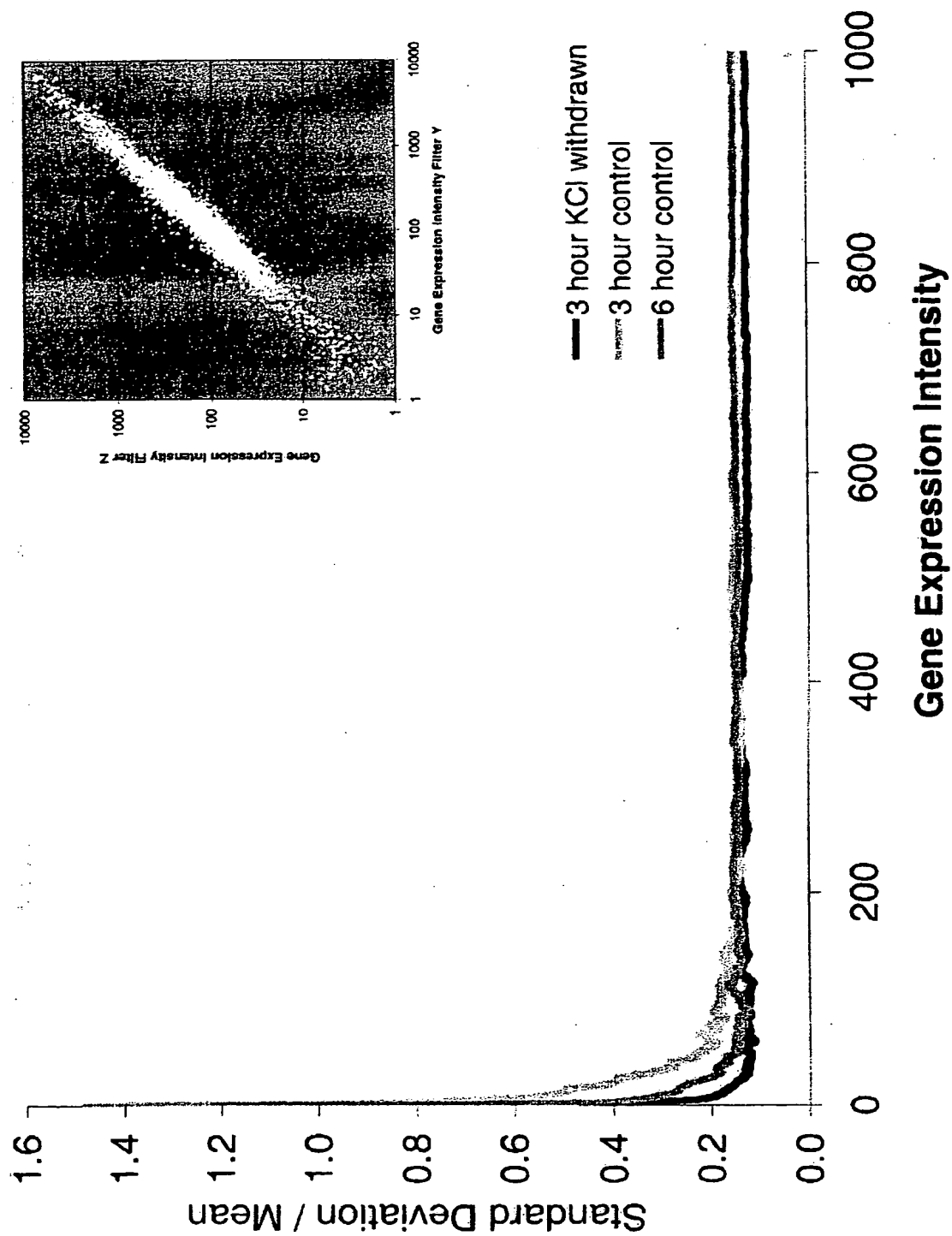


FIG. 2.

3/31

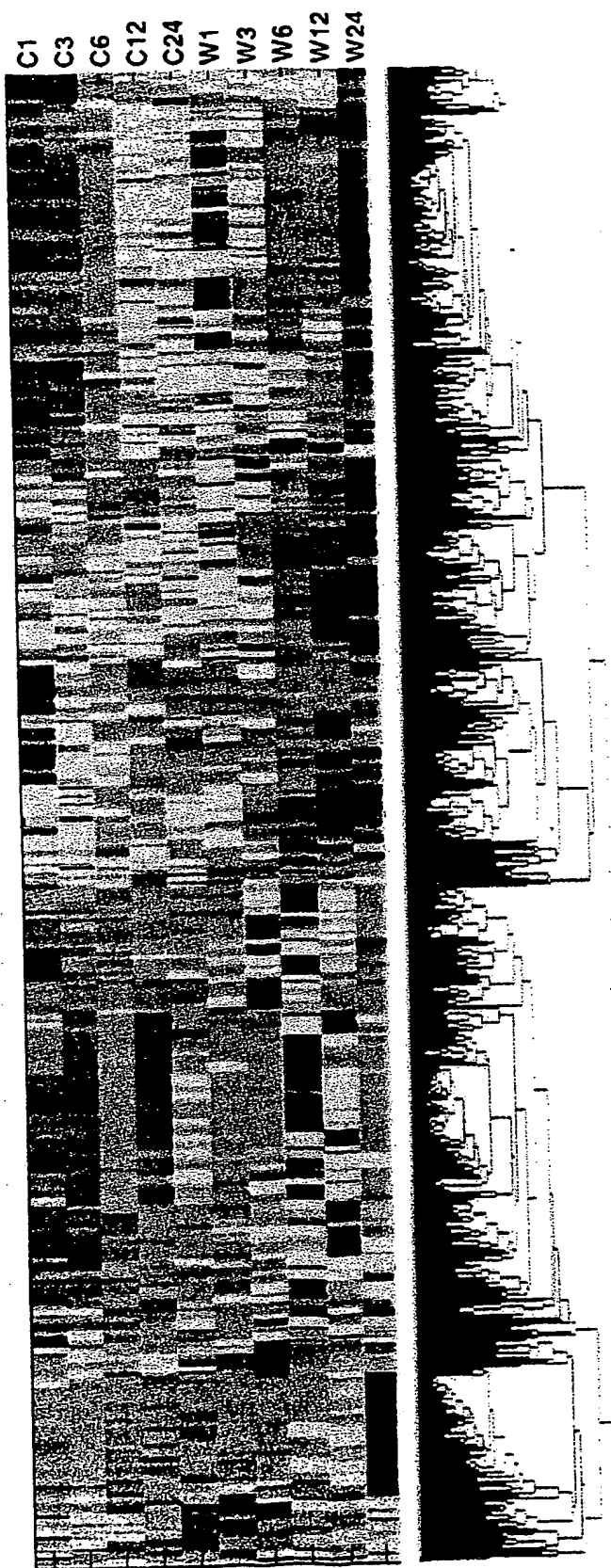
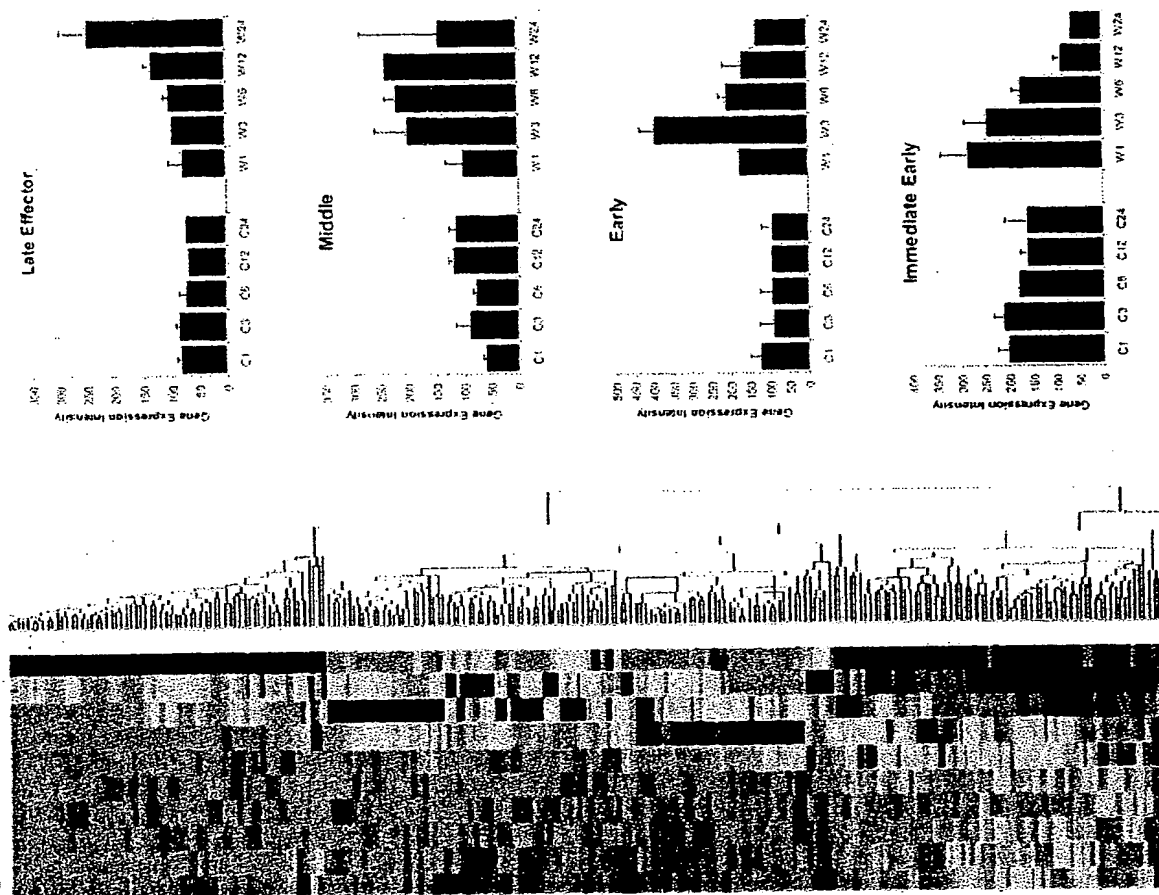


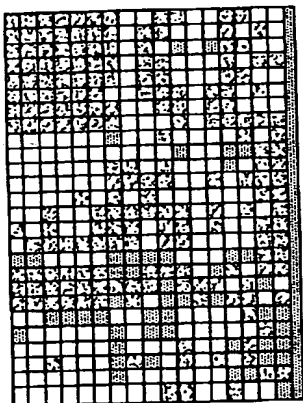
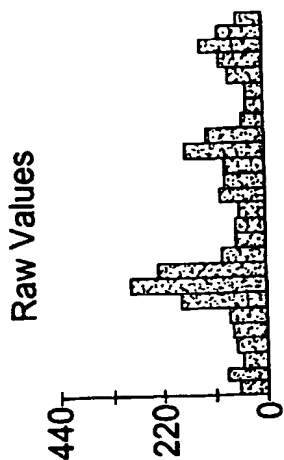
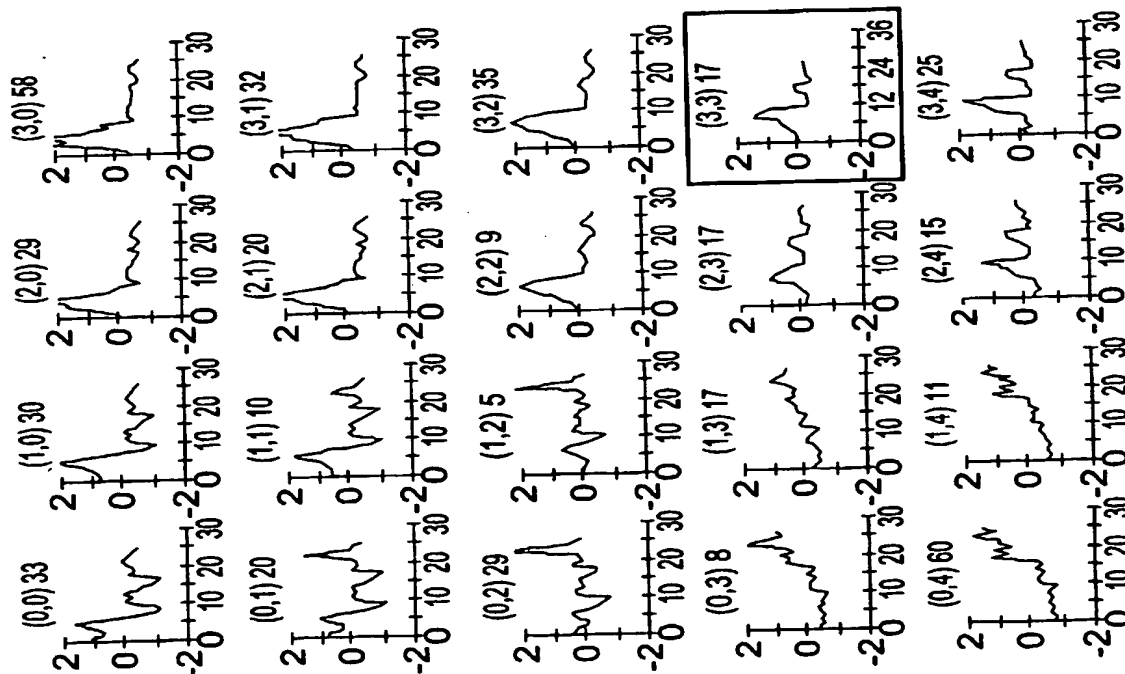
FIG. 3A.

FIG. 3B.



5/31

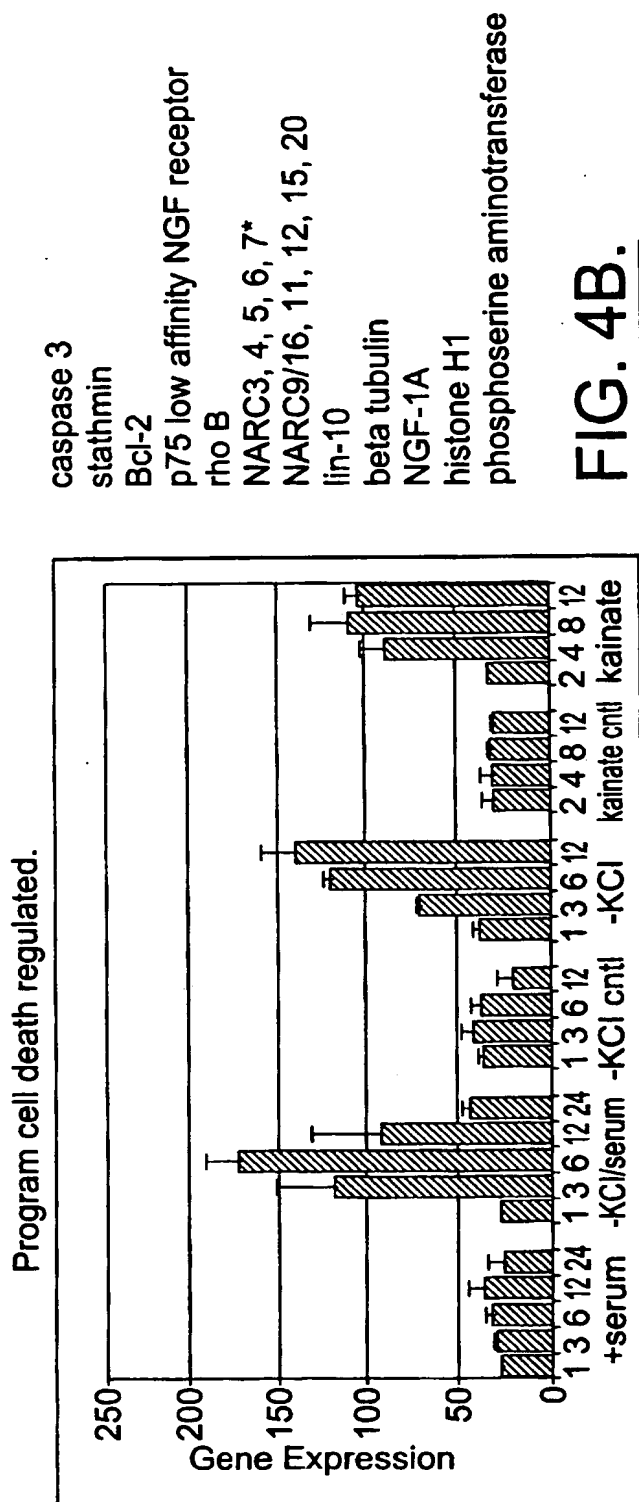
SOM expression clusters.



Array element	Distance
Wip 1	2.395
NARC2	2.470
NARC1	2.626
NARC1	2.635
NARC1	2.649
NARC1	2.784
jtrba021c10t1	3.013
NARC2	3.257
jtrba049c04t1	3.273
jlrxa057d03t1	3.509
caspase 3*	3.509
histone H1	3.654
caspase 3	3.712
histone H1	3.779
jtrba033c06t3	3.872
jlrxa026e08t1	4.368
jtrba080a08t1	4.492

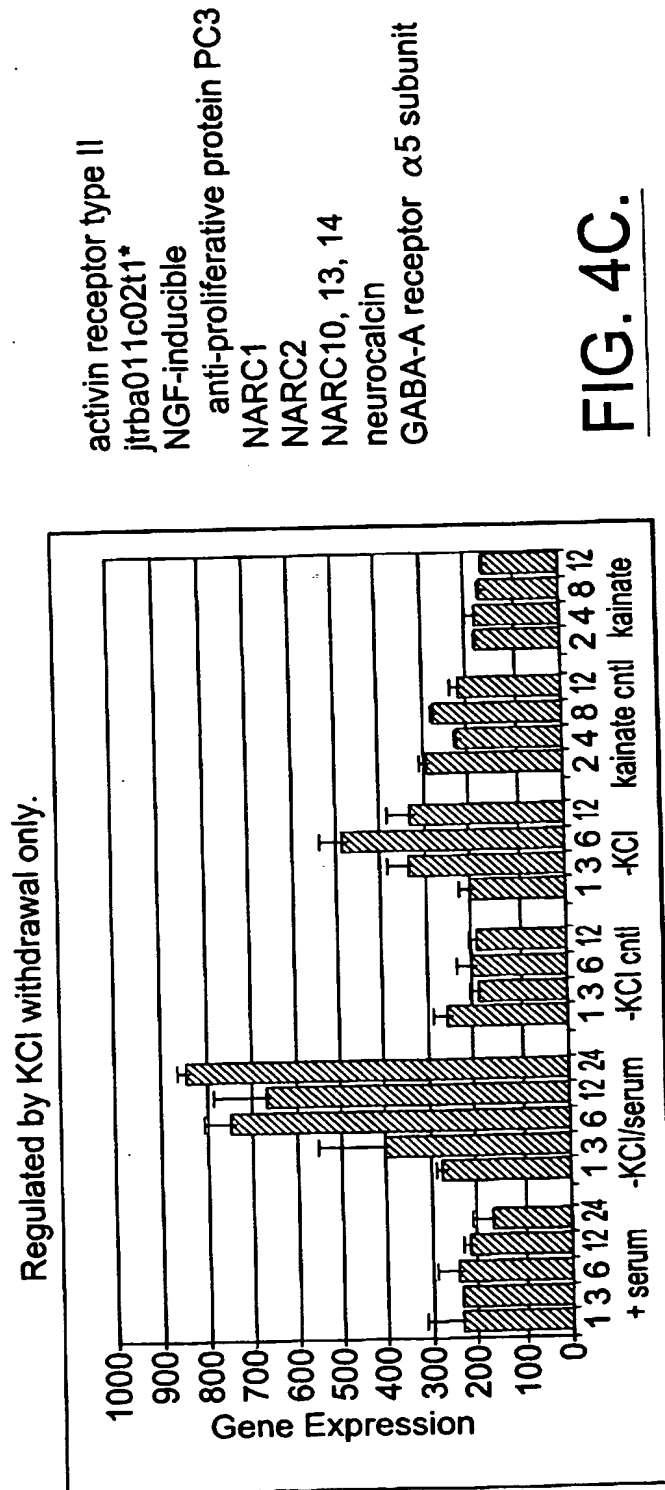
FIG. 4A.

6/31



SUBSTITUTE SHEET (RULE 26)

7/31



8/31

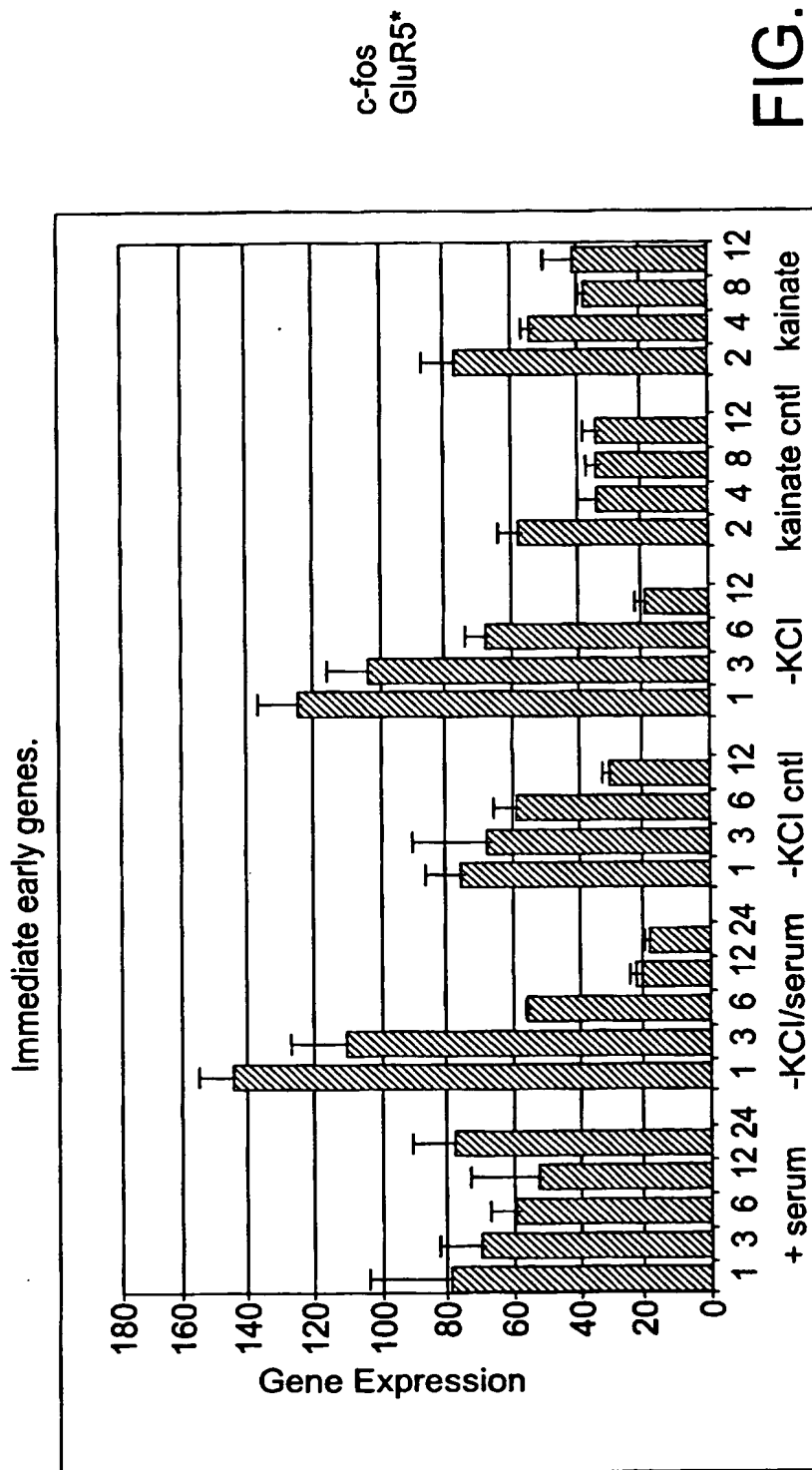


FIG. 4D.

9/31

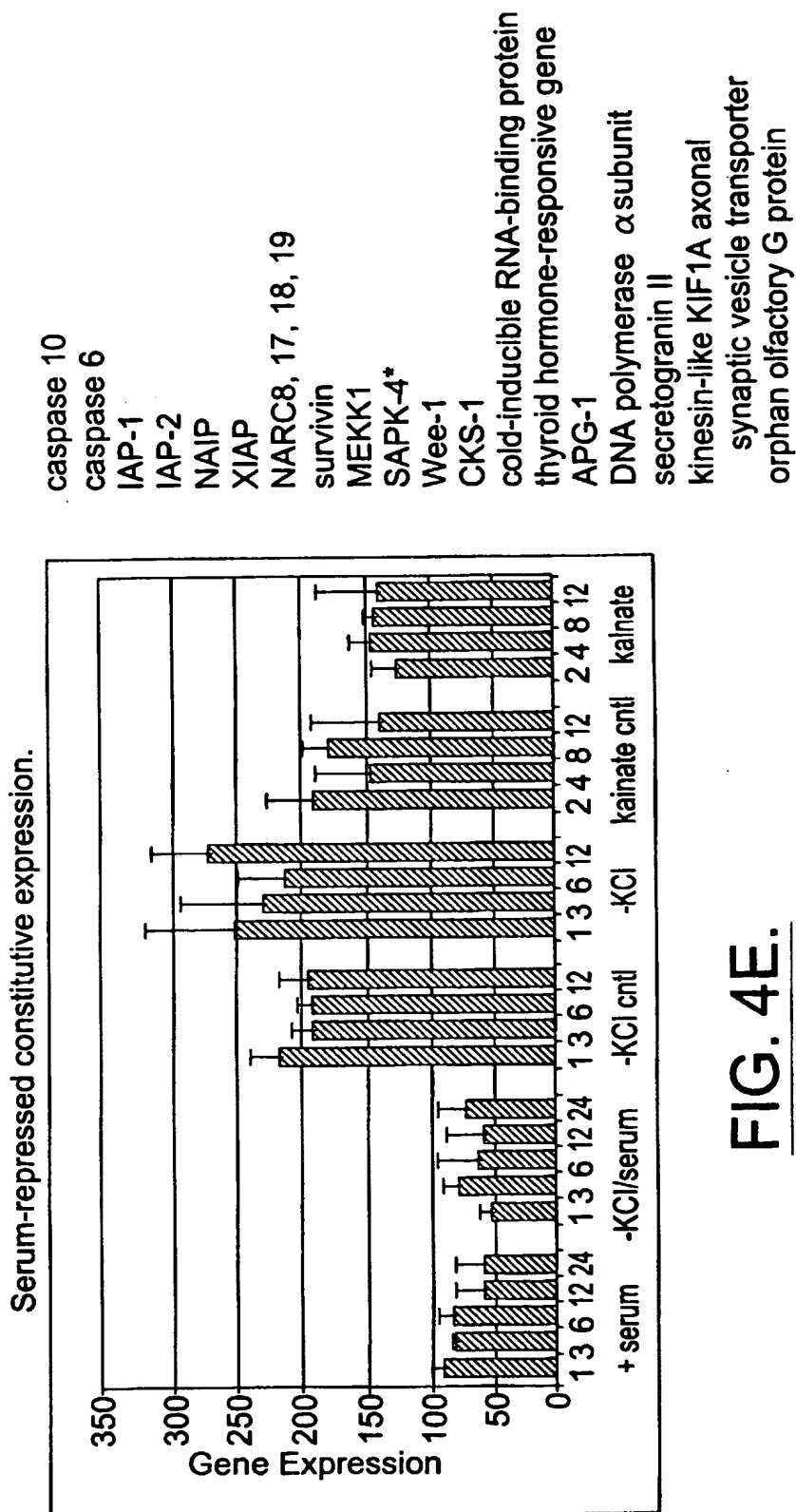


FIG. 4E.

10/31

NARC summary	Gene	Sequence info	Gene Expression Class	MTN	original EST Clone
NARC1	Rat fibNARC1:	1057 nt 3'-UTR.	KCl-regulated	3.4 kb liver testes	rtX020f06b1
NARC2	Rat AIXNARCxc1:	2019 nt; >516 aa CRF that extends upstream of 5' end of sequence. Human fibNARC2A: 1664 nt; >310 aa ORF that extends upstream of 5'; end of sequence. Novel with no apparent homology to any known protein.	KCl-regulated	2.3 kb ubiquitous	rtX022h07b1
NARC3	Rat fibNARC3:	637 nt; >94 aa ORF that extends upstream of 5' end of sequence. Novel with no apparent homology to any known protein.	PCD early		rtX024h09a1
NARC4	Rat fibNARC4:	1247 nt 3'UTR.	PCD intermediate	2.2, 1.8, 1.1 kb ubiquitous	jtrxa012h04f1
NARC5	Rat fibNARC5:	1837 nt; 129 aa ORF. Novel protein in a family including NARC6 containing coil-coil prot-prot domain and transcription factor motif. Homology with NARC6 extends upstream of apparent first ATG suggesting that the cDNA we have sequenced contains a nonsense mutation upstream of the apparent first ATG.	derepressed in low serum PCD intermediate	1.35 & 1.1 kb testes	jtrba034g08f3
NARC6	Rat fibNARC6:	1042 nt; 221 aa ORF. Novel protein in a family including NARC5 containing good coil-coil prot-prot domain and transcription factor motif.	derepressed in low serum PCD intermediate; ischemia 16/C6=3.1	1.35 & 1.1 kb testes+++ brain+	jtrba118a12f1
NARC7	Rat fibNARC7:	2809nt.	PCO intermediate	3.2 kb in brain kidney testes: 1.35 kb ubiquitous	jtrxa027h05f1
NARC8	Rat fibNARC8:	1475 nt; 298 aa ORF. Human fibNARC8B: 1390 nt; >378 aa ORF that extends upstream of 5' end of sequence. Novel splice variant of rat nuclear receptor binding factor 1 that produces a truncated form of the protein.	derepressed in low serum	3.0 kb in heart brain skeletal; 1.35 kb in testes	jtrba022a05f1
NARC9	Rat fibNARC9:	2393 nt; 249 aa ORF. Novel protein in family with NARC16 that has 50% identity, 65% similarity to C.elegans glycerophosphoryl diester phosphodiesterase and yeast Pho85.	PCD early	7.0 kb in brain; 5.0 kb in brain heart spleen lung skeletal testes (liver-may be bubble artifact)	jtrba022a05f1
NARC10	Rat fibNARC10A:	1791 nt; 155 aa ORF. Human fibNARD10C: 2034 nt; 183 aa KCl-regulated ORF of human homolog. Novel protein with 50% identity, 60% similarity to neuron-specific nucleosome assembly protein		2.4 & 2.0 kb brain +++ heart~	jtrba033c06f3

TO FIG. 5B.

FIG. 5A.

11/31

FROM FIG. 5A.

NARC11	Rat fibrinARC11: 1175 nt; ORF that extends upstream of 5' end of sequence with structure proposed in aa sequence figure. Novel splice variant of tyrosine phosphatase.	PCD early	3.5 kb in heart lung liver, 2.0 in spleen testes	jtrba124g04t1
NARC12	Rat fibrinARC12: 2250 nt. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	PCD early (less robust in defined medium)	4.2 kb brain	jtrba031b03t1
NARC13	Rat fibrinARC13: 2046NT; 151 aa ORF. Novel with no apparent homology to any known protein.	KCl regulated		jtrba031h11t1
NARC14	Rat fibrinARC14A: 863 nt; >74 aa ORF that extends upstream of 5' end of sequence. Novel protein similar to human AA316883.	KCl-regulated	1.6 kb in brain; 1.2 ubiquitous	jtrba011c02t1
NARC15	Rat fibrinARC15: 2485 nt 3'UTR. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	PCD down-regulated		jtrba018g08t1
NARC16	Rat fibrinARC16: 3381 nt; 626 aa ORF. Human fibrinARC168: 3206 nt; 673 aa ORF. Novel protein that is similar to C. elegans glycerophosphoryl diester phosphodiesterase and is larger splice variant of NARC9; in particular, homology is to starch binding motif.	PCD early	4.6 & 3.4 kb skeletal heart brain testis	jtrba043g05t1
NARC17	Rat fibrinARC17: 988 nt 3'UTR.	derepressed in low serum		jtrba045c08t1
NARC18	Rat jtrba013h07t1: 263 nt. Novel EST from PC12 NGF-treated cDNA library (1995, PNAS 92:8303).	derepressed in low serum	6.3, 4.6, 3.0, 2.4 kb in brain; 1.2 kb in testes; 1.0 kb in liver testes	jtrba013h07t1
NARC19	Rat fibrinARC19: 1596 nt; 150 aa ORF. Human homolog is epididymal secretory protein.	derepressed in low serum	1.35 kb ubiquitous	jtrba078d11t1
NARC20	Rat fibrinARC20: 3934 nt. Pre-mRNA of myelin basic protein (probably intronic sequence followed by 50 nt of MBP coding region and 3' UTR).	derepressed in low serum; PCD intermediate	2.8 kb brain	jtrba119f08t1
NARC21	Rat jtrba025e06t1: 1435 nt rat contig encodes KIAA0863, 46% identity, 71% similarity to mouse Activity-Dependent Neurotrophic Factor.	KCl/serum-regulated only	ND	jtrba025e06t1
NARC22	Rat jtrba109f02t1: 1749 nt 3'UTR detected in rat config.	serum-regulated		jtrba109f02t1
NARC23	Rat jtrba087f11t1: 724 nt 3'UTR.	PCD intermediate (very low expression)		jtrba087f11t1

FIG. 5B.

TO FIG. 5C.

12/31

FROM FIG. 5B.

NARC24	Rat jtrxa009d08t1: >385aa ORF in rat config (that extends past 5' end. 65% identity, 78% similarity to novel C.elegans ORF.	PCD early	2.5 kb brain	jtrxa009d08t1
NARC25	Rat ftrbNARC25: 1053 nt; >49 aa ORF that extends upstream of 5' end of sequence. Novel protein that is similar to synaptotagmin and inositol polyphosphate binding protein.	FN modulated ischemia-induced	4.8 kb brain	ftrb017g03b1 ftrb017f03b1 ftrb017e03b1
NARC26	Rat ftrbNARC26: 1788 nt; 333 aa ORF. Novel with no apparent homology to any known protein.	FN modulated ischemia-induced	4.4 kb	jtrba057f09t1
NARC27	Rat ftrbNARC27: 1403 nt; 429 aa ORF. Similar to "human secreted protein"; however, clone has much larger ORF with GTPase activation domain; longer ORf does not have signal peptide sequence; homology to centaurin.	FN modulated ischemia-induced	2.6 & 2.0 kb ubiquitous	jtrba094b04t1
NARC28	Rat ftrbNARC28: 1298 nt; >80 aa ORF that extends upstream of 5' end of sequence. Novel with no apparent homology to any known protein.	FN modulated ischemia-induced	6.4 kb in brain heart kidney; 4.5 kb in heart brain lung kidney testes	jtrba111f05t1
NARC30	Rat ftrbNARC30: 4015 nt; >1212 aa ORF that extends upstream of 5' end of sequence. Similar to cell-death regulated secreted protein CW976_1; clone has a much larger ORF.	PCD early	4.2 kb brain heart lung skeletal kidney testes	jtrba057e01t1
NARC31			2.3 & 1.8 kb	jtrxa001g08t2

FIG. 5C.

13/31

Clone	Comment	Expression pattern
jtrb073h10t1	activin receptor type II	induced by KCl-withdrawal
jtrba001e02t1	ApoE, apolipoprotein E	induced by serum-withdrawal
jtrxa057f10t1	A-Raf proto-oncogene serine/threonine-protein kinase	constitutive
jtrxa055h12t1,	BAD	constitutive
mBcl-2	Bcl-2, mouse	induced by PCD
jthtb050a01	BclZ-2, human	basal expression repressed by serum-free conditions
jtrba012d09t1	BDNF	constitutive
jtrxa049d04t1	BMP-2, bone morphogenetic protein 2	constitutive
jtrba041d08t1,	BRCA2	constitutive
jthma018f01	caspase 10, human	basal expression derepressed by serum-free conditions
jtrba114c12t1,	caspase 6	basal expression derepressed by serum-free conditions
c-fos r,	c-fos	immediate early expression in all treatments
c-jun r	c-jun cpg	immediate early expression after KCl+serum withdrawal only
jfrxa024e05t1	CREB-RP, CREB related protein	induced by serum-addback, down-regulated by serum-withdrawal
jtrxa006e04t1	cyclin A	basal expression repressed by serum-free conditions
jtrxa012d04t1	cyclin B2	induced by kainate treatment
jtrxa039e08t1,	cyclin D-interacting myb-like protein	induced by kainate treatment
jtrba053h10t1,	(Dmp1)	
jtrxa006c09t1,	cyclin G, P53-activated	constitutive
jtrba106b06t1,	cyclin-dependent kinase 5 regulatory	constitutive
jtrba004g05t1	subunit p35 (Cdk5r)	
jtrba029b06t1	cyclin-dependent kinases regulatory subunit 1 (CKS-1)	induced by serum-withdrawal, down-regulated by serum-addback; basal expression derepressed by serum-free conditions
GAP43f.l.r	GAP 43	constitutive
jtrxa029h09t2,	Hsp 27	constitutive
jtrba086f03t1,	Hsp 60	constitutive
Hsp70cpg271	Hsp 70 cpg	immediate early expression after kainate treatment only; basal expression repressed by serum-free conditions
jtrxa001d09t2	Hsp 86, chaperonin	constitutive
jthqb017f10	IAP 1, human	basal expression derepressed by serum-free conditions
jthdc128h09t2	IAP-2, human	2 basal expression derepressed by serum-free conditions

FIG. 6A.

14/31

Clone	Comment	Expression pattern
IGF	IGF, human	down-regulated by KCl-withdrawal; completely absent in kainate experiment
jtrxa006b09t1	IGF-II receptor	constitutive
jtrba071b09t1,	IL-6 receptor	constitutive
jtrba032a06t3	insulin receptor tyrosine kinase 53 kDa substrate	basal expression repressed by serum-free conditions
jtrxa001c01t2	insulin-induced growth response protein (CL-6)	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by
jtrba109c03t1	insulin-like growth factor II	constitutive
jtrba054b04t1	MAP kinase kinase	constitutive
jtrba096f01t1	MAP kinase kinase kinase 1 (MEKK1)	basal expression derepressed by serum-free conditions
erk2 h,	MAP kinase, human erk 2, p38	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by serum-free conditions
jtrba089d11t1	MAPKAP kinase, mitogen activated protein kinase activated protein kinase-3	constitutive
jtrxa027g07t1	MDM-2, p53 associated protein	constitutive
jthua018h08	NAIP	basal expression derepressed by serum-free conditions
p75 r	NGF receptor p75, low affinity	immediate early expression after induction of PCD
NGF m	NGF, mouse	induced by serum-addback, down-regulated by serum-withdrawal; basal expression repressed by serum-free conditions
jtrxa005d10t1	NGF-receptor, fast	constitutive
jtrxa001c03t2	p53	induced by serum-addback, down-regulated by
jtrba062e03t1	ras inhibitor	constitutive
jtrba002a07t1,	ras protein p21	constitutive
jtrba117b02t1	RB protein binding protein	constitutive
jtrxa024h04t1	RBP, retinoblastoma-binding protein (RbAp46)	constitutive
jtrba103c03t2	RBP1, retinoblastoma binding protein 1	
jtrxa047a06t1	RBP2, retinoblastoma binding protein 2	basal expression repressed by serum-free conditions
jtrxa007b03t1,	rho	constitutive
jtrba125h09t1	Rho GDP-dissociation inhibitor 1	induced by serum-withdrawal, down-regulated by serum-addback

FIG. 6B.

15/31

Clone	Comment	Expression pattern
jtrxa027d05t1	RIP, nucleoporin-like protein (Rev interacting protein, Rev/Rex activation domain binding protein)	constitutive
jtrxa005c04t1	SKP1, cell-cycle control	
jtrxa038f09t1	SOD-1	constitutive
jtrba041d05t1	SOD-2	constitutive
jtrxa044d10t1	Ste-20 like kinase YSK-1	constitutive
jtrxa025c05t1	Ste20-like kinase, 54% identity, 73% identity to human Ste20-like kinase	constitutive
jtrxa013g03t1	stress-activated protein kinase 4	basal expression derepressed by serum-free conditions
jthta102e01	survivin, human	basal expression derepressed by serum-free conditions
jtrxa001d05t2	trk A	induced by serum-addback, down-regulated by serum-withdrawal
trkB r 2	trk B	basal expression repressed by serum-free conditions
jtrba002b11t1	trk C	constitutive
jtrxa018h09t2	wee 1 tyrosine kinase	basal expression derepressed by serum-free conditions
jchrb018f03	XIAP, human	basal expression derepressed by serum-free conditions

FIG. 6C.

Clone	Comments	16/31	SOM Id
	<u>transient down-regulation by serum, down-regulated by KCl withdrawal (secretion/synoptic residue release)</u>		
jtrxa046c07t1	synaptotagmin synaptic vesicle protein		(0,0)
jtrxa040b12t1	novel		(0,0)
jtrba016d09t1			(0,0)
jtrba010a11t1	rat inositol 1,4,5-triphosphate receptor		(0,0)
jtrba083g04t1	CDC10 homologue involved in cytokinesis		(0,0)
jtrba086f01t1	kynein intermediate chain		(0,0)
jtrxa011g07t1	fodrin (cytoskeletal component of Ca ⁺⁺ -secretion)		(0,0)
jtrxa060g03t1	(NAD ⁺) isocitrate dehydrogenase in mitochondria.		(0,0)
jtrxa055g08t1	neuroendocrine secretory protein 55 chromagranin family		(0,0)
jtrba028h11t1			(0,0)
jtrba005a01t1			(0,0)
jtrba061f02t1			(0,0)
jtrba039g01t1	Na ⁺ , K ⁺ -ATPase beta subunit		(0,0)
jtrba049a02t1	rat inositol 1,4,5-trisphosphate receptor		(0,0)
jtrba027c04t1			(0,0)
jtrba021f01t1	14-3-3 protein tau multifunctional regulator of CaMKII and PKC		(0,0)
jtrxa041a03t1	secretogranin neurendocrine secretory granule protein chromagranin family		(0,0)
jtrba010e10t1	succinyl-coa Sketoacid-coenzyme in mitochondria		(0,0)
jtrba035a02t1	synaphin 2 associated with docking/fusion complex		(0,0)
jtrxa038g09t1	novel		(0,0)
jtrba032c01t3	complexin cytosolic proteins that regulate SNAP receptor function		(0,0)
jtrxa032h06t1	cytochrome c in mitochondria		(0,0)
secretograninc	secretogranin		(0,0)
jtrba021e08t1			(0,0)
jtrba067h01t1			(0,0)
jtrba015a11t1	RIG (down-Regulated in Glioma)		(0,0)
jtrba012g02t1	alpha-tubulin-1		(0,0)
jtrxa027b05t1			(0,0)
p75 r	low affinity NGF receptor p75		(0,0)
jtrba043g04t1	DAP-5, a novel homolog of eukaryotic translation initiation factor 4G isolated as a putative modulator of gamma interferon-induced cell death		(0,0)
jtrba090c02t1			(0,0)
jtrba020h04t1			(0,0)
jtrxa027f11t1	novel		(0,0)
	<u>transient down-regulation by serum, down-regulated by KCl withdrawal and kainate treatment</u>		
rtrX024a09a1			(0,1)
rtrX024b09a1			(0,1)
jfrxa006g12t1	secretogranin I chromogranin family		(0,1)
jtrxa044c10t1			(0,1)
rtrX024c09a1			(0,1)
rtrX024d09a1			(0,1)
rtrX024h09a1			(0,1)

FIG. 7A.

SUBSTITUTE SHEET (RULE 26)

17/31

rtrX024f09a1		(0,1)
rtrX024g09a1		(0,1)
jtrba089e08t1		(0,1)
rtrS024e09a1		(0,1)
jtrba035d02t1	neurocalcin calcium-binding protein	(0,1)
jtrba009b09t2	novel	(0,1)
jtrba122c04t1	voltage-gated potassium channel KV1.4	(0,1)
jtrba025e08t1		(0,1)
rtrX023e08b1		(0,1)
jtrba078f10t1		(0,1)
jtrba050b02t1		(0,1)
jtrba090c03t1		(0,1)
jtrba125h10t1	novel	(0,1)
jtrba045f03t1		(0,2)
jtrxa052b09t1		(0,2)
jtrba049h12t1		(0,2)
jtrxa046g07t1		(0,2)
jtrba069a03t2		(0,2)
jtrxa044d10t1		(0,2)
jtrba034c03t3		(0,2)
jtrba069h07t2		(0,2)
jtrba032h02t3		(0,2)
jtrba024h09t1		(0,2)
jtrba076f12t2		(0,2)
jtrba034h07t3		(0,1)
jtrba029a05t1		(0,2)
jtrxa050b11t1		(0,2)
jtrxa031c06t1		(0,2)
jtrxa054e11t1		(0,2)
jtrba007g02t1		(0,2)
jtrba063b08t1		(0,2)
jtrxa046g08t1		(0,2)
jtrba041b03t1		(0,2)
jtrba005c05t1		(0,2)
jtrba083a05t1		(0,2)
jtrba054b06t1		(0,2)
jtrba011b09t1		(0,2)
	<u>immediate early effect after any perturbation</u>	
jtrba005h05t1	navel	(0,2)
jtrba111e08t2	c-fos	(0,2)
jtrxa055b02t1		(0,2)
c-fos r	c-fos	(0,2)
jtrba018g08t1	novel EST from PC12 NGF-deprived cDNA library (1996, PNAS 92:8303)	(0,2)
	<u>serum-repressed genes</u>	
jtrba060c09t1	(cell proliferation)	(0,3)
jtrxa033d10t1		(0,3)
jtrba093g06t1		(0,3)

FIG. 7B.

SUBSTITUTE SHEET (RULE 26)

18/31

jtrba103b07t2	transferrin transports iron from sites of absorption and heme degradation to those of storage and utilization; serum transferrin may have a role in stimulating cell proliferation	(0,3)
jtrxa035d07t1		(0,3)
jtrxa015g08t1	novel	(0,3)
jtrxa045g12t1		(0,3)
jtrba005d06t1		(0,3)
jtrba046c05t1	<u>serum and KCl regulated</u> (secretion/synaptic residue released cytoskeletal reorganization) calcium-dependent actin-binding protein; activator protein for secretion/exocytosis	(1,0)
jtrxa030e12t1	microtubule-associated protein (MAP1B)	(1,0)
jtrba013g10t1	novel	(1,0)
jtrxa010h04t1	novel	(1,0)
jtrba032f05t3	Vap-33, vesicle-associated membrane binding protein; synaptobrevin binding protein	(1,0)
jtrba057g11t1	GABA-A receptor delta subunit	(1,0)
jtrxa022g12t2	novel	(1,0)
jtrba063e11t1		(1,0)
jtrba033b07t4	novel	(1,0)
jtrxa058h01t1	63% identical, 74% similar to mouse N-myo-downstream gene Ndr1	(1,0)
jtrxa005c02t1	novel	(1,0)
jtrba104d03t1	novel	(1,0)
jtrxa035f07t1	microtubule associated protein MAP-1b	(1,0)
jtrba057d04t1	plasma membrane Ca ²⁺ ATPase-isoform 2	(1,0)
jtrxa049g12t1	novel	(1,0)
jtrba006e10t1	CaMKIIbeta	(1,0)
jtrba009d10t2		(1,0)
jtrba036h03t2	myosin heavy chain	(1,0)
jtrba041h10t1	synapsin Ib	(1,0)
jtrba025h11t1		(1,0)
jtrba035g12t2		(1,0)
jtrba001a08t1	bovine cytochrome b-560, succinate-ubiquinone reductase OPs1	(1,0)
jtrxa056g12t1		(1,0)
jtrxa058a10t3	alpha tubulin	(1,0)
jtrxa058a10t3	alpha tubulin	(1,0)
jtrxa024f05t1	phosphofructokinase C	(1,0)

FIG. 7C.

19/31

jtrxa038e03t1	novel	(1,0)
jtrxa024b07t1	alpha tubulin	(1,0)
jtrxa024g09t1	cell death regulated	(1,0)
jtrba027g06t1	vesicle-associated calmodulin-binding kinase-like protein	(1,1)
jtrba040h07t1	beta-synuclein	(1,1)
jtrxa037f09t1	DNA replication licensing factor required for the entry into S phase and for cell division	(1,1)
jtrba044e03t1		(1,1)
jtrba057e01t1		(1,1)
jtrxa049c07t1	mammalian homolog of Drosophila circadian Perigene	(1,1)
jtrba109f02t1		(1,1)
jtrxa009d08t1		(1,1)
GluR5 r	GluR5	(1,1)
jtrba116b12t1	Hsp60, involved in mitochondrial protein import and macromolecular assembly; may prevent misfolding and promote refolding and proper assembly under stress conditions in the mitochondrial matrix	(1,1)
jtrba031h09t3		
jtrba009h10t2		
jtrxa039e01t1		
jtrba036e10t1		
jtrba035f12t2	<u>derepressed genes after growth in defined medium</u>	
jtrba119f08t1	novel (stress response/hormone response)	(1,3)
jtrba116h12t1	olfactory G protein involved in visual transduction and in mediating the effect of one or mor hormones/neurotransmitters	(1,3)
jtrba023d09t1	DNA polymerase alpha subunit	(1,3)
jtrxa040h01t1	cold-inducible RNA-binding protein mediating cold-inducible suppression (of mammlian cell growth; expressed transiently in developing neural tissue; LV-inducible-cDNA	(1,3)
jthma018t01	caspase 10	(1,3)
jtrxa009f03t1	novel	(1,3)
jtrba078d11t1		(1,3)
jtrxa045c08t1		(1,3)
jtrba016g02t1	thyroid hormone-responsive gene; gene from the Down syndrome optical region resembling a DNA binding protein with an SH3 domain	(1,3)
jtrba002f04t1		(1,3)
jtrba029e02t1		(1,3)
jtrba114c07t1	secretogranin II	(1,3)
jtrba124f02t1	GABA-A receptor alpha-5 subunit	(1,3)
jtrba013h07t1		(1,3)
jtrxa059d02t1		(1,3)
jtrxa053a03t1	lin-10	(1,3)
jtrba075f03t1		(1,3)
	<u>derepressed genes after growth in defined medium</u>	
jtrba034g06t3	novel (stress response/hormone response)	(1,4)

FIG. 7D.

20/31

jtrba109c06t1		(1,4)
jtrba053b09t1		(1,4)
jtrxa032g02t1	novel	(1,4)
jtrba018b01t1	Hsp70-related protein APG-1, induction by hyperosmolar salt stres:	(1,4)
jtrxa026f02t1		(1,4)
jfrxa044a11t1		(1,4)
jtrba118a12t1	novel	(1,4)
jtrxa006b04t1	novel	(1,4)
jtrba068f01t1	progesterone-induced messenger RNA resembling phosphoserine amino transferase required in major phosphorylated pathway of serine and pyridaxine biosynthesis	(1,4)
jtrxa013f01t1	progesterone-induced messenger RNA resembling phosphoserine amino transferase required in major phosphorylated pathway of serine and pyridaxine biosynthesis	(1,4)
<u>Serum and KCl regulated</u>		
jtrba105c11t1	(calcium signal transduction)	(2,0)
SV2a r 1	SV2a	(2,0)
jtrba017f06t1	cytosolic aspartate aminotransferase	(2,0)
jtrba001d01t1	novel (jtrba005f12t1)	(2,0)
jtrba077g09t1		(2,0)
CaMKIIb r	CaMKII beta	(2,0)
SV2a r 2	SV2a	(2,0)
CaMKIIdC r	CaMKII delta C	(2,0)
jtrba005f12t1	novel (jtrba001d01t1)	(2,0)
jtrxa045c04t1	14-3-3 protein gamma-subtype; regulation of protein kinase C	(2,0)
jtrba045c03t1	citrate synthase in mitochondrial matrix	(2,0)
jtrba027h11t4	novel SH3 containing protein	(2,0)
jtrba022g04t1	novel	(2,0)
jtrba075h01t1	vacuolar adenosine triphosphatase subunit responsible for acidifying a variety of intracellular compartments	(2,0)
jtrba073h12t1	14-3-3 protein gamma-subtype; regulation of protein kinase C	(2,0)
jtrba073h12t1	calcium-independent alpha-latrotoxin receptor (secretin GPCR family:	(2,0)
jtrba115h04t1	synapsin 1	(2,0)
jtrxa001e02t2		(2,0)
jtrba091c06t1		(2,0)
jtrba087e08t1	novel	(2,0)
NGFIB r	NGF-Ib	(2,0)
jtrba013b12t1	FUSE binding protein that activates the far upstream element of c-my:	(2,0)
jtrba029a12t1	novel	(2,0)
jtrxa059c04t1	testoterone induced splicing factor SF1	(2,0)
jtrxa019a09t2	TCP-1 chaperonin	(2,0)

FIG. 7E.

21/31

jtrba028g11t1	Stathmin, a ubiquitous phosphoprotein acting as an intracellular re ay of proliferation and differentiation	(2,3)
jtrxa021g01t1	rhoB ras-like immediate early gene	(2,3)
jtrba043g05t1	novel	(2,3)
jtrba022a05t1		(2,3)
jtrba018b03t1		(2,3)
jtrba014b01t1		(2,3)
jtrba043c06t1		(2,3)
jtrba124g12t1	beta tubulin-like protein	(2,3)
jtrba087f11t1		(2,3)
jtrba115d02t1	kinesin-like KIF1A axonal transporter of synaptic vesicles	(2,3)
	Stable transcripts	(2,4)
jtrba034e12t3	(survival factors)	(2,4)
jtrxa054d03t1		(2,4)
jtrxa001e10t2		(2,4)
jtrxa006h05t1		(2,4)
jtrba108b01t1	vimentin	(2,4)
jtrba081c02t1		(2,4)
jtrba113b04t1		(2,4)
jtrba112a12t1		(2,4)
jtrba086b12t1		(2,4)
jtrba023d12t1		(2,4)
jtrba005c11t1		(2,4)
jtrba117c06t1		(2,4)
jtrba005f07t1		(2,4)
jtrxa013g09t1		(2,4)
jtrxa045f01t1		(2,4)
Synaptophysin	synaptophysin	(3,0)
jtrxa045h03t1	synaptotagmin XI	(3,0)
p39	p39	(3,0)
jtrba002e06t1	clathrin-coated vesicle/synaptic vesicle proton pump 116 subunit vaccolar proton pump subunit 1	(3,0)
jtrba084a04t1	novel	(3,0)
N-sec 1 r	syntaxin-binding protein (n-sec1)	(3,0)
jtrba031d04t3	novel	(3,0)
jtrba060c03t1	sodium/potassium-transporting ATPase alpha-3 chain	(3,0)
flh286t		(3,0)
jtrba109d02t1		(3,0)
jtrxa001d05t2	trk A	(3,0)
rlrX019c05a1		(3,0)
jtrba037a07t1	sodium/potassium-transporting ATPase alpha-3 chain	(3,0)
jtrba057f09t1		(3,0)
jtrxa007f11t1	syntaxin-binding protein (n-sect)	(3,0)
jtrxa044b01t1	novel	(3,0)
jtrba030g12t1	novel	(3,0)
jtrba048f02t1	novel	(3,0)
PSD-95 r	PSD-95	(3,0)
jtrxa054b03t1	novel	(3,0)
jtrba013a08t1		(3,0)

FIG. 7F.

SUBSTITUTE SHEET (RULE 26)

22/31

<i>jtrba036a02t2</i>	(3,0)
<i>jtrba041a02t1</i>	(3,0)
<i>jtrba083f09t1</i>	(3,0)
<i>jtrxa056e08t1</i>	(3,0)
<i>jtrxa033f03t1</i>	(3,0)
<i>jtrba053d01t1</i>	(3,0)
<i>jtrba017b08t1</i>	(3,0)
<i>jtrxa046e09t1</i>	(3,0)
<i>jtrba055g03t1</i>	(3,0)
<i>jtrba049b09t1</i>	(3,0)
<i>jtrba094b12t1</i>	(3,0)
<i>jtrxa034e05t1</i>	(3,0)
<i>jtrba109c11t1</i>	(3,0)
<i>jtrba038h12t1</i>	(3,0)
<i>rtrX017e03b1</i>	(3,0)
<i>jtrba086f08t1</i>	(3,0)
<i>jtrxa004b09t1</i>	(3,0)
<i>jtrba019b02t1</i>	(3,0)
<i>jtrba019g02t1</i>	(3,0)
<i>jtrba019f12t1</i>	(3,0)
<i>jtrba117h02t1</i>	(3,0)
<i>rtrX017c03b1</i>	(3,0)
<i>rtrX017f03b1</i>	(3,0)
<i>jtrxa008c04t1</i>	(3,0)
<i>rtrX017g03b1</i>	(3,0)
<i>jtrxa005h09t1</i>	(3,0)
<i>jtrba029d07t1</i>	(3,0)
<i>rtrX015b01a1</i>	(3,0)
<i>jtrxa050h03t1</i>	(3,0)
<i>rtrX015a01a1</i>	(3,0)
<i>jtrba118c02t1</i>	(3,0)
<i>jtrba109f08t1</i>	(3,0)
<i>jtrba083c03t1</i>	(3,0)
<i>jtrba035h01t1</i>	(3,0)
<i>jtrba095g06t1</i>	(3,0)
<i>rfrX015g01a1</i>	(3,0)
<i>jtrba038a08t1</i>	(3,0)
<i>jtrba084b04t1</i>	(3,1)
<i>jtrba084h08t1</i>	(3,1)
<i>jtrba013a09t1</i>	(3,1)
<i>rGFAP 5'1/10 GFAP</i>	(3,0)
<i>RCSP r RGSP</i>	(3,1)
<i>jtrba084b09t1</i>	(3,1)
<i>jtrba045g09t1</i>	(3,0)
<i>syntaxin2 r</i>	(3,1)
<i>r Actin 5' beta actin</i>	(3,1)
<i>jtrxa011f11t1</i>	(3,1)
<i>jtrxa001c12t2</i>	(3,1)
<i>jtrxa038f02t1</i>	(3,1)

FIG. 7G.

SUBSTITUTE SHEET (RULE 26)

23/31

<i>rGFAP 5' RT-1 GFAP</i>	(3, 1)
<i>r Actin 5' 1/10 beta actin</i>	(3, 1)
<i>jtrba069b12t2</i>	(3, 1)
<i>jtrba108b06t1</i>	(3, 1)
<i>r GFAP 5' GFAP</i>	(3, 1)
<i>jtrba046g08t1</i>	(3, 1)
<i>jtrba022f09t1</i>	(3, 1)
<i>jtrba085g11t1</i>	(3, 1)
<i>jtrba073d05t1</i>	(3, 1)
<i>jtrba054g03t1</i>	(3, 1)
<i>r GAPDH glyceraldehyde-3-phosphate-dehydrogenase</i>	(3, 1)
<i>jtrba063h10t1</i>	(3, 1)
<i>erk2 h erk 2</i>	(3, 1)
<i>jtrxa043b04t1</i>	(3, 1)
<i>jtrxa050f07t1</i>	(3, 1)
<i>jtrba021f10t1</i>	(3, 1)
<i>rtrX017h03b1</i>	(3, 1)
<i>jtrba038f01t1</i>	(3, 1)
<i>jtrba078f09t1</i>	(3, 1)
<i>r GFAP M RT GFAP</i>	(3, 2)
<i>jtrba025g01t1</i>	(3, 2)
<i>rGFAP M 1/10 GFAP</i>	(3, 2)
<i>jtrxa005b01t1</i>	(3, 2)
<i>jtrba100e12t1</i>	(3, 2)
<i>rGFAP M GFAP</i>	(3, 2)
<i>rtrX018h04a1</i>	(3, 2)
<i>r Actin M 1/10 beta actin</i>	(3, 2)
<i>jtrxa004d06t1</i>	(3, 2)
<i>jtrxa030b02t1</i>	(3, 2)
<i>jtrba112h05t1</i>	(3, 2)
<i>jtrba120f04t1</i>	(3, 2)
<i>jtrba032e04t3</i>	(3, 2)
<i>jtrba118d04t1</i>	(3, 2)
<i>r Actin M beta actin</i>	(3, 2)
<i>jtrxa052f02t1</i>	(3, 2)
<i>jtrba112h06t1</i>	(3, 2)
<i>jtrba007a11t1</i>	(3, 2)
<i>jtrba125b04t1</i>	(3, 2)
<i>r 18S rRNA 18S rRNA</i>	(3, 2)
<i>jtrba057e05t1</i>	(3, 2)
<i>r nEnolase neuronal enolase</i>	(3, 2)
<i>jtrxa045d02t1</i>	(3, 2)
<i>jtrxa033a12t1</i>	(3, 2)
<i>r GFAP 3' GFAP</i>	(3, 2)
<i>jtrxa011c11t1</i>	(3, 2)
<i>jtrba045d09t1</i>	(3, 2)
<i>rGFAP 3' 1/10 GFAP</i>	(3, 2)
<i>rGFAP 3' RT-1 GFAP</i>	(3, 2)
<i>rGFAP 3'-1/10 GFAP</i>	(3, 2)
<i>jtrxa007b04t1</i>	(3, 2)

FIG. 7H.

24/31

<i>PC12-PTP1 r PC12-PTP1</i>	(3,2)
<i>jtrba087b01t1</i>	(3,2)
<i>jtrba061c01t1</i>	(3,2)
<i>jtrba079h08t2</i>	(3,2)
<i>KCl withdrawal unregulated</i>	
<i>rtrX019b05a1</i> <i>protein phosphatase Wip-1 (apoptosis)</i>	(3,3)
<i>rtrX022h07b1</i>	(3,3)
<i>rtrX020b06b1</i>	(3,3)
<i>rtrX020f06b1</i>	(3,3)
<i>rtrX020e06b1</i>	(3,3)
<i>rtrX020c06b1</i>	(3,3)
<i>jtrba021c10t1</i> <i>novel</i>	(3,3)
<i>rtrX022f07b1</i>	(3,3)
<i>jtrba049c04t1</i> <i>nucleolar ribosomal RNA precursor external transcribed spacer 1 (ETS1)</i>	(3,3)
<i>jtrxa057d03t1</i>	(3,3)
<i>rtrX030f10a1</i> <i>caspase 3</i>	(3,3)
<i>jtrxa057a03t1</i> <i>histone H1</i>	(3,3)
<i>rtrX030b10a1</i> <i>caspase 3</i>	(3,3)
<i>rtrX015h01a1</i>	(3,3)
<i>jtrba033c06t3</i> <i>novel</i>	(3,3)
<i>jtrxa026e08t1</i> <i>novel</i>	(3,3)
<i>jtrba080a08t1</i>	(3,3)
<i>Stable transcripts</i>	
<i>jtrba011c02t1</i> <i>(survival factors)</i>	(3,4)
<i>jtrba035f05t1</i>	(3,4)
<i>jtrxa004c06t1</i>	(3,4)
<i>jtrba076h01t2</i>	(3,4)
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<i>jtrxa015d09t1</i>	(3,4)
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<i>jtrba032b05t3</i>	(3,4)
<i>jtrxa041d11t1</i>	(3,4)
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<i>jtrba098h06t1</i>	(3,4)
<i>jtrba019f07t1</i>	(3,4)
<i>jtrxa005h03t1</i>	(3,4)
<i>jtrxa003a01t1</i>	(3,4)
<i>jtrba120d07t1</i>	(3,4)
<i>jtrba011b04t1</i>	(3,4)
<i>jtrba115b01t1</i>	(3,4)
<i>jtrba009a07t2</i>	(3,4)

FIG. 7I.

SUBSTITUTE SHEET (RULE 26)

25/31

Tissue distribution of rat brain EST array elements.	
Category/Tissue	# Array elements detected
Total rat brain ESTs arrayed	7296
Any tissue	6127
Brain	5329
Heart	4525
Kidney	2686
Liver	1658
Lung	2620
Pancreas	1365
Skeletal muscle	1861
Smooth muscle	2714
Spleen	3921
Testes	3545
Brain not heart	1000
Brain only	582
All tissues	1169

FIG. 8.

26/31

Table 2. Smart Chip Genes regulated by KCI/Serum-withdrawal.

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
nexin 1	Late	glia-derived protease that is a neurite-promoting factor	P07092	jtrba019f07t1
SPARC	Late	Secreted Protein Acidic and Rich in Cysteine (regulates cell growth through interactions with the extracellular matrix and cytokines)	P16975	jtrba034c12t3
β -2-microglobulin	Late	precursor of major histocompatibility complex class I molecules (Fc receptor)	P07151	jlrxa006h05t1
GAD	Late	glutamic acid decarboxylase (catalyzes the production of inhibitory neurotransmitter GABA).	P18088	jtrba120d01t1
PAF A	Late	platelet activating factor acetylhydrolase (modulates action of PAF)	Q60963	jtrba028a03t1
intestinal membrane A4 protein	Late	possible differentiation role in intestinal epithelium (gene localizes near synaptophysin locus)	Q04941	jlrxa003a01t1
glutathione S-transferase Yb-1	Late	conjugation of reduced glutathione to exogenous and	M11719	jlrxa001f09t2
myoblast cell surface antigen	Late	myoblast cell surface antigen	X16850	jlrxa004e01t1
Na ⁺ -dep. glutamate/aspartate transporter	Late	essential for terminating the postsynaptic action of excitatory neurotransmission by rapid removal of glutamate from the synaptic cleft	S75687	jtrba014e07t3
ATP citrate-lyase	Late	primary enzyme for synthesis of cytosolic acetyl-coa and <i>de novo</i> lipid synthesis in neurons	P16638 P16638	jlrxa005e05t1 jlrxa005e05t1
vimentin	Late	vimentin cytoskeletal protein	X62952	jtrba108b01t1
phosphacan	Late	chondroitin sulfate proteoglycan of brain that interacts with neurons and neural cell-adhesion molecules and is an extracellular variant of receptor-type protein tyrosine phosphatase	U04998	jtrba011f03t1
NARC19	Late	epididymal secretory protein		jtrba078d11t1
L4BD NADP-dependent oxidoreductase	Late	catalyses the key step of metabolic inactivation of leukotriene B4 in tissues other than leukocytes	U66322	jlrxa015a05t1
S100 beta	Late	Ca ²⁺ binding protein		
GABA _A receptor α -1 subunit	Late	inhibitory neurotransmitter GABA receptor	M63436	jtrba011b04t1
OXY B	Late	oxysterol-binding protein that may play a role in regulation of sterol metabolism	P22059	jlrxa048f03t1
cystatin C	Late	cysteine proteinase inhibitor that regulates cysteine protease activity including cathepsin B, H, and L	P14841	jtrba005h01t1
P0	Late	60S acidic ribosomal protein	P19945	jtrba076a01t2
Na ⁺ /Cl ⁻ -dependent GABA transporter protein 1	Late	terminates the inhibitory action of GABA by its high affinity Na ⁺ -dependent reuptake into presynaptic terminals	P31648	jtrba078h11t1
Apo E	Late	apolipoprotein E transcription factor	J02582	jtrba119b10t1

FIG. 9A.

27/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
T-cell receptor	Late	novel protein with some homology to retroviral GAG and murine T-cell receptor γ .	M18247	jlrxa004c06t1
γ -like protein	Late	novel protein with some homology to gelsolin involved in Ca^{2+} -regulated, actin-modulation; promotes assembly of monomers into filaments, as well as sever filaments already formed; binds fibronectin	P20305	jlrxa019e05t2
gelsolin-like protein	Late	mitochondrial NAD-dependent bifunctional methylenetetrahydrofolate dehydrogenase/cyclohydrolase	P18155	jlrxa042c04t1
NAD ⁺ -dep. MTHFDH cyclohydrolase	Late	hydrolyzes an epoxide moiety of LTA-4 to form LTB-4; also has some peptidase activity; 3rd step in the biosynthesis of leukotrienes; belongs to zinc metallopeptidase family	M63848	jlrxa049a09t1
leukotriene A-4 hydrolase	Late	CB1 cannabinoid G protein coupled receptor	X55812	jlrba035f05t1
SKR6	Late	proteasome delta chain precursor for a multicatalytic proteinase complex involved in ATP/ubiquitin-dependent non-lysosomal proteolytic pathway	P28073	jlrxa030g11t1
PRCD	Late	novel with some homology to N- α -acetyl transferase (acetylation of proteins and peptides, used to stabilise pharmaceuticals or induce herbicide resistance in plants)	P12945	jlrxa030g11t1
N- α -acetyl transferase-like protein	Late	electrically silent basolateral cotransporter which mediates Na^+ / Cl^- reabsorption required for the regulation of ionic balance and cell volume	P55012	jlrba066f05t2
Na^+ - K^+ -2 Cl^- cotransporter	Late	novel protein with some homology to SNK.	P53351	jlrxa060b04t1
SNK-like protein	Late	serum/phorbol ester induced serine/threonine kinase	D38380	jlrba103b07t2
transferrin	Late	transports iron from sites of absorption and heme degradation to those of storage and utilization; may have a role in stimulating cell proliferation	X92864	jlrba118g01t1
testican	Late	testican	P38659	jlrxa054d03t1
CaBP2	Late	Ca^{2+} -binding protein with protein disulfide isomerase activity involved in rearrangement of intrachain & interchain disulfide bonds in proteins to form native structures in the ER	P70583	jlrxa001b10t2
PIP 4	Late	deoxyuridine 5'-triphosphate nucleotidohydrolase PPAR-interacting protein 4 (decreases intracellular dUTP such that uracil cannot be incorporated into DNA; binds and prevents PPAR dimerization with retinoid X receptor by inducing PPAR translocation to the nucleus	L25274	jlrba009d09t2
DM-GRASP	Late	lg superfamily axonal surface protein that supports neurite extension	X92864	jlrba120c06t1
testican-agrin-like protein	Late	novel protein with homology to testican and agrin	U09256	jlrxa044b08t1
Transketolase	Late	implicated in Wernicke-Korsakoff syndrome caused brain damage	P26641	jlrxa020c04t2
EF1G	Late	elongation factor 1-gamma that may anchor mRNA elongation complex to other cellular components	AB001926	jlrba018b01t1
APG-1	Late	Hsp70-related protein induced by hyperosmolar salt stress	P07825	jlrxa004f07t1
synaptophysin	Middle	synaptophysin synaptic vesicle protein		

FIG. 9B.

28/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
NADH dehydrogenase subunit 4	Middle	mitochondrial gene that shares regions of homology with NADH dehydrogenase subunit 4	U40063	jlrxa020h01t2
cytochrome c oxidase	Middle	mitochondrial cytochrome c oxidase	X14848	jtrba053b12t1
MU-crystallin homologue	Middle	NADP-regulated thyroid-hormone binding protein involved in the regulation of the free intracellular concentration of triiodothyronine and access to its nuclear receptors	Q14894	jtrba087g05t1
ubiquinone oxidoreductase e-like protein	Middle	novel protein with domain homology to scavenger-receptor protein NADH-ubiquinone oxidoreductase chain 4	P05508	jtrba126d05t1
zinc finger protein	Middle	zinc finger protein	L03386	jtrba065c11t1
NARC4	Middle	novel	W57419	jlrxa012g04t1
L-proline transporter	Middle	high affinity L-proline transporter	M88111	jtrba030a12t1
NARC5	Middle	novel	X53581	jtrba034g08t3
NARC7	Middle	novel		jlrxa027h05t1
NARC6	Middle	novel		jtrba118a12t1
caspase 3	Middle	PCD	P55213	jlrxc015b03t1
Zfhpf-2	Middle	zinc finger homeodomain enhancer-binding protein-2 that binds T3-response elements	U51583	jlrxa032a03t1
heparan sulfate proteoglycan core-like protein	Middle	novel protein with domain homology to basement membrane heparan sulfate proteoglycan core cell adhesion	M85289	jtrba107h07t1
RasGAP	Middle	RasGAP		rlrX019a05a1
4F2 bc	Middle	splice variant of type II membrane glycoprotein, a cell growth antigen required for expression of system L-like neutral amino acid-transport activity in C6-Bu-1 rat glioma	X89225	jtrba085b10t2
BCR 1	Middle	breakpoint cluster region protein (GTPase-activating protein for RAC1 and CDC42; chromosomal translocation produces BCR-ABL oncogene)	U07000	jtrba085b10t2
hippocalcin	Middle	neuron specific calcium-binding protein of the recoverin family expressed in hippocampus	D12573	jlrxa054b09t1
RT1.Aw3	Middle	MHC class 1 protein	L40363	jtrba019c12t1
Y-box binding protein	Middle	mammalian equivalents of the bacterial stress-inducible oxyR gene	L35599	jlrxa013c05t1
p130	Middle	cyclin-dependent kinase homologue that is a phosphotyrosine-independent SH2 ligand		jtrba108f08t1
PITSLRE	Middle	novel	U59235	jtrba119f08t1
NARC20	Middle	apoptosis regulated protein phosphatase		rlrX019b05a1
Wip-1	Middle	pyruvate dehydrogenase kinase 2 subunit	U10357	jtrba088c07t1
p45	Middle	serine/threonine-protein kinase in the CDC2/CDKX subfamily	Q00537	jtrba038e02t1
PCTAIRE-2	Middle	novel	AA182996	jtrba011c02t1
NARC14	Middle	novel protein with similarity to acetyltransferase		jtrba073f01t1
acetyltransferase-like protein	Middle			

FIG. 9C.

29/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
hTOM34-like protein	Middle	novel protein with domain homology to outer mitochondrial membrane 34 kDa translocase hTOM34	U58970	jtrba032b11t3
scavenger-receptor-like protein	Middle	novel protein with domain homology to scavenger-receptor protein	X99336	jtrba058c09t1
CAT-1	Middle	cationic amino acid transporter-1	U70476	jtrba043f05t1
carbonic anhydrase-like protein	Middle	novel protein with some homology to carbonic anhydrase	Y07785	jtrba002g03t1
NET-1-like protein	Middle	novel protein with some homology to guanine nucleotide regulatory protein NET-1 neuroepithelium oncogene	U02081	jlrxa043f01t1
stathmin	Middle	ubiquitous phosphoprotein which acts as an intracellular relay of proliferation and differentiation	Q09004	jtrba028g11t1
DIF-1-like protein	Middle	novel protein similar to C. elegans mitochondrial carrier protein DIF-1	Z75532	jtrba044d04t1
pentraxin	Early	neuronal pentraxin	U18772	jtrba049h01t2
NARC1	Early	novel		rlrX020c06b1
NARC2	Early	novel		rlrX022f07b1
apoptosis associated tyrosine kinase	Early	apoptosis associated tyrosine kinase	AF011908	jlrxa055h07t1
NARC30	Early	cell-death regulated secreted protein		jtrba057e05t1
Na ⁺ channel β -1 subunit	Early	Na ⁺ channel β -1 subunit, neurotransmission	Q00954	jtrba041g04t1
rhoB	Early	ras-like immediate early gene	M74295	jlrxa021g01t1
NARC13	Early	novel	D80004	jlrxa031h11t1
histone H2A	Early	histone H2A	X05862	jtrba124h04t1
PC3	Early	NGF-inducible anti-proliferative protein involved in cell cycle regulation, in particular, growth arrest and differentiation of neuronal precursor cells.	P27049	jlrxa027g01t1
histone H1	Early	histone H1	X72624	jtrba010c12t1
NARC9	Early	similar to C.elegans glycerophosphoryl diester phosphodiesterase	Q10003	jtra022a05t1
histone H3	Early	histone H3	M17876	jtrba019a12t2
HAP1-A-like protein	Late	similar to huntingtin-associated protein HAP1-A (except missing expanded polyglutamine repeat)	U38373	jtrba022f11t1
TAI	Late	a highly conserved oncofetal mRNA from rat hepatoma that encodes an integral membrane protein associated with liver development, carcinogenesis, and cell activation		jtrba026f06t3
DP5-like protein	Middle	similar to mRNA for polypeptide DP5 induced during programmed neuronal death	D83697	jtrba007a11t1
Nel	Middle	protein kinase C-binding protein Nel	U48245	jtrba016c05t1
CaMKII γ B	Imm. Early	CaMKII γ B involved in neuronal plasticity	U48245	jtrba016c05t1
dual specificity protein phosphatase 3	Imm. Early	dual specificity (tyrosine-, serine-) non-receptor protein phosphatase 3 involved in neuronal plasticity	P51452	jtrba073b11t1

FIG. 9D.

30/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
LDL receptor	Imm. Early	low-density lipoprotein receptor that binds LDL, the major cholesterol-carrying lipoprotein of plasma and transports it into cells by endocytosis after clustering in clathrin-coated pits	P35952	jlrxa012e10t1
NGF-In ZO-2	Imm. Early Imm. Early	NGF responsive transcription factor zonula occludens 2 protein, a tight junction protein homologous to the Drosophila discs-large tumor suppressor protein (contains guanylate kinase homology and three PDZ domains)		rlrX017b03b1
GluR5 vesicle-associated calmodulin-binding kinase-like protein	Imm. Early Imm. Early	excitatory neurotransmitter glutamate receptor vesicle-associated calmodulin-binding kinase-like protein	L22557	jtrba027g06t4
Neuro-endocrine secretory protein 55 NARC24	Imm. Early	neuroendocrine secretory protein 55 in the chromogranin family	U77614	jlrxa055g08t1
β -synuclein	Imm. Early	novel		jlrxa009d08t1
NARC21	Imm. Early	β -synuclein	S69965	jtrba040h07t1
14-3-3 protein	Imm. Early	novel	AA442186	jlrxa025e06t1
tau	Imm. Early	multifunctional regulator of CaMKII and PKC involved in neuronal plasticity	P35216	jtrba021f01t1
synapsin Ib	Imm. Early	synapsin Ib	M27924	jtrba041h10t1
NSG-1 P19 protein	Imm. Early	expressed in the Golgi apparatus of neural/neuroendocrine cells	P47759	jlrxa011c11t1
NARC12	Imm. Early	novel	H35593	jlrxa031b03t1
synaphin 2	Imm. Early	Protein associated with docking/fusion complex for synaptic vesicle release		jtrba035a02t1
Ca ²⁺ -dependent action-binding protein	Imm. Early	activator protein for secretion/exocytosis	U16802	jtrba046c05t1
ribonucleoprotein L	Imm. Early	heterogeneous nuclear ribonucleoprotein L	X16135	jtrba025h11t1
TCP-1	Imm. Early	chaperonin	P80317	jlrxa019a09t2
MCM2	Imm. Early	DNA replication licensing factor required for the entry into S phase and for cell division	P97310	jlrxa037f09t1
cytochrome b-560	Imm. Early	succinate-ubiquinone reductase	U31241	jtrba001a08t1
α -tubulin-1	Down-reg.	α -tubulin-1 cytoskeletal protein		jtrba012g02t1
vacuolar H ⁺ -ATPase	Down-reg.	active proton transport	M88690	jlrxa011g01t1
Ribonuclear particle U-like protein	Down-reg.	novel protein with some domain homology to heterogeneous ribonuclear particle protein U	D14048	jlrxa034f09t1
spindlin	Down-reg.	major maternal transcript expressed in the mouse during the transition from oocyte to embryo	U48972	jlrxa037b02t1
fodrin	Imm. Early	cytoskeletal component of Ca ²⁺ -secretion		jlrxa011g07t1

FIG. 9E.

SUBSTITUTE SHEET (RULE 26)

31/31

Gene ^a	Class ^b	Comment ^c	Accession ^d	SC EST ^e
MAP-1B	Imm. Early	microtubule associated cytoskeletal protein	A56577	jlrxa035f07t1
Synaptotagmin 1	Imm. Early	synaptic vesicle protein that binds to neurexins, syntaxin, and AP2 in Ca ²⁺ -independent fashion	P48018	jlrxa046c07t1
thyroid hormone-responsive gene	Imm. Early	gene from the Down syndrome critical region resembling a DNA binding protein with an SH3 domain	D83407	jtrba016g02t1
neurocalcin	Imm. Early	calcium-binding protein	D10884	jtrba035d02t1
NAD ⁺ -isocitrate dehydrogenase	Imm. Early	mitochondrial (NAD ⁺) isocitrate dehydrogenase	P50213	jlrxa060g03t1
NF-H	Imm. Early	heavy neurofilament cytoskeletal protein	M37227	jtrba073g11t1
K ⁺ channel β -subunit	Imm. Early	K ⁺ channel β -subunit, neurotransmission	L47665	jtrba022g06t1
NADH-ubiquinone oxidoreductase	Imm. Early	transfers electrons from NADH to ubiquinone in the respiratory chain	P34942	jlrxa026e02t1

(Table 2 footnotes)

^aThe gene identified as the most similar to the array element EST by BLAST. Genes are listed in the same order of hierarchical clustering as in Figure 4B.

^bGene expression class identified based on hierarchical clustering (Figure 4B). Following KCl/serum-withdrawal, Imm. Early, Early, Middle, and Late genes peaked at 1 hr., 3 or 6 hr., 12 hr., and 24 hr., respectively.

^cDescription of the gene identified by BLAST. The description was obtained from the Medline annotation for each sequence with indicated accession number. Red font highlights possible function physiologically relevant to programmed cell death or survival.

^dGenBank accession number.

^eSmart Chip EST identification number.

FIG. 9F.

SEQUENCE LISTING

<110> Chiang, Lillian Wei-Ming

<120> Nucleic Acid Molecules Derived from
Rat Brain and Programmed Cell Death Models

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ctg gag ctg gcc ttg aag ttg ccc cat gtc gac tac atc gag gag gac Leu Glu Leu Ala Leu Lys Leu Pro His Val Asp Tyr Ile Glu Glu Asp 135 140 145	670
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 630 635 640

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<213> Homo sapiens

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His Arg Cys Ala Lys Asp Pro Trp Arg Leu Pro Gly Thr Tyr Val Val
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Val Leu Lys Glu Glu Thr His Leu Ser Gln Ser Glu Arg Thr Ala Arg
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Arg Leu Gln Ala Gln Ala Ala Arg Arg Gly Tyr Leu Thr Lys Ile Leu
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 His Val Phe His Gly Leu Leu Pro Gly Phe Leu Val Lys Met Ser Gly
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 Pro Glu Glu Asp Gly Thr Arg Phe His Arg Gln Ala Ser Lys Cys Asp
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 Ser His Gly Thr His Leu Ala Gly Val Val Ser Gly Arg Asp Ala Gly
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 Val Ala Lys Gly Ala Ser Met Arg Ser Leu Arg Val Leu Asn Cys Gln
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 Lys Ser Gln Leu Val Gln Pro Val Gly Pro Leu Val Val Leu Leu Pro
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 Ile Gly Ala Ser Ser Asp Cys Ser Thr Cys Phe Val Ser Gln Ser Gly
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 Thr Ser Gln Ala Ala Ala His Val Ala Gly Ile Ala Ala Met Met Leu
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 Ser Ala Glu Pro Glu Leu Thr Leu Ala Glu Leu Arg Gln Arg Leu Ile
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 His Phe Ser Ala Lys Asp Val Ile Asn Glu Ala Trp Phe Pro Glu Asp
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 Gly Cys Ser Ser His Trp Glu Val Glu Asp Leu Gly Thr His Lys Pro
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 Pro Val Leu Arg Pro Arg Gly Gln Pro Asn Gln Cys Val Gly His Arg
 580 585 590
 Glu Ala Ser Ile His Ala Ser Cys Cys His Ala Pro Gly Leu Glu Cys
 595 600 605
 Lys Val Lys Glu His Gly Ile Pro Ala Pro Gln Glu Gln Val Thr Val
 610 615 620
 Ala Cys Glu Glu Gly Trp Thr Leu Thr Gly Cys Ser Ala Leu Pro Gly

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Arg Ser Arg Asp	Val Ser Thr Thr Gly Ser Thr Ser Glu Glu Ala Val					
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 Met Gly Thr Val Ser Ser
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 Arg Arg Ser Trp Trp Pro Leu Pro Leu Leu Leu Leu Leu Leu Leu
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 Leu Gly Pro Ala Gly Ala Arg Ala Gln Glu Asp Glu Asp Gly Asp Tyr
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 Glu Glu Leu Val Leu Ala Leu Arg Ser Glu Glu Asp Gly Leu Ala Glu
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 Ala Pro Glu His Gly Thr Thr Ala Thr Phe His Arg Cys Ala Lys Asp
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 Pro Trp Arg Leu Pro Gly Thr Tyr Val Val Val Leu Lys Glu Glu Thr
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 Ala Arg Arg Gly Tyr Leu Thr Lys Ile Leu His Val Phe His Gly Leu
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 Leu Lys Leu Pro His Val Asp Tyr Ile Glu Glu Asp Ser Ser Val Phe
 135 140 145 150

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cgc gtc ctc aac gcc gcc tgc cag cgc ctg gcg agg gtt ggg gtc gtg Arg Val Leu Asn Ala Ala Cys Gln Arg Leu Ala Arg Val Gly Val Val 295 300 305 310			1026
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 Glu Asp Gly Leu Ala Glu Ala Pro Glu His Gly Thr Thr Ala Thr Phe
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 His Arg Cys Ala Lys Asp Pro Trp Arg Leu Pro Gly Thr Tyr Val Val
 65 70 75 80
 Val Leu Lys Glu Glu Thr His Leu Ser Gln Ser Glu Arg Thr Ala Arg
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 His Val Phe His Gly Leu Leu Pro Gly Phe Leu Val Lys Met Ser Gly
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 Ile Thr Pro Pro Arg Tyr Arg Ala Asp Glu Tyr Gln Pro Pro Asp Gly
 165 170 175
 Gly Ser Leu Val Glu Val Tyr Leu Leu Asp Thr Ser Ile Gln Ser Asp
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 His Arg Glu Ile Glu Gly Arg Val Met Val Thr Asp Phe Glu Asn Val
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 Pro Glu Glu Asp Gly Thr Arg Phe His Arg Gln Ala Ser Lys Cys Asp
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 Ser His Gly Thr His Leu Ala Gly Val Val Ser Gly Arg Asp Ala Gly
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 Val Ala Lys Gly Ala Ser Met Arg Ser Leu Arg Val Leu Asn Cys Gln
 245 250 255
 Gly Lys Gly Thr Val Ser Gly Thr Leu Ile Gly Leu Glu Phe Ile Arg
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 Lys Ser Gln Leu Val Gln Pro Val Gly Pro Leu Val Val Leu Leu Pro
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 Gly Ala Thr Asn Ala Gln Asp Gln Pro Val Thr Leu Gly Thr Leu Gly
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 370 375 380
 Thr Ser Gln Ala Ala Ala His Val Ala Gly Ile Ala Ala Met Met Leu
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 420 425 430
 Gln Arg Val Leu Thr Pro Asn Leu Val Ala Ala Leu Pro Pro Ser Thr
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 Ser Gly Pro Thr Arg Met Ala Thr Ala Ile Ala Arg Cys Ala Pro Asp

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Asn Ala Phe Gly Gly Glu Gly Val Tyr Ala Ile Ala Arg Cys Cys Leu
          515          520          525
Leu Pro Gln Ala Asn Cys Ser Val His Thr Ala Pro Pro Ala Glu Ala
          530          535          540
Ser Met Gly Thr Arg Val His Cys His Gln Gln Gly His Val Leu Thr
545          550          555          560
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                                     Met Val
                                     1

ctg tcg gtg cct gtg atc gcg ctg ggc gcc acg ctg ggc aca gcc acc      166
Leu Ser Val Pro Val Ile Ala Leu Gly Ala Thr Leu Gly Thr Ala Thr
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agc atc ctc gcg ttg tgc ggg gtc acc tgc ctg tgt cgg cac atg cac      214
Ser Ile Leu Ala Leu Cys Gly Val Thr Cys Leu Cys Arg His Met His
          20          25          30

ccc aag aag ggg ctg ctg ccg cgg gac cag gac ccc gac ctg gag aag      262
Pro Lys Lys Gly Leu Leu Pro Arg Asp Gln Asp Pro Asp Leu Glu Lys
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gcg aag ccc agc ttg ctc ggg tct gca caa cag ttc aat gtt aaa aag      310
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tcc acg gaa cct gtt cag ccc cgt gcc ctc ctc aag ttc cca gac atc      358
Ser Thr Glu Pro Val Gln Pro Arg Ala Leu Leu Lys Phe Pro Asp Ile
          70          75          80

tat gga ccc agg cca gct gtg acg gct cca gag gtc atc aac tat gca      406
Tyr Gly Pro Arg Pro Ala Val Thr Ala Pro Glu Val Ile Asn Tyr Ala
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gac tat tca ctg agg tct acg gag gag ccc act gca cct gcc agc ccc      454
Asp Tyr Ser Leu Arg Ser Thr Glu Glu Pro Thr Ala Pro Ala Ser Pro
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caa ccc ccg aat gac agt cgc ctc aag agg cag gtc aca gag gag ctg      502
Gln Pro Pro Asn Asp Ser Arg Leu Lys Arg Gln Val Thr Glu Glu Leu
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Phe Ile Leu Pro Gln Asn Gly Val Val Glu Asp Val Cys Val Met Glu
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cac tac tgc ctg gac tat gac tgt cag aag gca gaa ttg ttt gtg act 646
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cgc ctg gaa gct gtg acc agc aac cac gac gga ggc tgt gac tgc tac 694
Arg Leu Glu Ala Val Thr Ser Asn His Asp Gly Gly Cys Asp Cys Tyr
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Val Gln Gly Ser Val Ala Asn Arg Thr Gly Ser Val Glu Ala Gln Thr
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gcc cta aag aag cgg cag ctg cac acc acc tgg gag gag ggc ctg gtg 790
Ala Leu Lys Lys Arg Gln Leu His Thr Thr Trp Glu Glu Gly Leu Val
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ctc ccc ctg gcg gag gag gag ctc ccc aca gcc acc ctg acg ctg acc 838
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ttg agg acc tgc gac cgc ttc tcc cgt cac agc gtg gcc ggg gag ctc 886
Leu Arg Thr Cys Asp Arg Phe Ser Arg His Ser Val Ala Gly Glu Leu
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cgc ctg ggc ctg gac ggg aca tct gtg cct cta ggg gct gcc cag tgg 934
Arg Leu Gly Leu Asp Gly Thr Ser Val Pro Leu Gly Ala Ala Gln Trp
260 265 270

ggc gag ctg aag act tca gcg aag gag cca tct gca gga gct gga gag 982
Gly Glu Leu Lys Thr Ser Ala Lys Glu Pro Ser Ala Gly Ala Gly Glu
275 280 285 290

gtc cta cta tcc atc agc tac ctc ccg gct gcc aac cgc ctc ctg gtg 1030
Val Leu Leu Ser Ile Ser Tyr Leu Pro Ala Ala Asn Arg Leu Leu Val
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Leu Gly Lys Asp Val Ser Val Lys Val Thr Leu Lys His Gln Ala Arg
325 330 335

aag ctg aag aag aag cag act aaa cga gct aag cac aag atc aac ccc 1174
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INTERNATIONAL SEARCH REPORT

International Application No

PC1/US 00/29132

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/12 C07K14/47 C07K16/18 C12Q1/68 G01N33/53
A61K38/17

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C07K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EMBL, EPO-Internal, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE EMBL [Online] 17 August 1999 (1999-08-17) BLAKEY S.: "KIAA1434" retrieved from EBI Database accession no. AL109935 XP002163761	1,4-17, 26
Y	abstract	2,3
X	DATABASE EMBL [Online] 19 March 1999 (1999-03-19) POUSTKA A. ET AL.: " EST DKFZp434B2072_r1" retrieved from EBI Database accession no. AL046038 XP002163762	1,4-17, 26
Y	abstract	2,3

	-/--	

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

23 March 2001

Date of mailing of the international search report

05 June 2001

Name and mailing address of the ISA

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Authorized officer

Gurdjian, D

INTERNATIONAL SEARCH REPORT

International Application No

PC., US 00/29132

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 98 49297 A (AMERICAN HOME PROD) 5 November 1998 (1998-11-05) abstract; claims 1-7 ---	1-17,26, 28
A	AOKI TOMOKAZU ET AL: "Rat TAFII-31 gene is induced upon programmed cell death in differentiated PC12 cells deprived of NGF." BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 234, no. 1, 1997, pages 230-234, XP002163760 ISSN: 0006-291X abstract; figures 4,5 ---	1-17,26, 28
A	RICKMAN DENNIS W ET AL: "Characterization of the cell death promoter, Bad, in the developing rat retina and forebrain." DEVELOPMENTAL BRAIN RESEARCH, vol. 115, no. 1, 8 June 1999 (1999-06-08), pages 41-47, XP000990537 ISSN: 0165-3806 abstract ---	1-17,26, 28
A	HEMACHANDRA REDDY P ET AL: "Transgenic mice expressing mutated full-length HD cDNA: A paradigm for locomotor changes and selective neuronal loss in Huntington's disease." PHILOSOPHICAL TRANSACTIONS OF THE ROYAL SOCIETY OF LONDON B BIOLOGICAL, vol. 354, no. 1386, June 1999 (1999-06), pages 1035-1045, XP000990516 June, 1999 ISSN: 0962-8436 abstract -----	1-17,26, 28

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 00/29132

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 22-25 and claims 18-21, as far as they concern an in vivo method, are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☒ Claims Nos.: 27
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims: it is covered by claims Nos.:
see subject 1. on extra sheet

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 27

Claim 27 relating to binding agents to the polypeptide of claim 10 could not be searched as its subject-matter was insufficiently disclosed .

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.1 or 4 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

2. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.2 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

3. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.3 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

4. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.5 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

5. Claims: 1-28 partly

nucleic acids with nucleotide sequence with seq.id.6 or 8 and corresponding polypeptides , vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe.

6. Claims: 1-28 partly

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

nucleic acids with nucleotide sequence with seq.id.10 and corresponding polypeptides, vectors ,host cell , method of preparing a polypeptide , antibody , method of assaying the presence of anucleic acid ,method of detecting a polypeptide ,method of modulating the activity of a polypeptide and kit comprising a nucleic acid probe .

INTERNATIONAL SEARCH REPORT

formation on patent family members

International Application No

PC., US 00/29132

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9849297 A	05-11-1998	AU 6969998 A EP 0977846 A	24-11-1998 09-02-2000

Form PCT/ISA/210 (patent family annex) (July 1992)